Table S1. Sequences for PCR primers to generate templates for RNA probe synthesis					
Gene name		Primer sequence	Product size		
Slc34a1, probe 1	F	5'-CTTCTTCAACATCTCGGGCATC-3'			
	R	5'-TCTGTCTTTCTACTGTGGGCATTG-3'	556 bp		
Slc34a1, probe 2	F 5'-CGCTGGTGTTTGGCATTTCC-3'				
	R	5'-GCACTAATGGTCACACAGGCTCAG-3'	580 bp		
Slc12a3, probe 1	F	5'-CCTTTGATACCCAGAGCCATAATG-3'			
	R	5'-AATGAATGCAGGTCAGCCAGG-3'	290 bp		
Slc12a3, probe 2	3, probe 2 F 5'-GTGGCACCTATTTCCTTATTTCCC-3				
	R	5'-CCCTTACGGTTTCTGCAAAGC-3'	129 bp		
Wnt4, probe 1	F	5'-GGAGACGTGCGAGAAACTCAAAG-3'			
	R	5'-TGTGTCACCACCTTCCCAAAGAC-3'	192 bp		
Wnt4, probe 2	F	5'-CGCTAAAGGAGAAGTTTGACGGTG-3'			
	R	5'-GGTCCTCATCTGTATGTGGCTTG-3'	111 bp		
Podxl, probe 1	F	5'-GCCAAGCAACCCTACACCATTC-3'			

T7 sequence 5'-ATTGTAATACGACTCACTATAGGG-3'

To generate probes for the genes listed, we amplified two different PCR products specific for exon sequences of each gene using mouse genomic DNA as a template. The reverse primers had a 5' extension that included the T7 promoter for in vitro transcription. The two resulting probes for each gene were combined in the hybridization solution.

5'-TCGCTGTGCTCGGTGAAGAATC-3'

5'-AATGGTCTGTGATGGTCACGGG-3'

5'-GCCTCATTCTGCTGGACATTCC-3'

359 bp

394 bp

F, forward primer; R, reverse primer. The sequence of the T7 promoter is highlighted.

R

R

Podxl, probe 2

Table S2. Total number of collecting duct tips in kidneys of normal and mutant littermates
Number of collecting dust time

and material material				
Number of collecting duct tips				
E13.5	E15.5	E17.5		
89.6±25.5	337.4±39.2	874±61.9		
40.3±10.1 (45%)	90.5±25.1 (27%)	148±25.3 (20%)		
79.5±22.7	397.2±33.9	1098±147		
60.1±15.3 (76%)	284.3±55 (72%)	518±61 (47%)		
	Nun E13.5 89.6±25.5 40.3±10.1 (45%) 79.5±22.7	Number of collecting duct E13.5 E15.5 89.6±25.5 337.4±39.2 40.3±10.1 (45%) 90.5±25.1 (27%) 79.5±22.7 397.2±33.9		

Total number of collecting duct tips in kidneys of normal and mutant littermates was quantified as previously described (Cebrián et al., 2004). Briefly, $100 \, \mu m$ vibratomesectioned kidneys were permeabilized and assayed for immunofluorescent staining using primary antibodies against Calbindin $D_{28K}(CD)$. CD-positive duct tips were visualized under fluorescent microscopy and quantified in all sections of a given kidney. A minimum of seven kidneys at each developmental stage was analyzed with values expressed as mean $\pm s.d.$

Reference

Cebrián, C., Borodo, K., Charles, N. and Herzlinger, D. A. (2004). Morphometric index of the developing murine kidney. *Dev. Dyn.* 231, 601-608.

mutant littermates					
	Number of renal corpuscles				
	E15.5	E18.5			
Normal littermates	379±28	2519±374			
Mild Fgf8 hypomorph	265±32 (70%)	912.5±83.3 (36%)			
Normal littermates	427±34	2812±409			
Severe <i>Fgf</i> 8 hypomorph	201±37 (47%)	531±76.5 (19%)			

Table 3. Total number of renal corpuscles in kidneys of normal and

Samples were analyzed as described in Table S2, using primary antibodies against Wilms' tumor 1 protein (WT1). Overprojection bias was corrected as previously described by Cebrián et al. (Cebrián et al., 2004). A minimum of five kidneys at each developmental stage was analyzed with values expressed as mean±s.d.

Reference

Cebrián, C., Borodo, K., Charles, N. and Herzlinger, D. A. (2004). Morphometric index of the developing murine kidney. Dev. Dyn. 231, 601-608.