Table S1. Markers and primers generated for mapping the mutation in the line JF1781

Marker	Primer sequence (5′ to 3′)	Polymorphism	Enzyme	Amplicon (bp)	
				Col	C24
F12K11 14/15	AAGAAGAGCCTGTCACCAACTAC TTGACTCTAACCTAATCGCAGAT	In/del	–	300	250
F7G19 13/14	TTCATCGGTCGGCACAAAAGTGT CAAACTCGCTTATTCAACAAGAGAC	In/del	–	350	300
F10K1.15	TGCTGATGGTGAACTCAAGG CAGTCGACCATGTTGGCATA	In/del	–	350	400
F22G5 8/9	TCTTAGTCGGGTCAACGAAAA TTTTTGTGTGGTGAGACCAATC	In/del	–	325	275
T23G18 4/5	AAGAGGAAAATGAAGGCAACAG TCCCTAACTACTGGTTTTCACC	In/del	–	275	300
T27G7.11	TCAGGCCCATATTCGTTTGG AGGCTTTCAGTTTGACCCCA	In/del	–	250	275
F24B9 13/14	GCAGTATAGGCTGATGTGTGTGTT CCAAACCGAATCCGACTCTA	In/del	–	300	320
T6D22 24/25	GCAGAAATATGTTCCCCTGCT GTCGATGAACATGTTTGGAAGA	CAPS	*Bst*NI	–	+
T6D22 8/9	TCAAGAAGTTGGAGTGTTGGAA AAGGTAAAGCAGGGGAGGAA	CAPS	*Nsi*I	–	+
T6D22.2	ATCGTTTGCGTGAGGTTAGC ATGCTGCTCCAGAATTACCC	CAPS	*Mwo*I	–	+
T6D22 12/13	GGTGTTTTTAGTTAGGCTGAATTTGGA- TCTTATTGTATAATCTTATTCAGCT GGTTCTGTGATTGCTGGTT	CAPS	*Pvu*II	+	–

The markers have been named according to the BAC and the ORF in which they were identified. In/del indicates an insertion/deletion has taken place, and the size of PCR fragments obtained for each ecotype is specified. CAPS, cleavage amplified polymorphic sequence. Plus and minus signs indicate from which ecotype the corresponding PCR fragment will be cut or not, respectively. Underlined bases in the T6D22 12/13 forward primer were introduced to create a CAPS between the Col and the C24 sequence.