

Table S2. Plasmids**A Plasmids used for data presented in main text**

Plasmid	Description	Reference/Source/Construction
pGC464	<i>ins-33</i> rescue and overexpression	This work: ~8kb genomic region of <i>ins-33</i> was PCR amplified using the following primers: AAGGAGAACAACTGTATCGAGATTGAGGG and CATCGTCTGGAACAAATGAAGAACGAATGGCGG and TA cloned into pCR-XL-TOPO (Invitrogen)
pGC467	<i>ins-3</i> rescue and overexpression	This work: ~11.5kb genomic region of <i>ins-3</i> was PCR amplified using the following primers: ATGAAGCGGAGAGAAGAAGTCGGAGAGAAGG and GTTATGGACATATCGTACTAAGTCTGCTGCC and TA- cloned into pCR-XL-TOPO (Invitrogen)
sjj_C46A5.9	<i>hcf-1</i>	(Kamath et al., 2003) ^a
sjj_T07A9.6	<i>daf-18</i>	(Kamath et al., 2003) ^a
sjj_R13H8.2 ^b	<i>daf-16</i>	(Kamath et al., 2003) ^a
sjj_R13H8.1 ^b	<i>daf-16</i>	(Kamath et al., 2003) ^a
mv_CAA10315	<i>daf-18</i>	(Rual et al., 2004; C.elegans ORF-RNAi library (Geneservice Ltd.)) ^a
	RNAi-targeting: <i>ins-1</i> , <i>ins-2</i> , <i>ins-3</i>	Kindly provided by Monica Driscoll ^a
pGC488	<i>daf-2</i>	This work: created by ligating the <i>Kpn</i> I/ <i>Xba</i> I fragment from pKDK33 (Wolkow et al., 2000) to similarly-digested L4440
pGC461	Distal tip cell expression of DAF-16	This work: <i>daf-16::GFP</i> fusion (gift of T. Johnson) was fused to the <i>lag-2</i> promoter in pJK590 (Blelloch et al., 1999; Mathies et al., 2003).
pGC492	Germline expression of DAF-16	This work: <i>daf-16::GFP</i> fusion (gift of T. Johnson) was fused to nos-2 3'UTR. The <i>rpl-11.1</i> promoter (5'-cgcgttcaatccccgggtccggccctttt.....tcacagttcaaatttatgtatgc-3') and then cloned along with the <i>C. briggsae unc-119(+)</i> gene into pPD117.01 (kind gift of Barth Grant).

B Plasmids used for data presented in supplementary material

Plasmid	Description	Reference/Source/Construction
pGC487	<i>ins-3</i> expression	This work: A 5565 bp 5' fragment of <i>ins-3</i> was PCR amplified from pGC467 using primers: CAAGCTAGCTAAGTAAGTTGTATTGTTACAAACG and CAAGGGCCCGTGTAGTCGACTTCAGATCATG, digested with <i>Nhe</i> I and <i>Apal</i> , and ligated to similarly digested pGC305 (Voutev and Hubbard, 2008), to create pGC485. Next, a 5749 bp fragment covering the first intron to the 3' downstream region, was PCR amplified from pGC467 using primers: AACCTGCAGGGGTTGTCGACATGAAGCGGAGAG and CAACCCGGGTATTCAGAACAGGAATTGATAAATGTGTC, digested with <i>Sbf</i> I and <i>Xma</i> I, and ligated to similarly digested pGC485, to create pGC487
pGC486	<i>ins-33</i> expression	This work: A 1287 bp 5' fragment of <i>ins-33</i> was PCR amplified from pGC464 using the following primers: CAAGGATCCAAGGAGAACAACTGTATCGAG and CAAGAGCTTTGTTCAAAAAATCAGCAC, digested with <i>Bam</i> H I and <i>Sac</i> I, and ligated to similarly digested pGC305 (Voutev and Hubbard, 2008), to create pGC482. Next, a 5800 bp fragment covering the first intron to the 3' downstream region, was PCR amplified from pGC464 using the following primers: CAAGCTAGCTAAGTAAGCGATGAAAATCGATAGAACAC and CAAGGGCCCCATCGTCTGGAACAAATGAAGAACG, digested with <i>Nhe</i> I and <i>Apal</i> , and ligated to similarly digested pGC482, to create pGC486
	RNAi-targeting: <i>ins-4</i> , <i>ins-5 ins-6</i> , <i>ins-8</i> , <i>ins-9</i> , <i>ins-10</i> , <i>ins-11 ins-12</i> , <i>ins-13</i> , <i>ins-14</i> , <i>ins-15</i> , <i>ins-16</i> , <i>ins-17</i> , <i>ins-18</i> , <i>ins-19</i> , <i>ins-20</i> , <i>ins-21</i> , <i>ins-22</i> , <i>ins-23</i> , <i>ins-26</i> , <i>ins-27</i> , <i>ins-28</i> , <i>ins-29</i> , <i>ins-30</i> , <i>ins-32</i> , <i>ins-33</i> , <i>ins-34</i> , <i>ins-35</i> , <i>ins-36</i> , <i>ins-37</i>	Kindly provided by Monica Driscoll ^a
sjj_F21E9.4	<i>ins-39</i>	(Kamath et al., 2003) ^a
pGC314	<i>ins-7</i>	This work ^c : ATATTCTAGAACATGCCACCAATAATTTGG and ATATCTCGAGTTAAGGACAGCACTGTTTC
pGC315	<i>ins-24</i>	This work ^c : ATATTCTAGAACATGAGATCTCCACCTTG and ATATCTCGAGTTAGAAAACGAAGCCAGATG
pGC316	<i>ins-31</i>	This work ^c : ATATTCTAGAACATGAGATGCCCTTGATC and ATATCTCGAGCTAGAAAAGCTGGACG
pGC317	<i>ins-38</i>	This work ^c : ATATTCTAGAACATGAGCTTCTCCTCG and ATATCTCGAGCTAGCTTGTGGGGC
pGC318	<i>daf-28</i>	This work ^c : ATATTCTAGAACATGCAAGCTCATCG and ATATCTCGAGGTGGTACAGGCGTCTC

^a Verified by DNA sequencing.^b These two reagents gave similar results.^c In each case, plasmids were made by PCR amplification from a cDNA library (Invitrogen) using indicated primers, digested with *Xba*I/*Xho*I, and ligated into similarly-digested L4440.