Table S2. Primer sequences for quantitative and semi-quantitative PCR and cloning of *ATHB15::LhG4* and *10OpTATA::ATHB8-δmiR*

cioning of ATHBTS::LING4 and TUOPTATA::ATHB8-omik	
Primer	Sequence (5' to 3') or source
S16	QT00833819 (Qiagen)
KAN1	QT00830802 (Qiagen)
KAN2	GCAGCTTCGTCAGGACAATC and
	TCTCCGGAAGAATTGGTCCA
REV	CGAATAGTCCTGCTGGATTG and
	GATCTCTGCAATCTTCATAG
PHB	QT00716583 (Qiagen)
PHV	QT00870877 (Qiagen)
AtHB15	AGTCCTGCAGGACTTTTGTC and
	CAATCTCTGCAACCCTTGTA
AtHB8	AAGCAGAGGAAACTCAAG and
	AAATGTGAATACCGGAGAT
PIN1	QT00892654 (Qiagen)
APL	QT00744926 (Qiagen)
MP	QT00861847 (Qiagen)
Cyclophilin	TCTTCCTCTTCGGAGCCATA and
	AAGCTGGGAATGATTCGATG
AS2	ATGGCATCTTCTTCAACAAAC and
	TCAAGACGGATCAACAGTAC
PIN1	ATGATTACGGCGGCGGACTTC and
	TCATAGACCCAAGAGAATGTAGTAGAG
PIN3	CAAGATGATCTCATGGCACG and
	GCTTATAACCCGAGTAGAATG
KAN1-GR	GGAGAAGAAATGGGC and
	CTTTGCCCATTTCACTGC
PIN4	GGAAAAATGATTACGTGGC and
	GGGACCAATTCAAAGGCC
AtHB15 promoter	GGCGCGCCATAGAGAGCAAATGTGATGGTG and
	GGGGTACCTACTCCTCAGCAAAACTCTT
ATHB8-δmiR	ACCTCGAGATGGGAGGAGGAAGCAATA and

Either the QuantiTect Primer Assay (Qiagen) or oligonucleotide primers (Microsynth, Balgach, Sg, Switzerland) were used at a concentration of 0.5 μ M. Real-time quantitative RT-PCR was performed with an iCycler iQ Real-Time PCR Detection System (BioRad, Hercules, CA, USA), using Absolute SYBR Green Fluorescein, which binds non-specifically to double-stranded DNA (ABgene, Epsom, Surrey, UK). The ribosomal S16 gene was used as a standard control. Reactions were performed in triplicate in 20 μ l using 100 ng cDNA and 0.4 mM MgCl $_2$ per reaction. Negative controls were performed in duplicate. Thermal cycling parameters were 95°C for 15 minutes, followed by 45 cycles at 95°C for 30 seconds, 59.7°C for 30 seconds and 72°C for 45 seconds. RT-PCR specificity was tested with melting-curve analysis and electrophoresis. Experiments were repeated three times. All calculations were performed with the Gene Expression Macro 1.1 (Bio-Rad) macros for Excel (Microsoft, Redmond, WA, USA). Semi-quantitative real-time PCR on 1 μ l cDNA was performed using ExTaq (TaKaRa Biotech).

GCGGTACCCATATAAAAGACCAGTTGA