Table S1. Primers

Number		
	ChIP primers – *Fgf10* promoter	**Amplicon position**
1	For: 5′-acctccttggaagctggcagg-3′ Rev: 5′-atgtgtttctgccttccagatcc-3′	–804 to –625
2	For: 5′-ttcatttgcgccaagagagagg-3′ Rev: 5′-ccagcttccaaggaggtctcg-3′	–942 to –788
3	For: 5′-ggctttctggggctaagctaagagg-3′ Rev: 5′-ccaaaggaaaagctgacatagctttttag-3′	–2765 to –2644
4	For: 5′-gaagtaactgagaatgattcagg-3′ Rev: 5′-agcctgagaaaagttccacagg-3′	–2899 to –2761
5	For: 5′-gggtaccattctaatctgaagc-3′ Rev: 5′-gactgaaattctacctacctaccttcc-3′	–3032 to –2889
	Cloning primers – WT *Fgf10* promoter	**Cloning site**
	For: 5′-atctg<u>gaagctc</u>attctgccctttcaatcttaacag-3′ Rev: 5′-ctgct<u>aagctt</u>tcttgggcgagaggagtggc-3′	*Sac*I *Hin*dIII
	Cloning strategy – del (HD6-9) *Fgf10* promoter	
	Wild-type promoter was cut at *Sac*I and *Stu*I restriction sites, which flank homeodomain-binding sites sites 6-9. The cut construct was end filled by T4 DNA polymerase for blunt-end cloning, resulting in a construct that contains a small 131 bp deletion and lacks HD sites 6-9.	*Sac*I *Stu*I T4 DNA polymerase filled in and blunt end ligated