

Fig. S1. Upd distribution is graded prior to specification of follicular epithelium. ( $\mathbf{A}, \mathbf{A}^{\prime}$ ) Fluorescence signal intensity of Upd protein (red) as detected by anti-Upd antisera processed for extracellular protein was quantified along the apical surface of the follicle cells at the anterior and posterior poles. DNA is blue. ( $\mathbf{B}, \mathbf{B}^{\prime}$ ) Signal intensity is plotted relative to position along the epithelium. Colored arrows on the image and graphs mark corresponding positions. Scale bar: $20 \mu \mathrm{~m}$.


Fig. S2. sdc and trol do not influence the distribution of Upd during oogenesis. (A-D) Follicular clones of $s d c$ (A,B) and trol (C,D) mutant cells are outlined in white as marked by lack of GFP (green). Upd distribution (A,D, red) and pnt-lacZ expression which serves as a reporter of JAK/STAT activity at the posterior (C, red) are shown. Upd distribution appears to be normal in chambers with mosaic follicle cells of either mutation. Anterior follicle cells mutant for $s d c$ (B, loss of GFP) undergo normal morphological transitions for their region, including extension of stretched cells (arrowheads) and migration of the adjacent cells (outline) and cannot be differentiated from the wild-type anterior follicle cells (B, GFP). In large trol clones (C, loss of GFP), graded expression of pnt-lac $Z$ is unaffected.


Fig. S3. Loss of Hs2st does not alter upd expression. (A,B) RNA in situ hybridization to upd shows comparable expression between wild-type (A) and Hs2st mutant (B) ovaries. Scale bar: $20 \mu \mathrm{~m}$.


Fig. S4. dally misexpression does not alter upd expression. Flip-out misexpression clones of dally (marked with GFP, green) in the background of an enhancer trap in the upd gene (PD) show expression of the $\beta$-galactosidase reporter (red) only in the polar cells (asterisks), but not in dally-misexpressing cells.

Table S1. Primers used to amplify templates from cDNA for in situ hybridization

| Target gene <br> dally | Primer pairs <br> 5'-GTAATACGACTCACTATAGGGCCAGCTTTTGCTATTTACCCTGC-3' |
| :--- | :--- |
|  | $5^{\prime}$--AATTAACCCTCACTAAAGGGAGTCCGGCATATTCCGCCG-3' |

Primer sets used to synthesize template for RNA labeling are indicated for each gene of interest.
Underlines indicate T7 and T3 RNA polymerase binding sites, respectively.

Table S2. Stalk size is reduced in HSPG mutant ovarioles

|  | Number of cells in stalk |  |  |
| :--- | :---: | :---: | :---: |
|  | $1-2$ | $3-4$ | $\geq 5$ |
| Control | 0 | 0 | 39 |
| dally $^{\text {gem }}$ dally $^{527}$ | 9 | 16 | $15^{*}$ |
| dlp $^{A 187}$ clone in stalk cells | 0 | 0 | 3 |
| $d^{A 187}$ clone in polar cells | 0 | 0 | 20 |
| dally $^{80}$, dlp $^{A 187}$ clone in stalk cells | 9 | 1 | $0^{*}$ |
| dally $^{80}, d l p^{A 187}$ clone in polar cells | 0 | 0 | 14 |

* $\mathrm{P}<0.05$ by Fisher's exact probability test.

The number of stalk cells was counted for stage 2-7 egg chambers of the indicated dally mutant combination or for chambers with the indicated locations of mutant clones. For polar cell clones, cells were counted for the adjacent stalk. For stalk cell clones, only clones in which the entire stalk was mutant were scored. Stalk cell numbers that differ statistically from the wild-type control are marked with an asterisk.

Table S3. pSTAT92E in Hs2st mutant ovaries

| Genotype | pSTAT92E | pSTAT92E positive stage 7-9 egg chambers |  |
| :--- | :---: | :---: | :---: |
|  | positive germaria | Anterior follicle cells | Posterior follicle cells |
| Control | $84 \%(n=25)$ | $90.9 \%(n=33)$ | $78.8 \%(n=33)$ |
| $H s 2 s t$ | $2.9 \%(n=34)^{*}$ | $12.5 \%(n=32)^{*}$ | $21.9 \%(n=32)^{*}$ |

*P<0.05 $\chi^{2}$ test
Anti-pSTAT92E fluorescence signal was observed in ovaries of wild-type and Hs2st mutants.
Using the non-specific ring canal staining as an internal control, signal was scored as background (negative) or above background (positive). In the posterior germarium, pSTAT92E was scored in the follicle cell precursors. In stage 7-9 egg chambers of the vitellarium, anterior and posterior follicle cells were scored separately for pSTAT92E. Statistically significant differences between wild-type and $H s 2 s t$ ovaries are marked with an asterisk.

