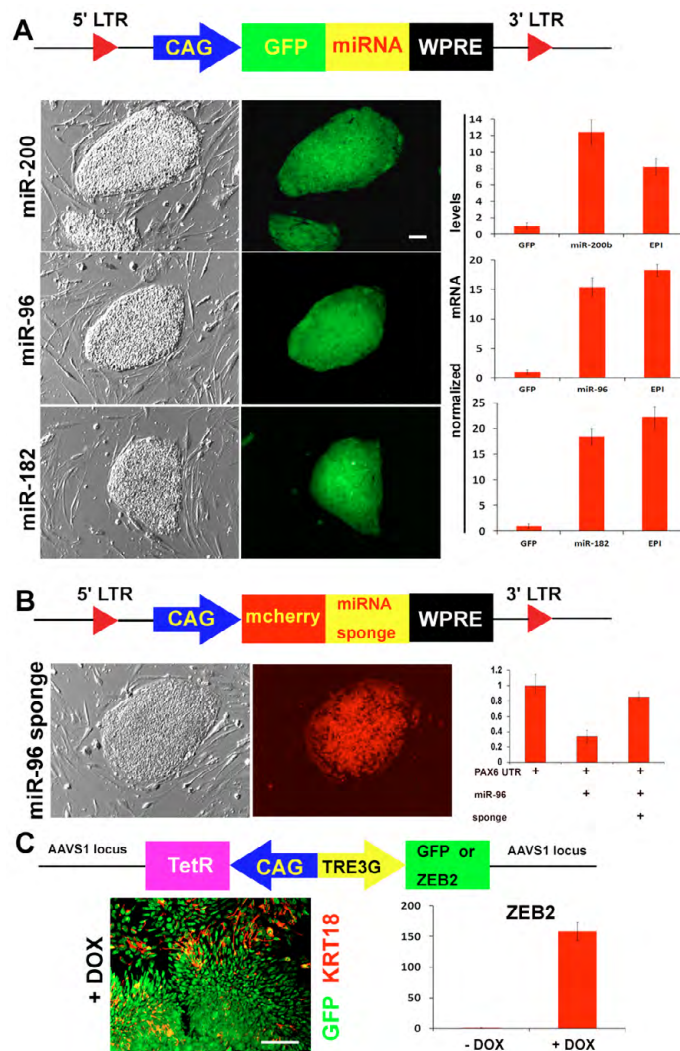
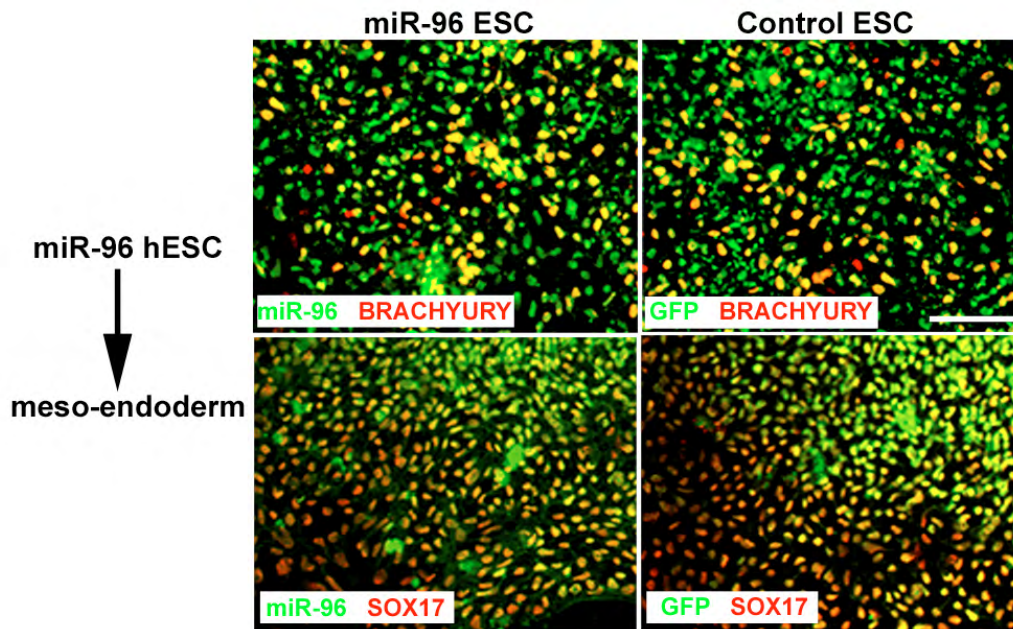


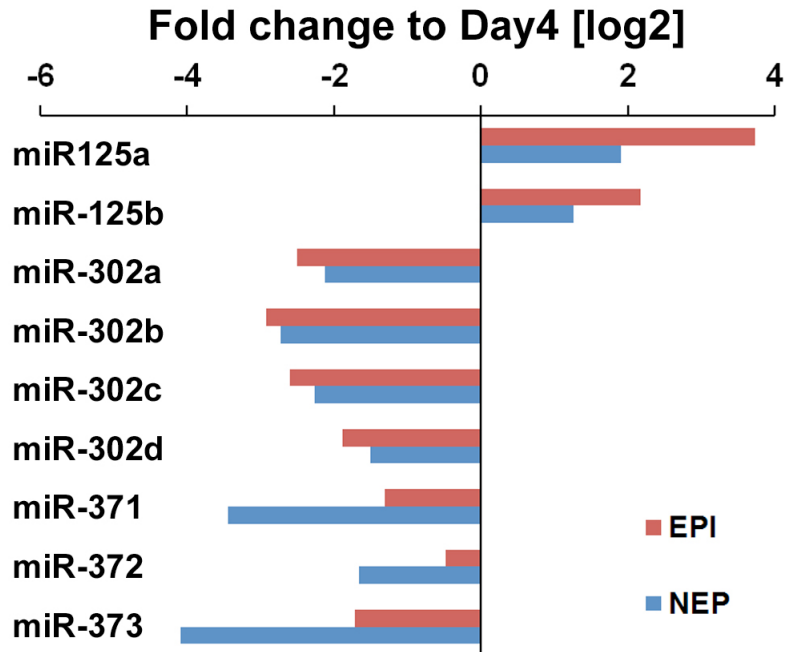
**Fig. S1. The expression pattern of miR-200 and miR-96 families during neural induction from hESCs.** (A) The differential expression of nine members of miR-200 and miR-96 families in day 10 NEP and EPI cells differentiated from hESC line H1. (B) The expression of ZEB2/miR-200 and PAX6/miR-96 was quantified by qPCR during hESC differentiation using the dual Nodal/BMP inhibition method. The relative gene expression levels were normalized to day 0 hESCs.



**Fig. S2. Establishment of miRNA overexpressing and miRNA sponge hESC lines.** (A) The structure of lentiviral vector for overexpressing miRNAs. The miR-200, miR-96 and miR-182 hESCs were visualized under phase-contrast and fluorescent microscopy. Scale bars: 50  $\mu$ m. The right column shows the miRNA levels of corresponding cell lines at day 10 of differentiation when compared with the control GFP hESCs and EPI cells. (B) The structure of lentiviral vector for expressing miRNA sponge. The miR-96-182 sponge hESCs were visualized under phase-contrast and fluorescent microscopy. Scale bars: 50  $\mu$ m. The right column showed the inhibiting activity of miR-96-182 sponge in the reporter assay. (C) Shown is the structure of the inducible expression system in the AAVS1 locus. Induced GFP expression was visualized under fluorescent microscopy in EPI cells (red). Scale bars: 50  $\mu$ m. Induced ZEB2 expression was quantified by qPCR.



**Fig. S3. Overexpression of miR-96 family does not interfere with meso-endoderm differentiation from hESCs.** The miR-96 hESCs (left) and GFP control hESCs (right) were differentiated toward meso-endoderm lineages, and stained for mesoderm marker brachyury and endoderm marker SOX17. Scale bars: 50  $\mu$ m.



**Fig. S4. The expression pattern of miR-125, miR-302 and miR-371 families.** The expression of members of miR-125, miR-302 and miR-371 families was compared during hESC differentiation into the neuroectoderm (NEP) with that of the epidermis (EPI) at day 10.

**Table S1. Fold change of miRNAs in day 10 NEP and EPI cells when compared with day 4 cells (shown as log<sub>2</sub>)**

<b>miRNA</b>	<b>NEP</b>	<b>EPI</b>
let-7b	-2.781158	-2.904227
miR-1	7.388859	5.373822
miR-100	2.096294	2.858324
miR-122a	-3.659820	-2.049011
miR-125a	1.912681	3.732400
miR-125b	1.267455	2.168344
miR-149	1.896574	2.391545
miR-181a	2.154590	3.481077
miR-181b	2.646930	3.764541
miR-181c	2.558172	3.934785
miR-181d	2.683006	3.770286
miR-199a	1.629753	1.917602
miR-199b	2.173490	2.128746
miR-216a	1.995136	2.034965
miR-216b	2.899187	2.256661
miR-217	4.393095	3.552109
miR-26b	2.269018	2.055425
miR-27a	3.883983	2.003635
miR-28	1.790957	2.519526
miR-29c	2.180244	3.367905
miR-301a	2.303150	2.459690
miR-301b	1.937745	2.334515
miR-302a	-2.125153	-2.505480
miR-302b	-2.726693	-2.925864
miR-302c	-2.268908	-2.596383
miR-302d	-1.500470	-1.887767
miR-31	3.382716	3.718355
miR-371	-3.444514	-1.302560
miR-373	-4.091098	-1.720301
miR-432	2.704942	1.688595
miR-495	-15.18792	-18.46438
miR-515	-4.297706	-1.994740
miR-516b	-2.706411	-3.153307
miR-517a	-3.769689	-3.043031
miR-517b	-3.351724	-1.884623
miR-517c	-3.944521	-2.659739
miR-518b	-2.996644	-3.478819
miR-518c	-3.444238	-4.019979
miR-518d	-3.712232	-1.994429
miR-518e	-3.602468	-3.060808

miR-518f	-4.159568	-2.816399
miR-519a	-3.662864	-2.817062
miR-519d	-3.187913	-2.696241
miR-520a	-3.172559	-2.756888
miR-520b	-2.427431	-3.271626
miR-520g	-4.083886	-4.088064
miR-522	-4.611940	-2.272161
miR-523	-3.541162	-2.967402
miR-524	-2.629966	-1.921285
miR-525	-3.885591	-3.721121
miR-526b	-2.376941	-2.360448
miR-614	-2.086076	-2.298528
miR-626	2.352647	2.446251
miR-663	-2.171612	-2.190557
miR-9	3.360588	3.145603
miR-92a	4.278383	2.939442
miR-99a	2.892189	3.902149

**Table S2. Oligonucleotides used in qPCR, cloning and mutagenesis studies**

<b>Primer</b>	<b>Sequence</b>
<b>qPCR</b>	
<b>ZEB1 forward</b>	CTACAACAACAAGACACTGCTGT
<b>ZEB1 reverse</b>	TGTTCTTTCAGAGAGGTAAAGCG
<b>ZEB2 forward</b>	CAA GAG GCG CAA ACA AGC
<b>ZEB2 reverse</b>	GGT TGG CAA TAC CGT CAT CC
<b>CDH2 forward</b>	CAGGGTGGACGTCATTGTAG
<b>CDH2 reverse</b>	AGGGTCTCCACCACTGATTC
<b>EOMES forward</b>	ATCATTACGAAACAGGGCAGGC
<b>EOMES reverse</b>	CGGGGTTGGTATTTGTGTAAGG
<b>SOX7 forward</b>	ACGCCGAGCTCAGCAAGAT
<b>SOX7 reverse</b>	TCCACGTACGGCCTCTTCTG
<b>Cloning</b>	
<b>miR-200 forward</b>	GTCGACCCACTCCGACCTAGTCCTC
<b>miR-200 reverse</b>	GCGGCCGCCTCCGGGTATCTGTGACTGTGAC
<b>miR-96 forward</b>	CTCGAGTCCTTGAAGGTCATCTTGGGCT
<b>miR-96 reverse</b>	GCGGCCGCAGGCAGTGTAAGGCGATCT
<b>miR-182 forward</b>	CTCGAGATGCCTGCCACAGGAACT
<b>miR-182 reverse</b>	GCGGCCGCTGCAGGGAAACACAGAGTGTC
<b>PAX6 3'UTR forward</b>	GTTTAAACGGGACACAACAGTTGAGCTTTC
<b>PAX6 3'UTR reverse</b>	GTCGACAGGCTGACAATGGAAATCTGCC
<b>ZEB2 3'UTR forward</b>	GTTTAAACATACTAGTGGAGTTGGAGCTGGGTATTG

<b>ZEB2 3'UTR reverse</b>	GTCGACACTAGTTGGAATCAGGATCAGTTGAGAA
<b>TS1 site forward</b>	AAACAGATATAAATTCAAGGAAGAAAAAAGTTGA TAGCTAAAAGGTAGAGTGTC
<b>TS1 site reverse</b>	TCGAGACACTCTACCTTTTAGCTATCAACTTTTTTCT TCCTTGAATTTATATCTGTTT
<b>TS2 site forward</b>	AAACCAAATTAACCATTGTTGATTGTAAAAACCA TGCCAAAGCCTTTGTATTC
<b>TS2 site reverse</b>	TCGAGAATACAAAGGCTTTGGCATGGTTTTTTACAAT CAACAATGGTTAATTTTGGTTT
<b>Mutagenesis</b>	
<b>TS1 site</b>	ATTCAAGGAAGAAAAAAGTTGATAGGGAAAAGGT AGAGTGTGTCT
<b>TS2 site</b>	AAAATTAACCATTGTTGATTGTAAAAAACCATGGGA AAGCCTTTGTATTC