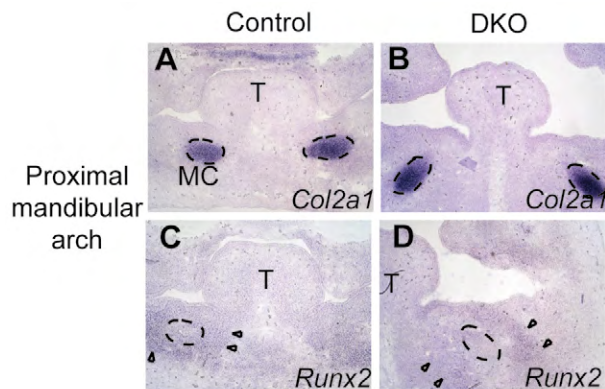
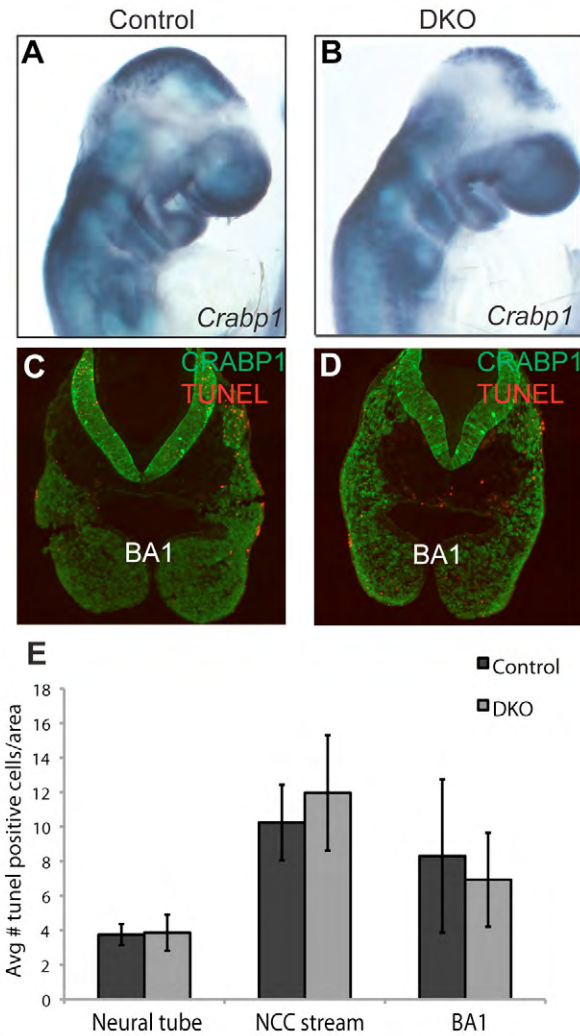


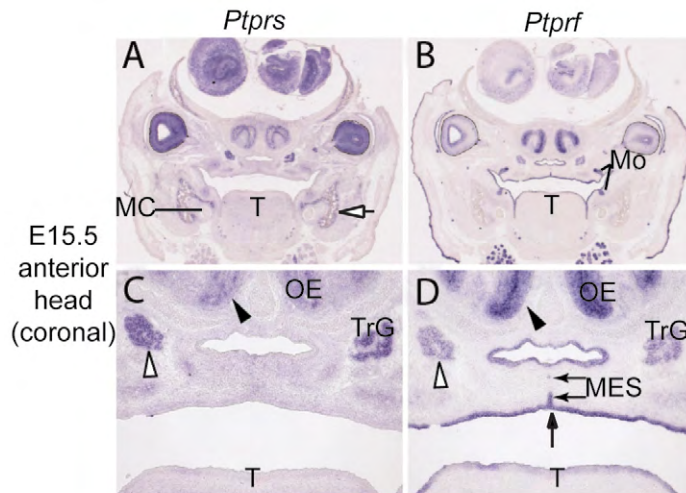
**Fig. S1. Appendicular and axial skeletal development is normal in *Ptprs<sup>-/-</sup>;Ptprf<sup>-/-</sup>* embryos.** Alizarin Red and Alcian Blue stainings of bone and cartilage. (A,B) Bone and cartilage development of the DKO body is normal at E18.5. (C-F) Bone and cartilage development of the DKO body (C,D) and middle ear capsule (E,F) are normal at E14.5. Note that Meckel's cartilage articulates normally with the middle ear. MC, Meckel's cartilage; ME, middle ear.



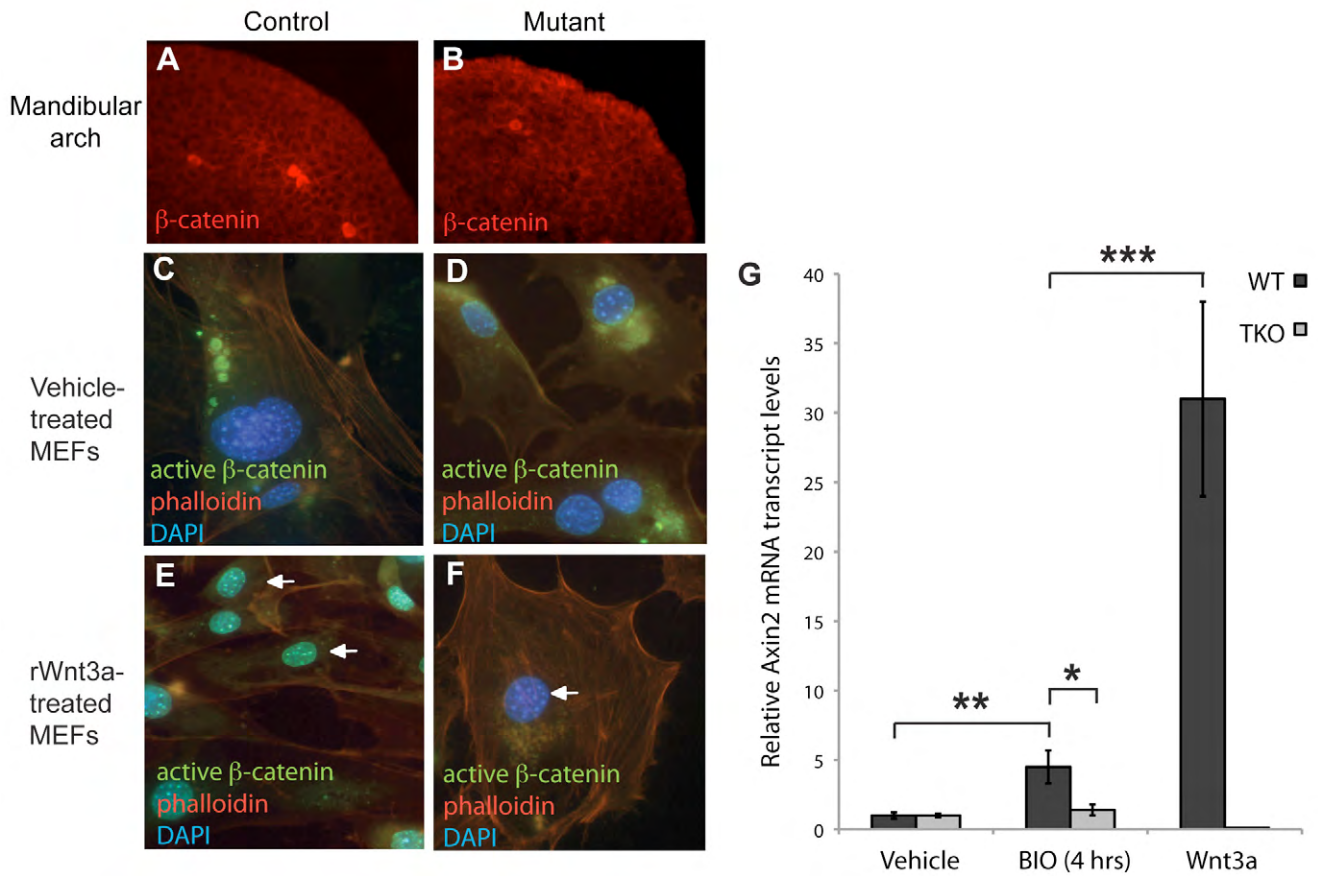
**Fig. S2. Proximal Meckel's cartilage is specified normally in *Ptprs<sup>-/-</sup>;Ptprf<sup>-/-</sup>* embryos.** (A-D) *In situ* hybridization using probes to *Col2a1* and *Runx2* reveals normal proximal Meckel's cartilage specification in DKO mandibular arch at E12.5. Differentiation of proximal bone precursors also occurs normally (arrowheads). MC, Meckel's cartilage; T, tongue.



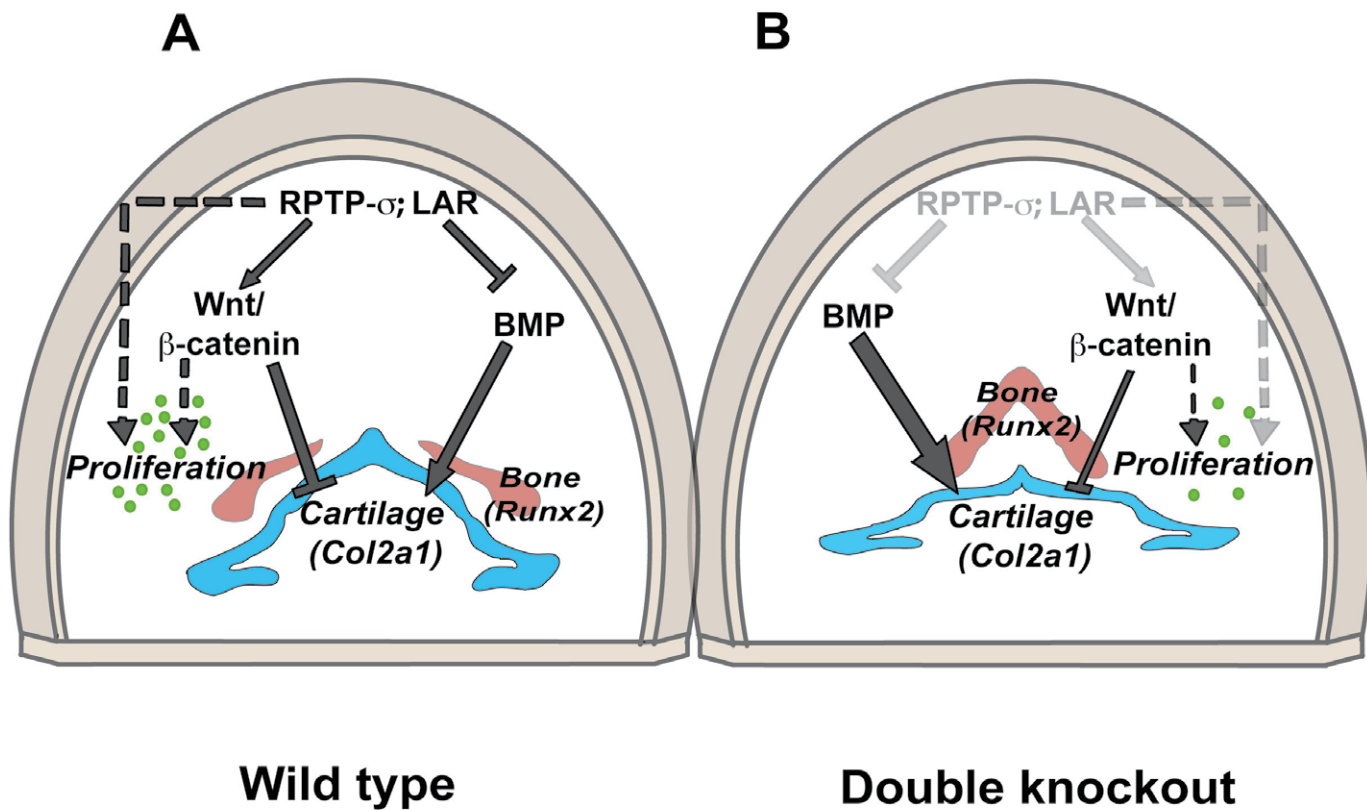
**Fig. S3. Neural crest migration and survival in the first branchial arch.** (A,B) Whole-mount *in situ* hybridization using a probe for cellular retinoic acid binding protein (CRABP1) as a marker of migratory neural crest cells reveals normal migration in DKO embryos at E9.5. (C-E) Immunofluorescent labeling of CRABP1 and TUNEL reveals no significant difference in neural crest cell (NCC) apoptosis in either the migratory stream or first branchial arch (BA1) at E9.5.



**Fig. S4. Expression of *Ptpvf* and *Ptpvs* during palatal shelf fusion.** *In situ* hybridization using probes for *Ptpvf* (A,C) and *Ptpvs* (B,D) on coronal E15.5 sections. (A,C) *Ptpvf* is expressed in the medial epithelial seam (MES, black arrows) where palatal shelf fusion occurs, as well as throughout the oral epithelium. (B,D) *Ptpvs* is expressed diffusely in the palatal shelf mesenchyme and in ossification centers surrounding Meckel's cartilage (open arrow). Additional regions of strong LAR family expression are indicated by arrowheads. MC, Meckel's cartilage; Mo, molar tooth buds; OE, olfactory epithelium; T, tongue; TrG, trigeminal ganglion.



**Fig. S5.  $\beta$ -catenin activation is impaired in mouse embryonic fibroblasts mutant for LAR family phosphatase activity.** (A,B) Immunofluorescent staining for total  $\beta$ -catenin protein on coronal E10.5 pharyngeal arch sections. There is no significant difference in  $\beta$ -catenin levels between control (A) and *Ptprs*<sup>-/-</sup>;*Ptprf*<sup>-/-</sup> (B) embryos. (C-F) Immunofluorescent staining of (C,E) wild-type (WT) mouse embryonic fibroblasts (MEFs) and (D,F) *Ptprs*<sup>-/-</sup>;*Ptprf*<sup>-/-</sup>;*Ptprd*<sup>-/-</sup> triple knockout (TKO) MEFs for Ser27/37 dephosphorylated (active)  $\beta$ -catenin reveals a specific defect in the activation of  $\beta$ -catenin in TKO MEFs upon rWNT3A stimulation (arrows). (G) qPCR analysis of *Axin2* transcription reveals that treatment with the GSK3 $\beta$  inhibitor BIO alone is insufficient to restore  $\beta$ -catenin activity, confirming the specificity of the rescue to the Wnt pathway. \* $P$ <0.1, \*\* $P$ <0.05, \*\*\* $P$ <0.005.



**Fig. S6. Model for LAR family phosphatase activity during pharyngeal arch development.** (A) Normally, PTPRS (RPTP $\sigma$ ) and PTPRF (LAR) act to dampen Bmp signaling activity and promote canonical  $\beta$ -catenin activity, which serves to correctly specify Meckel's cartilage and downstream initiation of mandibular bone development. In addition, PTPRS and PTPRF promote mandibular arch proliferation, potentially through their action on canonical Wnt signaling. (B) In *Ptprs*<sup>-/-</sup>;*Ptprf*<sup>-/-</sup> embryos, loss of the phosphatases permits concomitant intensification of Bmp signaling and downregulation of canonical Wnt restriction of cartilage development, thereby altering the specification of Meckel's cartilage and indirectly affecting mandibular bone positioning. Loss of the LAR family also disrupts proliferation, further affecting mandibular development.

Table S1. Primer sequences for qPCR or ISH

Gene	Use	Forward primer	Reverse primer
beta-microglobin	qPCR	GACTGGTCTTTCTATATCCTGG	CTTTCTGCGTGCATAAATTG
<i>Axin2</i>	qPCR	CCATTTTGGACGACCACCTCTC	GAAGAAGGGTATGACACTGCTGATG
cyclin D1	qPCR	TCTTTCATTGGGCAACGGG	TCCTCAGTTTGGATGGCTCTCC
<i>Ptc1</i>	qPCR	TCTGCTGGGTGTACTGATGC	TCAGGACACGGTCCAAGA
<i>Msx1</i>	qPCR	TCCTCAAGCTGCCAGAAGAT	TTGGTCTTGTGCTTGCGTAG
<i>Dlx1</i>	qPCR	TTCATCTGACGCTGAGTGTTGG	TTTCTGTCTTGTTCCTCTTC
<i>Dlx5</i>	qPCR	TCAGGAATCGCCAACCTTGC	TGCCATAAGAAGCAGAGGTAGGAG
<i>Gsc</i>	qPCR	TAAGAACCGCCGAGCCAAG	CCGAGTCAAATCGCTTTTACC
<i>Sox9</i>	qPCR	TCGGAAGTGCCTGGAACTTC	GAGGGAGGGAAAACAGAGAACC
<i>Col2a1</i>	qPCR	GAGCAGCAAGAGCAAGGAAAAG	CAGTGGACAGTAGACGGAGGAAAAG
<i>Runx2</i>	qPCR	ACCAGTCTTACCCCTCCTATCTGAG	GCAGTGTTCATCATCTGAAATACGC
<i>Msx1</i>	ISH	CCTACGCAAGCACAAGACCAAC	TGGGGACCACGGATAAATCTC
<i>Col2a1</i>	ISH	CATTATTGACATCGCACCC	CAGAATAGCACCATTGTGTAGG
<i>Runx2</i>	ISH	GAGCTATTAAGTGACAGTGGACGG	CAACACCATCATTCTGGTTAGGC
<i>Crabp1</i>	ISH	CAGCAGAGGTGGGTGCCTGCC	GCAGCCAACCAGTTTAATGACTGG

All primers are shown 5' to 3'.