

Fig. S1. Splice-blocking efficacy, primers used to assay splicing, left-right markers examined in 3-OST-5 morphants, tracks of fluorescent beads injected into the KV at 6 SS, 3-OST-5 RNA expression, midline marker expression. (A) Representative *in situ* images illustrating normal (black arrows) and reversed (red arrows) heart (*cmlc2*) and gut (*fkd2*) looping in 3-OST-5 morphants. (B) Table containing primers used for detecting spliced versus unspliced transcript when embryos were injected with translational splice blocking morpholinos. (C) Dose-dependent splice blocking efficacy of 3-OST-5 MO1. Increasing amounts of MO injected resulted in a 70±12% decrease of spliced mRNA. (D) Dose-dependent splice blocking efficacy of control 3-OST-3Z MO1. Increasing MO dose resulted in a 93±4% decrease of spliced mRNA. ImageQuant TL software was used to semi-quantitatively measure band intensity. The values listed are the percentage of spliced product compared with uninjected WT. (E,F) Whole-mount *in situ* analysis of 3-OST-5 mRNA, which was expressed ubiquitously throughout the embryo during epiboly (E) and early somitogenesis (F). Black arrow indicates staining in DFCs, white arrow indicates the position of KV. (G,H) Whole-mount *in situ* analysis of *shh* expression in uninjected (G) and 3-OST-5 morphants (H). (I,J) Whole-mount *in situ* analysis of *lefty1* expression in the midline of uninjected (I, *n*=113) and 3-OST-5 morphants (J, *n*=82), 3-OST-5 morphants had normal expression of *lefty1* expression in the midline. (K-M) Tracks of fluorescent beads injected into KV of uninjected (K) and 3-OST-3Z MO1 embryos (L) with normal bead flow, and 3-OST-5 MO1 embryos (M), displaying uncoordinated bead movement due to short but motile cilia.

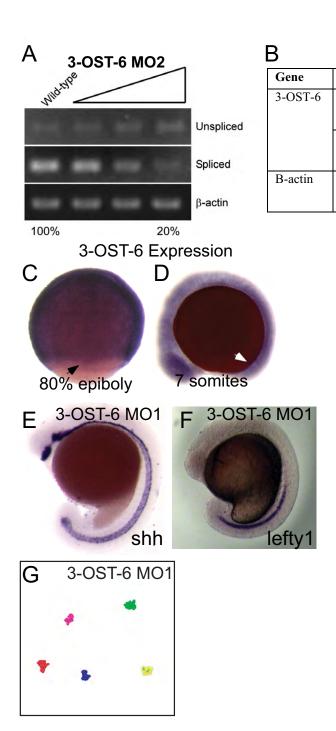


Fig. S2. Splice-blocking efficacy, primers used to assay splicing, tracks of fluorescent beads injected into the KV at 6 SS, 3-OST-6 RNA expression, midline marker expression. (A) Analysis of 3-OST-6 MO2 splice-blocking efficacy. Increasing amounts of MO injected resulted in an  $80\pm20\%$  decrease of spliced mRNA. Analysis was performed as described in Fig. S1. (B) Table containing primers used for detecting spliced versus unspliced transcript when embryos were injected with translational splice blocking morpholinos. (C,D) Whole mount *in situ* analysis of 3-OST-6 mRNA, which was expressed ubiquitously throughout the embryo during epiboly (C) and early somitogenesis (D). Black arrow indicates staining in DFCs, white arrow indicates the position of KV. (E) Whole-mount *in situ* analysis of *shh* expression, which was normal in 3-OST-6 morphants (n=109). (F) Whole-mount *in situ* analysis of *lefty1* expression in the midline, was normal in 3-OST6 morphants (n=49). (G) Bead tracks of fluorescent beads injected into 3-OST-6 MO1 embryos showing a lack of bead movement, indicative of non-motile cilia.

**Splice State** 

Spliced

Unspliced

Load control

Primer

5'-GCTTCCACACGCGCTGATCATC-3'

5'-GCTTCCACACGCGCTGATCATC-3'
5'-AGCCCATATACATTGTTTGCAACC-3'

5'-GGTATGGGACAGAAAGACAG-3'

5'-AGAGTCCATCACGATACCAG-3'

5'-ATGCGGCCTGGAGTTTCTGGAGTC-3'



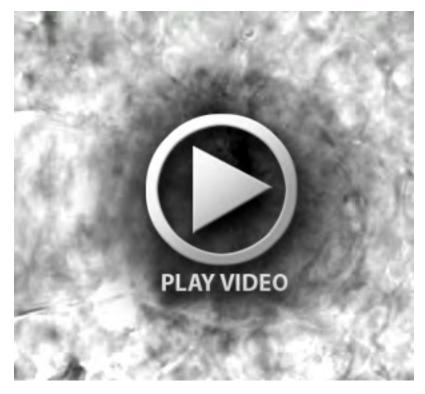
**Movie 1. Uninjected control embryos, normal KV flow.** Embryos were injected at 8 SS with fluorescent beads and filmed by DIC microscopy (to show orientation of notochord, to the left of each panel) and then by fluorescent imaging to capture bead movement.



**Movie 2. 3-OST-3Z MO, normal KV flow.** Embryos were injected at 8 SS with fluorescent beads and filmed by DIC microscopy (to show orientation of notochord, to the left of each panel) and then by fluorescent imaging to capture bead movement.



**Movie 3. 3-OST-5 MO, aberrant KV flow.** Embryos were injected at 8 SS with fluorescent beads and filmed by DIC microscopy (to show orientation of notochord, to the left of each panel) and then by fluorescent imaging to capture bead movement.



**Movie 4. 3-OST-6 MO, absent KV flow.** Embryos were injected at 8 SS with fluorescent beads and filmed by DIC microscopy (to show orientation of notochord, to the left of each panel) and then by fluorescent imaging to capture bead movement.