

Fig. S1. Quantification of the number of Lhx2/9⁺ cells in control and caBmprIb electroporated spinal cords. Electroporation of fGFP (n=69 sections from three embryos) or caBmprIb-IRES-fGFP+ (n=68 sections from four embryos) does not affect the number of Lhx2/9⁺ neurons generated. Equivalent numbers of Lhx2/9⁺ neurons are present on the electroporated and non-electroporated sides of HH stage 18 spinal cords (fGFP: probability of similarity, P>0.29; caBmprIb: probability of similarity, P>0.16). Error bars represent s.e.m.

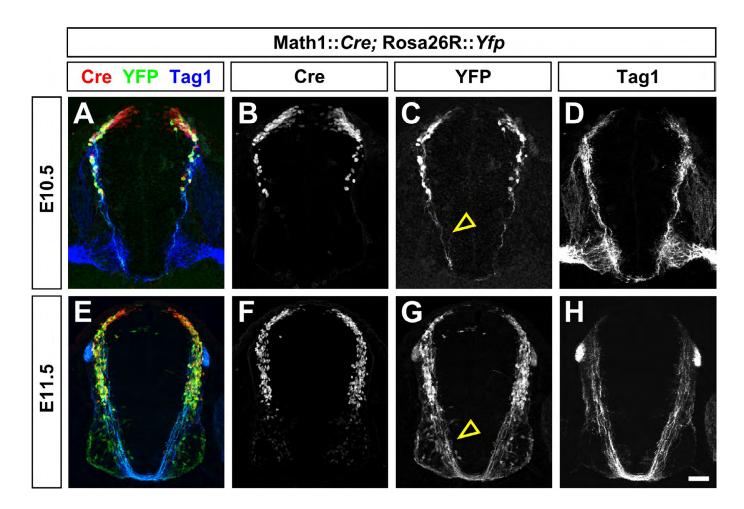


Fig. S2. The Math1 enhancer drives YFP expression in commissural neurons. (A-H) The Math1::Cre line can drive the expression of YFP in Tag1⁺ commissural axons (arrowheads, C,G) when crossed to the Cre reporter strain, Rosa26R(lox-stoplox)::Yfp. Transverse spinal sections, taken from E10.5 (A-D) and E11.5 (E-H) Math1::Cre; Rosa26R::Yfp embryos, were labeled with antibodies against Cre (red, A,B,E,F), GFP (green, A,E,C,G) and Tag1 (blue, A,E,D,H). Scale bars: 45 µm in A-D; 55 µm in E-H.

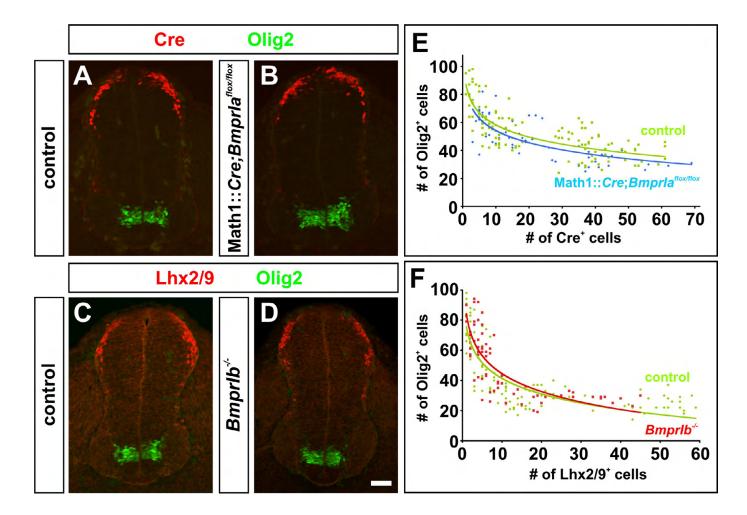


Fig. S3. The loss of BmprIa or BmprIb has no effect on the fate of commissural neurons. (A-D) There was no observable difference in the number of Lhx2/9⁺ cells in the presence or absence of either BmprIa or BmprIb. Transverse sections were taken from brachial or thoracic levels of the spinal cord from E10.5 Math1::*Cre*; *BmprIa*^{+/+}(control, A), Math1::*Cre*; *BmprIa*^{flox/flox} (B), *BmprIb*^{+/+} (control, C) and *BmprIb*^{-/-} (D) embryos and labeled with antibodies against Cre (red, A,B), Lhx2/9 (red, C,D) and Olig2 (green). (E,F) The numbers of Cre⁺ (E) or Lhx2/9⁺ (F) cells were plotted as a function of Olig2⁺ cell number to normalize the extent of development between embryos. A logarithmic regression analysis reveals no difference between the distribution of Cre⁺/Olig2⁺ cells in sections from Math1::*Cre*; *BmprIaflox/flox* (*n*=59 sections from four embryos) and *BmprIb*^{+/+} (*m*=40 sections from four embryos) littermates. Scale bar: 25 µm.