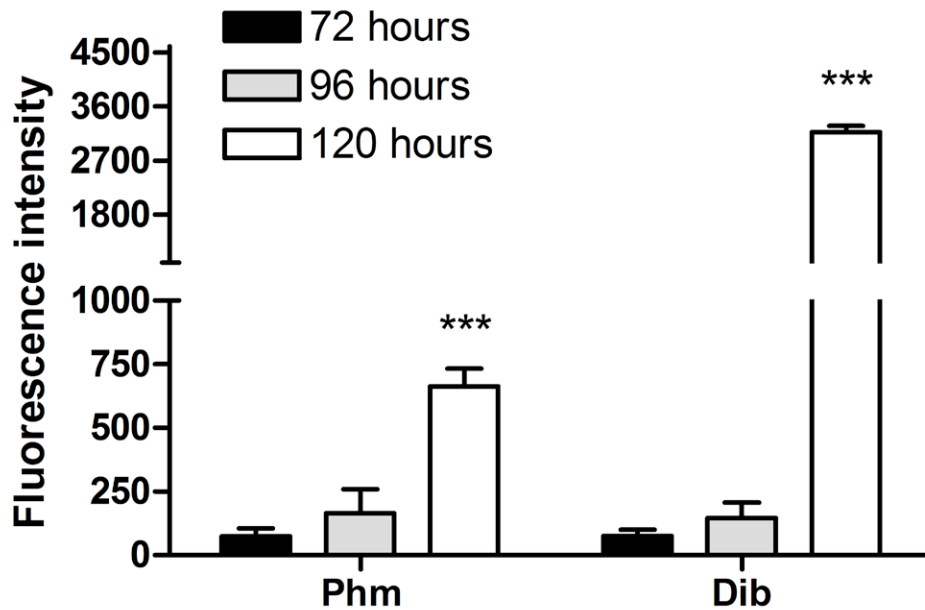
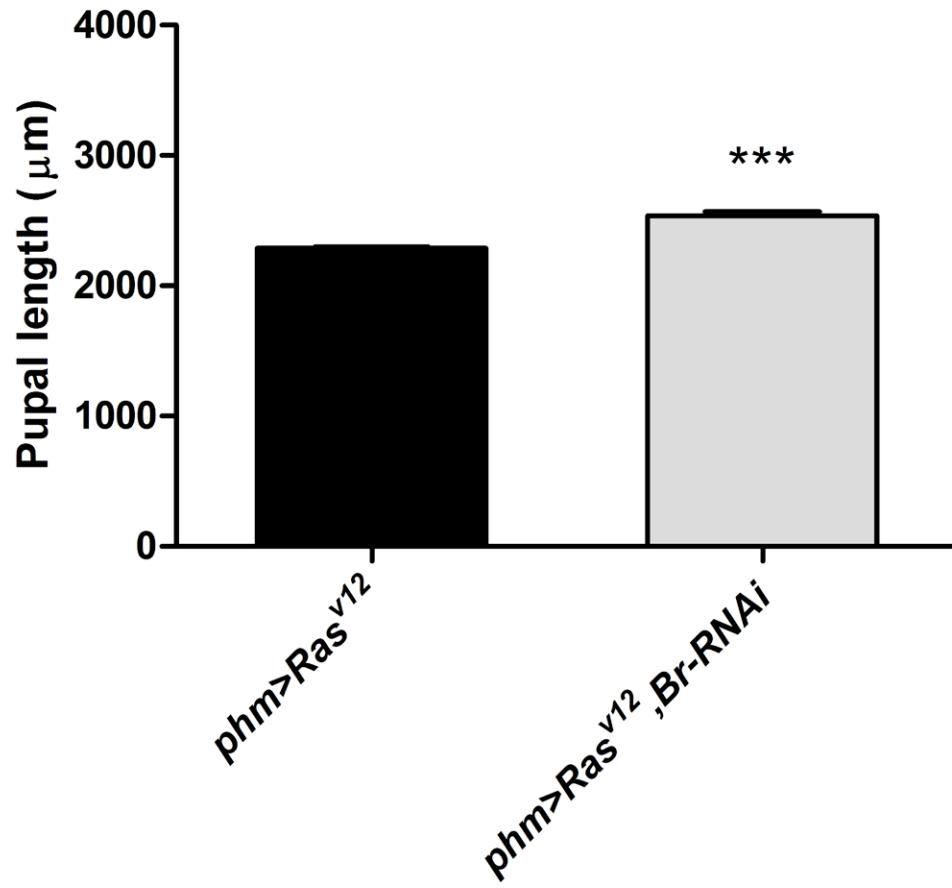


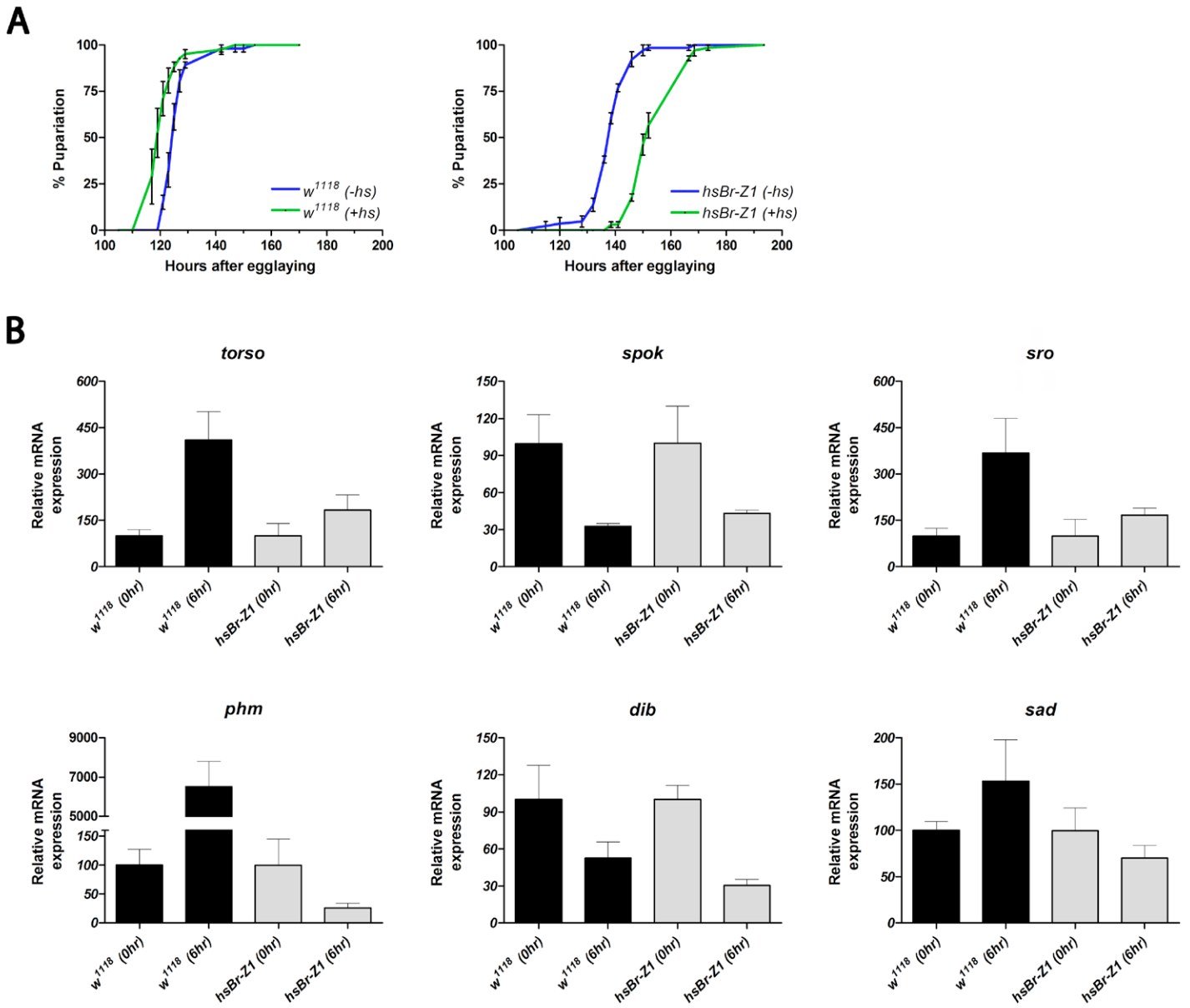
**Fig. S1. Expression of Br protein in nuclear extract from S2 cells.** Western blot with anti-Br-core show Br isoforms in S2 cell nuclear extract (lane 1) and tissue from third instar *w<sup>1118</sup>* wandering larvae (lane 2). The Br-Z4 isoform (~92 kDa) and the Br-Z1/Z3 isoforms (~75 kDa) are present in both S2 cell nuclear extract and in third instar larval tissue.



**Fig. S2. Quantification of Phm and Dib protein expression during the third instar.** Immunohistochemistry images (related to figure 3) of Phm and Dib expression during the third instar at 72, 96 and 120 hours AEL were subjected to image quantification analysis. Fluorescence intensity within the PG was quantified using Image J ( $n \geq 5$ ).



**Fig. S3. Reducing *Br* expression causes overgrowth in larvae overexpressing *Ras*<sup>V12</sup> in PG.** Effect on size of *Br-RNAi* in the PG of larvae overexpressing *Ras*<sup>V12</sup> (n=40-55).



**Fig. S4. EcR and Br-Z1 are involved in negative feedback that inhibits PG steroidogenic activity after pupariation.** (A) Heat shock at 112 hours AEL (prior to the high level ecdysone pulse) delays pupariation of *hs-Br-Z1* larvae compared to non-heat shocked controls. Heat shock of wild type (*w<sup>1118</sup>*) larvae at this time does not delay pupariation (B) Heat shock induced Br-Z1 over-expression 112 hours AEL (T=0 hr) inhibits the developmental increase in expression of genes involved in ecdysone biosynthesis and *torso* (*tor*) observed 6 hours later (118 hours AEL: T=6 hr) in the *phm*>+ control.

**Table S1. Primers for amplification of genomic fragments**

Name	5'-3'
<i>phmF1</i>	aatAGATCTcggcatatattatgggtctgc
<i>phmF2</i>	aatAGATCTaggtagacggtcgattgaata
<i>phmF3</i>	aatAGATCTcgaactgatcccagatttctac
<i>phmF4</i>	aatAGATCTtgggtgtaatgaatgtgcatac
<i>phmF5</i>	aatAGATCTgatgccgaaaccctgtatgct
<i>phmF6</i>	aatAGATCTgtatatggtatatatgggtggca
<i>phmF7</i>	aatAGATCTgagaatctgggaaccaagag
<i>phmR1</i>	aataGCTAGCcacttctgattcctcctgctc
<i>phmR2</i>	aataGCTAGCccttcttggttcccagattctc
<i>phmR3</i>	aataGCTAGCtgccacacatatacatatac
<i>phmR4</i>	aataGCTAGCaaggagagtgcgaaagtaaaatt
<i>dibF1</i>	tctggtcctcccttcggtcaccg
<i>dibR1</i>	attcgtattcacgcagcagctga

Capital letters indicate restriction enzyme sites: *Bgl*III in forward (F) primers and *Nhe*I in reverse (R) primers.

**Table S2. Primers for site-directed mutagenesis**

Name	5'-3'
<i>phmF mut1</i>	CCCGGGCTGCAGGAATTCAGATCTTGGCATATGTGAATGTGCATACGAT AAACGGGCAATTTC
<i>phmR mut1</i>	GAAATTGCCCGTTTATCGTATGCACATTCACATATGCCAAGATCTGAATT CCTGCAGCCCGGG
<i>phmF mut2</i>	GCTGCAGGAATTCAGATCTTGGGTGTAATGAACATATGTACGATAAACG GGCAATTTCAATTTCAAATTTACTTTTCGC
<i>phmR mut2</i>	GCGAAAGTAAAATTTGAAAATGAAATTGCCCGTTTATCGTACATATGTT CATTACACCCAAGATCTGAATTCCTGCAGC
<i>phmF mut3</i>	CAGATCTTGGGTGTAATGAATGTGCATACGCATATGGGGCAATTTCAATTT TCAAATTTACTTTTCGCACTC
<i>phmR mut3</i>	GAGTGCAGAAAGTAAAATTTGAAAATGAAATTGCCCCATATGCGTATGCA CATTACATTACACCCAAGATCTG
<i>phmF mut4</i>	GGTGTAATGAATGTGCATACGATAAACGGGCAACATATGTTTCAAATTT TACTTTTCGCACTCTCCTTCTTCG
<i>phmR mut4</i>	CGAAGAAGGAGAGTGCAGAAAGTAAAATTTGAAACATATGTTGCCCGTTT ATCGTATGCACATTCATTACACC
<i>phmF mut5</i>	GCATACGATAAACGGGCAATTTCAATTTCAAATTCATATGTCGCACTCTC CTTCTTCGATGCCGAAACCC
<i>phmR mut5</i>	GGGTTTCGGCATCGAAGAAGGAGAGTGCACATATGAATTTGAAAATGA AATTGCCCGTTTATCGTATGC

**Table S3. EMSA oligos**

Name	Forward, 5'-3'	Reverse, 5'-3'
** <i>phm</i> <i>Br-</i> <i>Z1/Z4</i>	GCAATTT <u>CATTTTCAAATTTCCCTG</u> TCGC	TGCGACAGGGAAATTTGAAAATG AAATTG
** <i>phm</i> <i>Br-</i> <i>Z1/Z4</i> <i>mut</i>	GCAATTT <u>CATCCCTGAATTTCCCTG</u> TCGC C	TGCGACAGGGAAATTCAGGGATG AAATTG
<i>dib Br-</i> <i>Z4</i>	TGAGCTAACGGAAACAGATCAGCA	GTGCTGATCTGTTTCCGTTAGCTC
<i>dib Br-</i> <i>Z4 mut</i>	TGAGCTAAC <u>CCCTG</u> CAGATCAGCA	GTGCTGATCTGCAGGGGTTAGCT C
*** <i>Br-</i> <i>Z4/Z1</i>	CCTTTTTTTTTTATTTATGAAGTAATT	GAATTACTTCATAAATAAAAAAA AAAG
Non- specific	AACGTAGCTGATCGAATCGGTTAC	AGTAACCGATTCGATCAGCTACG T

Mutations introduced are underlined.

\*\*Oligos have a mutated downstream Br-Z3 site allowing the study of Br-Z1/Z4 site alone.

\*\*\*Oligos described in Wang et al. (Wang et al., 2009).

**Table S4. qPCR primers**

Name	Forward, 5'-3'	Reverse, 5'-3'
* <i>rpL23</i>	GACAACACCCGGAGCCAAGAACC	GTTTGCGCTGCCGAATAACCAC
<i>E74A</i>	TTTCTCTGCCGTTGTCGTC	GCACTGAGACCCGCTCAC
<i>E74B</i>	CGCGAGTTCAAAGTGCTCTA	GGAGGGAGAGTGGTGGTGT
<i>E75B</i>	CAACAGCAACAACACCCAGA	CAGATCGGCACATGGCTTT
* <i>phantom (phm)</i>	GGATTTCTTTTCGGCGCGATGTG	TGCCTCAGTATCGAAAAGCCGT
* <i>disembodied (dib)</i>	TGCCCTCAATCCCTATCTGGTC	ACAGGGTCTTCACACCCATCTC
* <i>spookier (spok)</i>	TATCTCTTGGGCACACTCGCTG	GCCGAGCTAAATTTCTCCGCTT
* <i>neverland (nvd)</i>	GGAAGCGTTGCTGACGACTGTG	TAAAGCCGTCCACTTCCTGCGA
* <i>shadow (sad)</i>	CCGCATTCAGCAGTCAGTGG	ACCTGCCGTGTACAAGGAGAG
<i>shroud (sro)</i>	AGCAGCTGAAGGTCGATAGC	GCGATTCGTGGCAGTAAAC
<i>torso (tor)</i>	TGCTTGGATTGGTATCCCTATAA	TGGGTACACAGTAAGATTCTCTGG
<i>Br-Z4</i>	CAAAGGCACACACACACACA	TCGGTCGGTTCTTCTCCTTC
<i>Br-Z1</i>	CCAGACCAACACCCACACAC	ACATTCGCTTCCTTCGTCCT

\*Primers described previously (McBrayer et al., 2007).