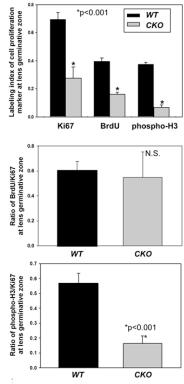
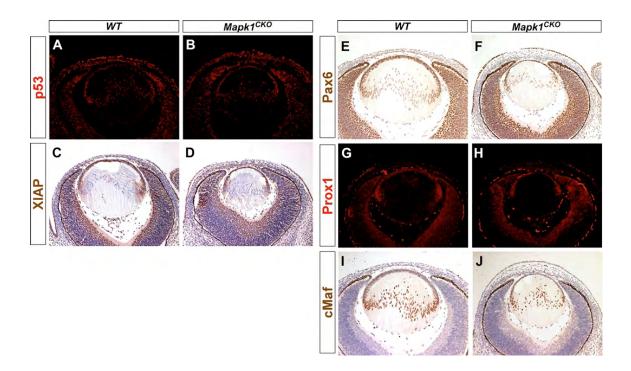


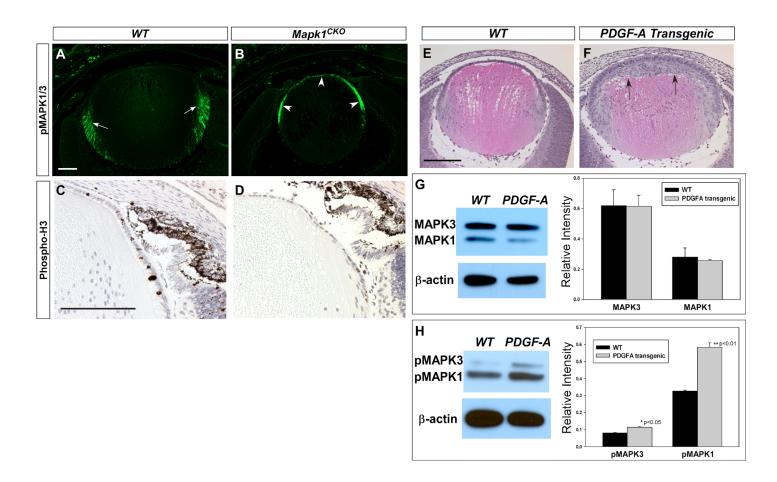
**Fig. S1. Phalloidin staining and lens/eye histology.** (**A**,**B**) Phalloidin labels F-actin in P2 mouse lenses. Compared with WT lens,  $Mapk1^{CKO}$  lens had disorganized fiber cells. Some areas showed a degenerative phenotype (arrow in B). (**C**,**D**) Anterior segment defects. The corneal endothelium and anterior chamber were absent in  $Mapk1^{CKO}$  eyes. The iris developed abnormally and was attached to the cornea in  $Mapk1^{CKO}$  eyes. (**E-H**) Despite the lens defects, retinal development did not seem to be affected at E14.5 (F) and P0 (H) in  $Mapk1^{CKO}$  eyes when compared with WT eyes (E,G, respectively). (**I**,**J**) No eye phenotype was observed in E14.5 or E16.5  $Mapk3^{-/-}$  mice. Scale bars: 100 µm.



**Fig. S2. Cell proliferation markers in the lens germinative zone (10-30° as shown in Fig. 3A,B).** The labeling indices for all three markers were significantly reduced (top panel). When the ratios of BrdU/Ki67 and phospho-H3/Ki67 were calculated, we found no significant change in the BrdU/Ki67 ratio between WT and  $Mapk1^{CKO}$  lenses (middle panel), whereas the ratio of phospho-H3/Ki67 was significantly reduced in  $Mapk1^{CKO}$  lenses (bottom panel). These data suggest that the G2-M phase was most affected by MAPK1 deficiency. We also speculate that the decrease in Ki67 index in  $Mapk1^{CKO}$  lens could be a result of a shortened late G2/M phase, extended G0/G1 phase, or a combination of both.



**Fig. S3.** Expression of apoptosis regulatory proteins (p53 and XIAP) and transcription factors (PAX6, PROX1 and cMAF) important for lens development in E14.5 mouse lens. (A-J) No change in expression patterns was observed for these proteins between WT and *Mapk1<sup>CKO</sup>* lenses.



**Fig. S4. MAPK activation and cell proliferation analysis.** (A,B) pMAPK1/3 immunofluorescence was performed without TSA enhancement. In *Mapk1<sup>CKO</sup>* lens, the level of pMAPK (equivalent to pMAPK3) was increased in the epithelium, likely as a result of an adaptive response to compensate for the loss of MAPK1 activity. Furthermore, the hyperactive region of pMAPK (arrows in A, WT) was shifted anteriorly in *Mapk1<sup>CKO</sup>* lens (arrowheads in B). This change could be caused by the reduction in lens size and consequent anterior shift of the developing iris and ciliary body in the *Mapk1<sup>CKO</sup>* eye. (C,D) Phospho-H3 immunohistochemistry. Increase of pMAPK3 did not rescue cell proliferation defects in *Mapk1<sup>CKO</sup>* lens. (E-H) *PDGFA* transgenic mice. PDGF-A overexpression in the lens induced overproliferation of the lens epithelial cells (arrows in F). MAPK1/3 protein levels were unaltered, but pMAPK1/3 levels were higher in *PDGFA* transgenic lenses (G,H) than in WT lenses. A 40% increase in the pMAPK3 levels and an 80% increase in the pMAPK1 levels occurred in *PDGFA* transgenic lenses, compared with WT lenses, suggesting that MAPK1 is more responsive to PDGF-A stimulation. Scale bars: 100 µm. \**P*<0.05, \*\**P*<0.01.