

SUPPLEMENTARY MATERIALS AND METHODS

Fly stocks and genetics

Mutants used included *slmb*¹, *slmb*², *slmb*⁸, *dlg*⁴⁰⁻², *Roc1a*^{G1}, *AP-2sigma* and *yki*^{B5} (described in FlyBase). Transgenes included *tub>slmb-myc* (Ko et al., 2002), *UAS-Ci*^{M1-4} (Chen et al., 1999), *UAS-Arm*^{S10} (Pai et al., 1997), *UAS-Plk4*^{SBM} (Rogers et al., 2009), *UAS-CapH2*^{SBM} (Buster et al., 2013), *UAS-Par1*^{T408A} (Lee et al., 2012), *UAS-aPKC*^{CAAX-DN} (Sotillos et al., 2004) and *UAS-aPKC*^{SN} (Betschinger et al., 2003) driven by *MS1096-GAL4*, as well as *hs-Wls-V5* and *UAS-Wls-V5* (Belenkaya et al., 2008). Entirely mutant wing discs were generated using *UbxFLP/FM7; cl FRT82B/TM6B* and entirely mutant eye discs were generated using *eyFLP cl GMRhid FRT82B/TM6B*. MARCM clones in the eye and neuroblast were generated with *eyFLP* and *hsFLP* stocks, respectively. Follicle cell clones were generated as described (Lu and Bilder, 2005).

Immunohistochemistry

The following primary antibodies were used: mouse anti-Mmp1 (1/100), mouse anti-Arm (N27A1, 1/100), mouse anti-Dlg (4F3, 1/100), mouse anti-Coracle (1/100), mouse anti-FasIII (7G10, 1/20), mouse anti-Notch^{ECD} (C458.2H, 1/50), mouse anti-Lamin (1/100), rat anti-Elav (9F8A9, 1/50) (all from Developmental Studies Hybridoma Bank, see references therein), rat anti-Crb (1/750; U. Tepass, E. Knust), guinea pig anti-Cad87E (1/1000; U. Tepass), guinea pig anti-Scrib (1/200), rabbit anti-PKCζ (sc-216, Santa Cruz Biotechnology, 1/200), rabbit anti-Miranda (1/500), mouse anti-Prospero (1/100). TRITC-phalloidin was used to visualize F-actin (1/400, Sigma) and either TO-PRO-3 (1/400) or DAPI (1/3000) was used to visualize DNA. Secondary antibodies were from Molecular Probes.

Supplementary references

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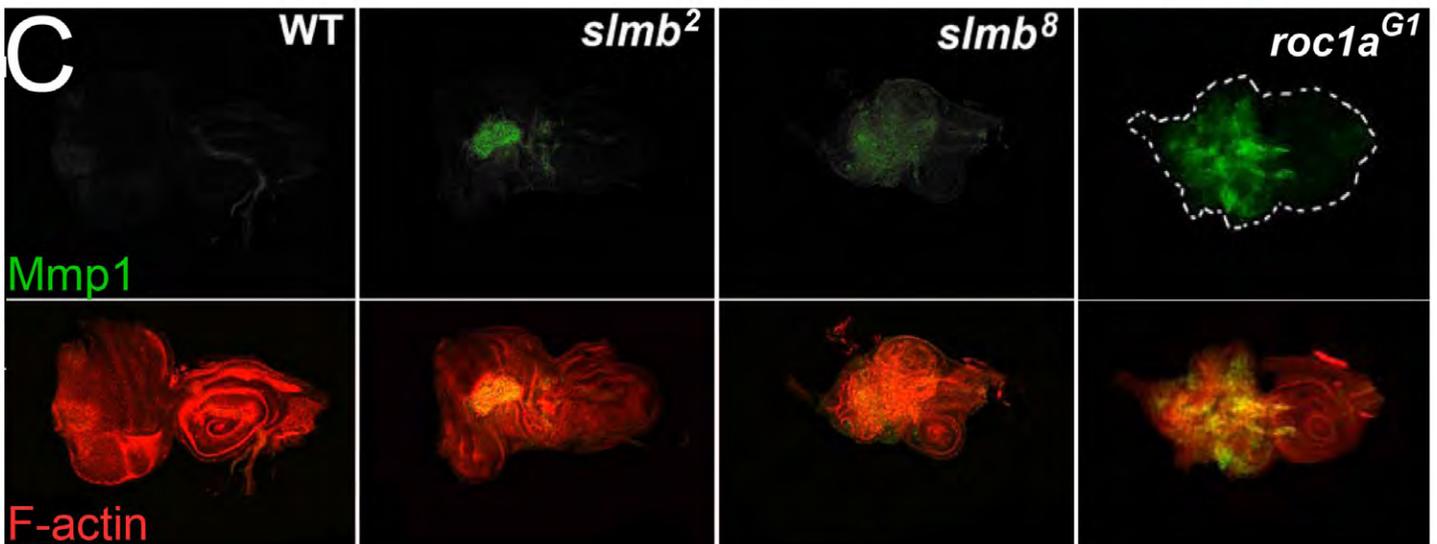
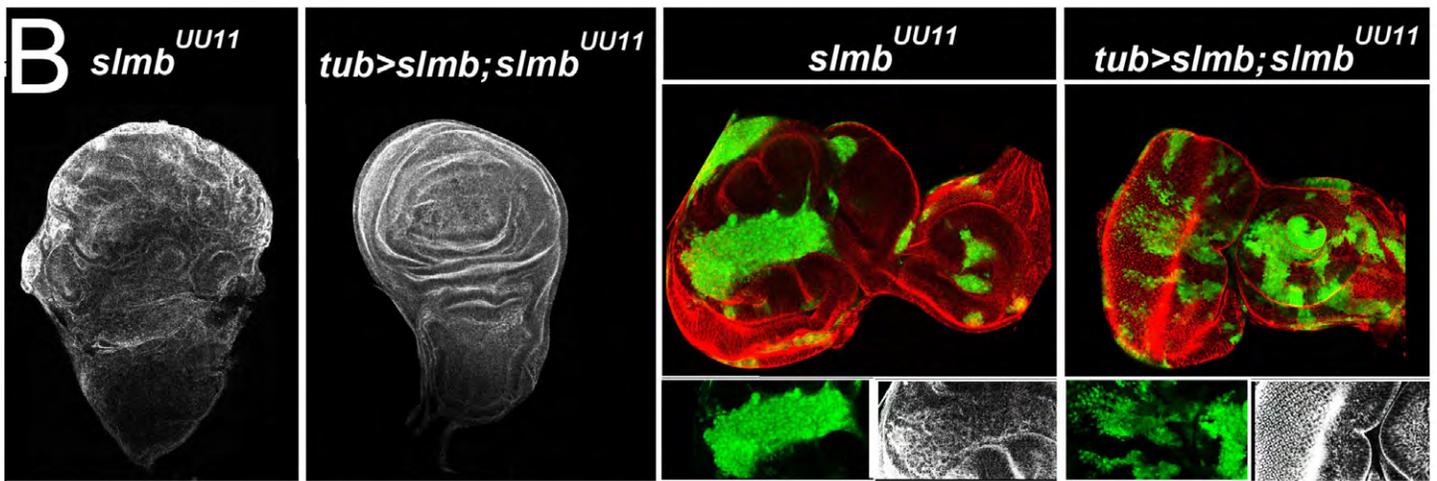
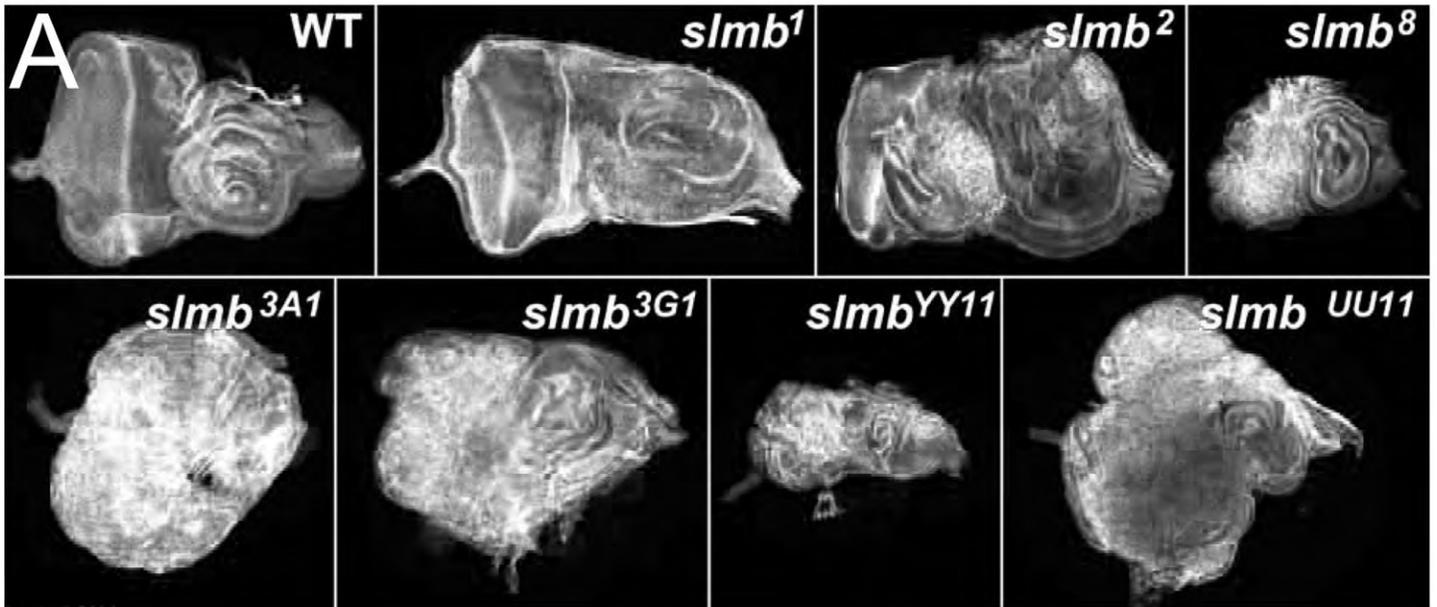
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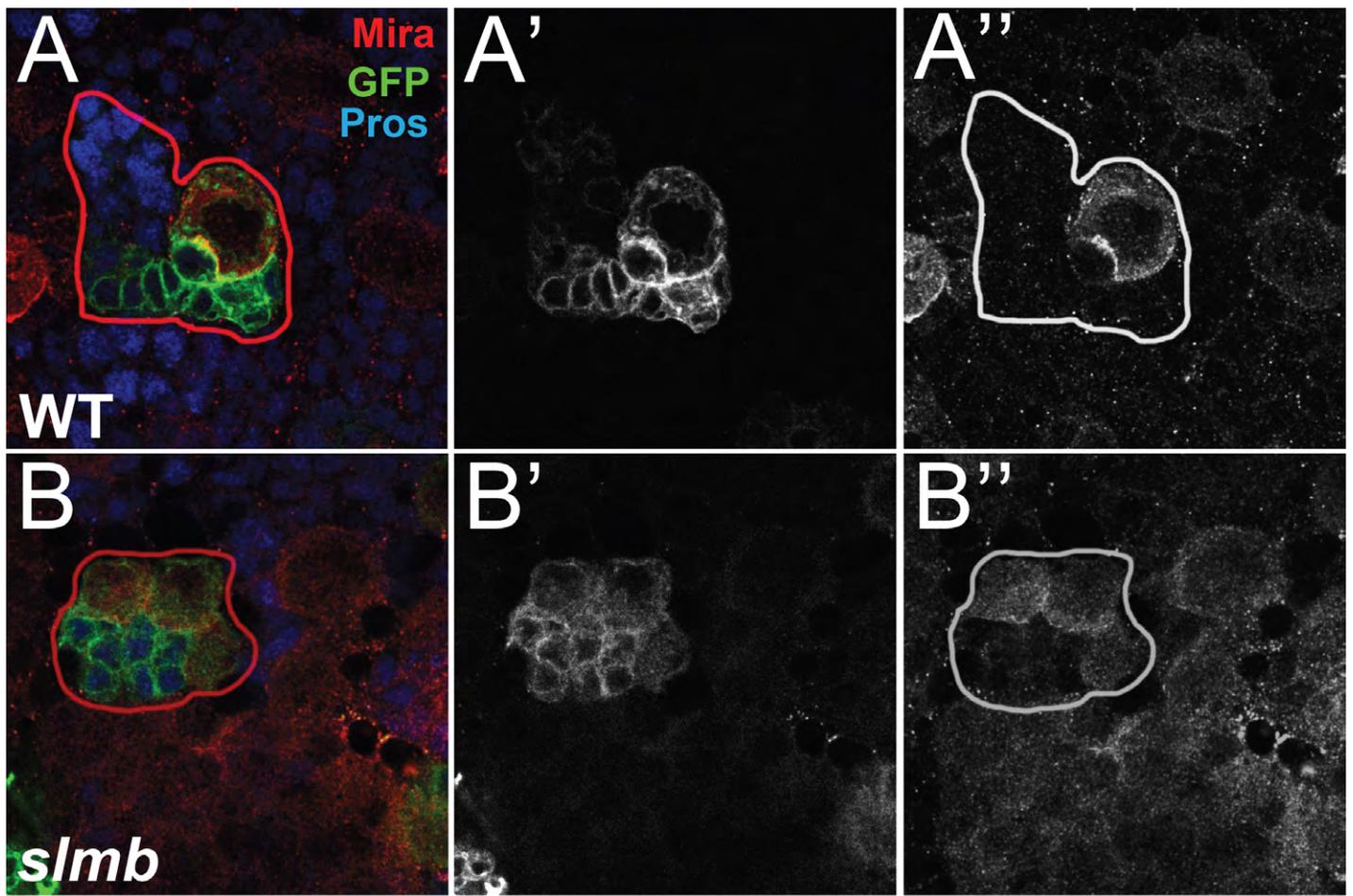
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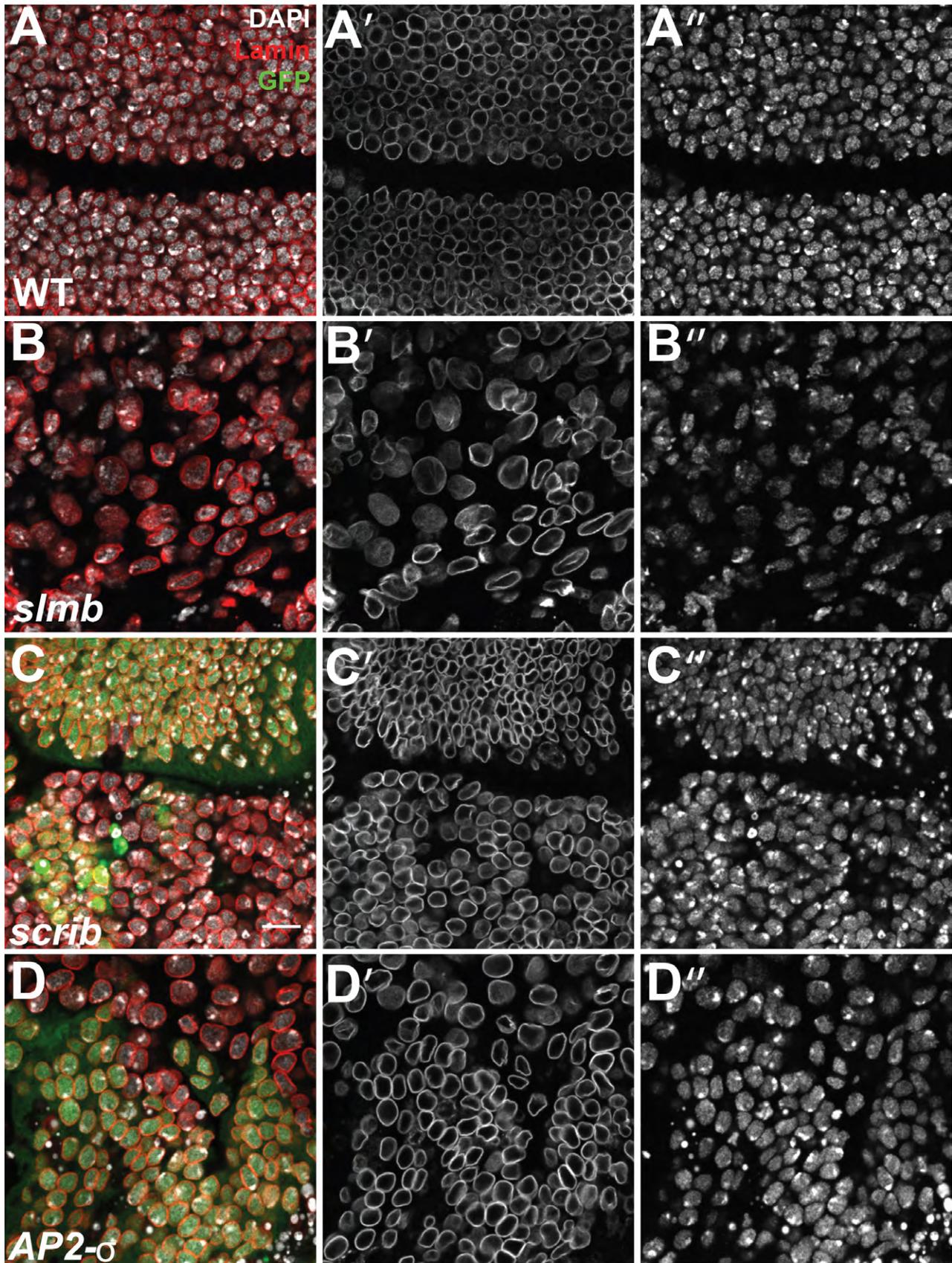
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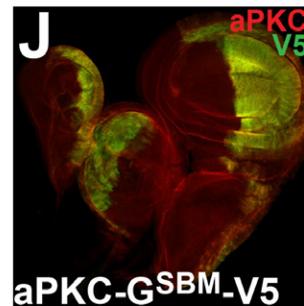
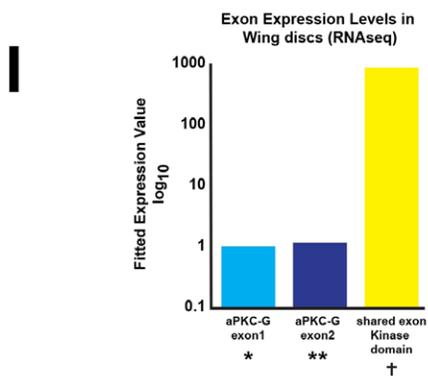
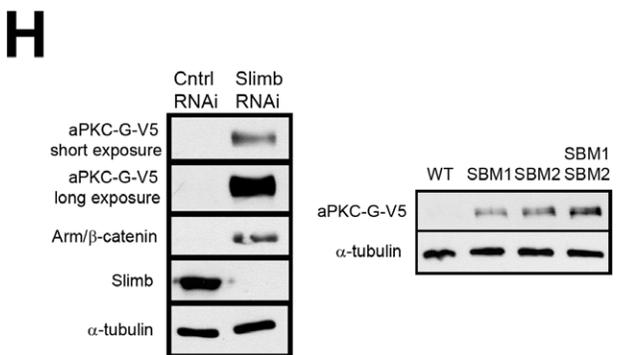
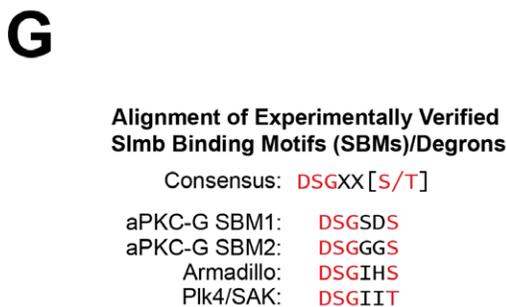
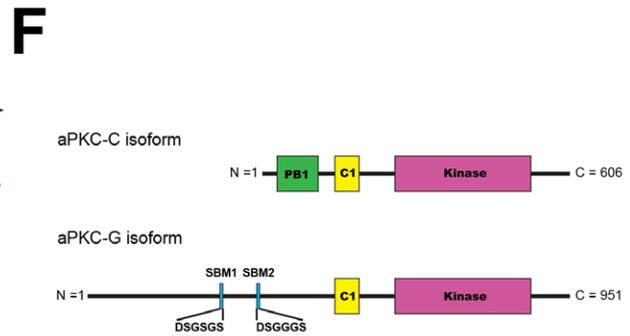
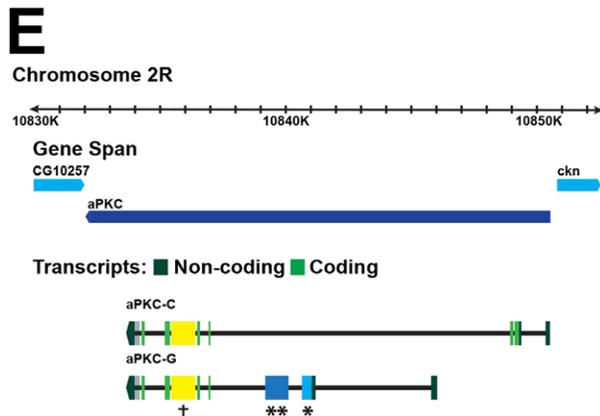
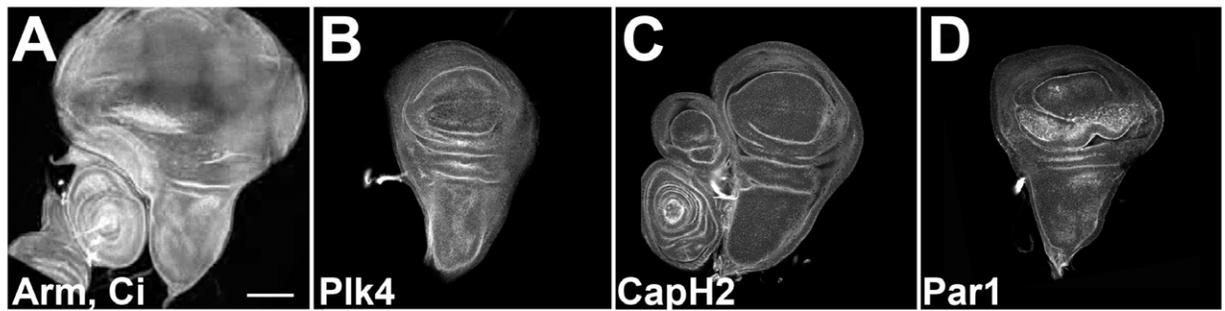
Supplementary Figure 1. Analysis of *slmb* allelic series. (A) Phalloidin staining of *slmb* mutant eye discs demonstrates that strong alleles show the most severe neoplastic transformation. (B) A *slmb* transgene rescues the neoplastic phenotypes of *UU11* in predominantly mutant wing discs and GFP-marked eye disc mosaics. (C) Discs derived from the deletion allele *slmb*⁸ and null mutation in the SCF core component *roc1a*^{G1} also display hallmarks of neoplasia, including disrupted F-Actin and upregulation of Mmp1.



Supplementary Figure 2. Effect of loss of *slmb* in neuroblasts. (A, B) GFP marks clones generated using the MARCM system. Larval type I neuroblasts divide asymmetrically to produce a new Miranda-positive neuroblast (red) and a smaller ganglion mother cell that will differentiate into a neuron or glia (Prospero positive, blue). *slmb* mutant neuroblasts display defects in asymmetric cell division, with a fraction of clones containing multiple Miranda positive neuroblast-like cells.



Supplementary Figure 3. Junctional scaffold and endocytic class tumor suppressors do not regulate Slmb activity. (A, B) Cells mutant for strong *slmb* alleles show chromosome condensation defects leading to a swollen nuclear lamina, reflecting misregulation of Condensin components. (C,D) In contrast, *scrib* and *AP2-sigma* mutant cells have WT nuclei and lamina size. Presence of GFP marks mutant cells. Scale, 10 μ m.



Supplementary Figure 4. Misregulation of known substrates cannot account for the *slmb* phenotype. (A-D) Overexpression of stabilized versions of known Slimb substrates throughout the presumptive wing pouch and notum using *MS-1096GAL4* does not phenocopy loss of *slmb*. (E, F) Gene and protein models comparing a common aPKC isoform C with the G isoform containing two Slimb binding motifs (SBM). (G) Alignment of aPKC-G SBMs with experimentally validated SBM degrons from other Slimb targets. (H) Western blots demonstrating that RNAi mediated knockdown of *slmb* in S2 cells results in stabilization of the aPKC-G isoform, as does mutation of the SBMs. (I) RNAseq data from third instar wing discs comparing levels of the unique aPKC-G exons with an exon encoding the shared Kinase domain; values shown are derived from RPKM. (J) Overexpression of a stabilized version of aPKC-G in the posterior domain of the wing disc (*en>GFP*, green) does not affect polarity or growth.