

Fig. S1

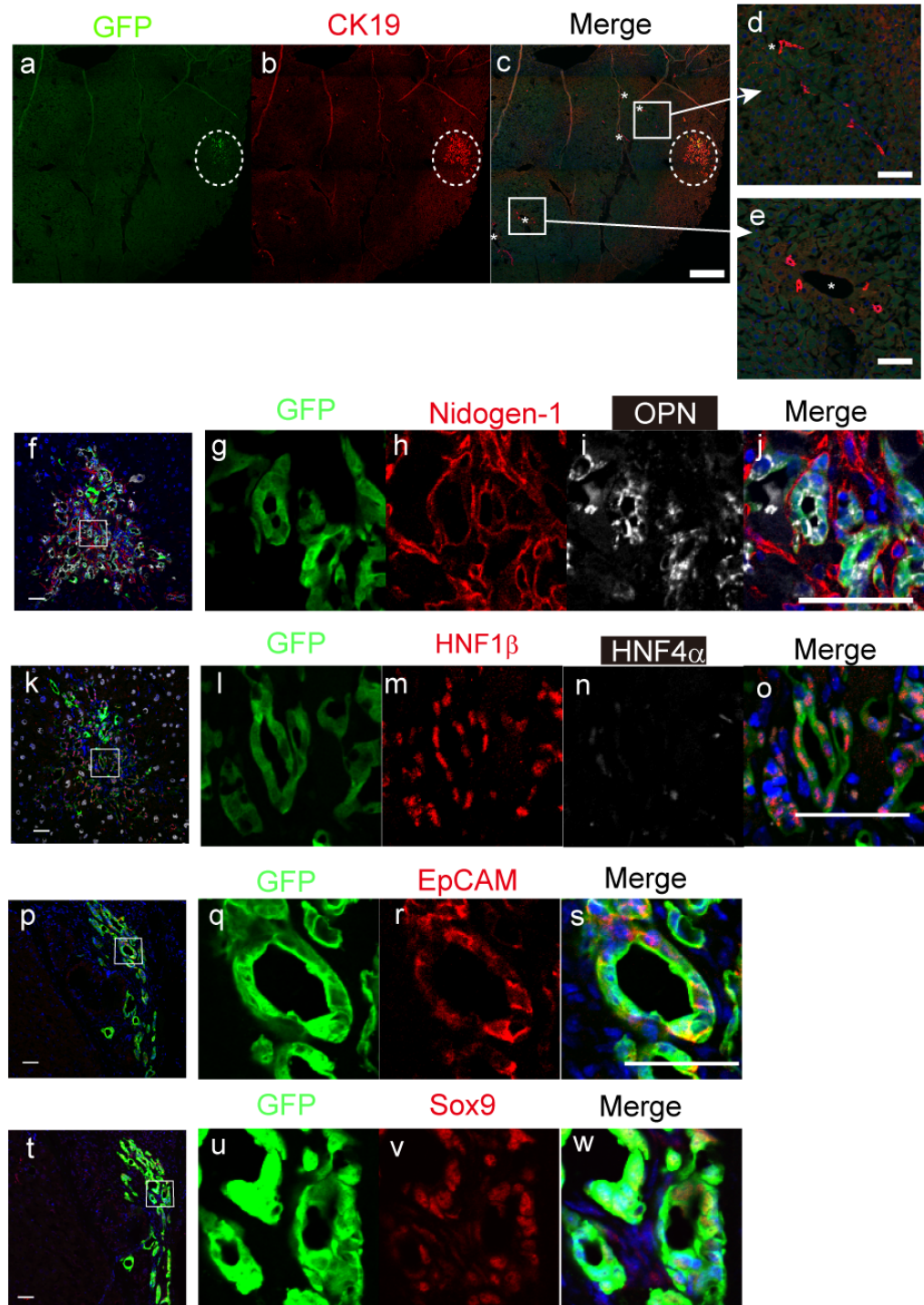


Fig. S1. Adult clone 1 differentiates to cholangiocyte-like cells and forms ductular structures in the recipient liver. GFP⁺ donor cells form ductular structures, surrounded by dotted circles, apart from portal veins (asterisks). Boxes in panel c are enlarged in panels d and e. Bars in panel c and d&e represent 500 and 100 μm, respectively. GFP⁺ donor cells express cholangiocyte markers including OPN, HNF1β, EpCAM and Sox9 but not a hepatocyte marker HNF4α. GFP⁺ ductular structures are associated with the ECM layer recognized with Nidogen-1. Bars in panels a, f, p, and t represent 100 μm. Bars in panels e, j, o, and s represent 50 μm.

Fig. S2

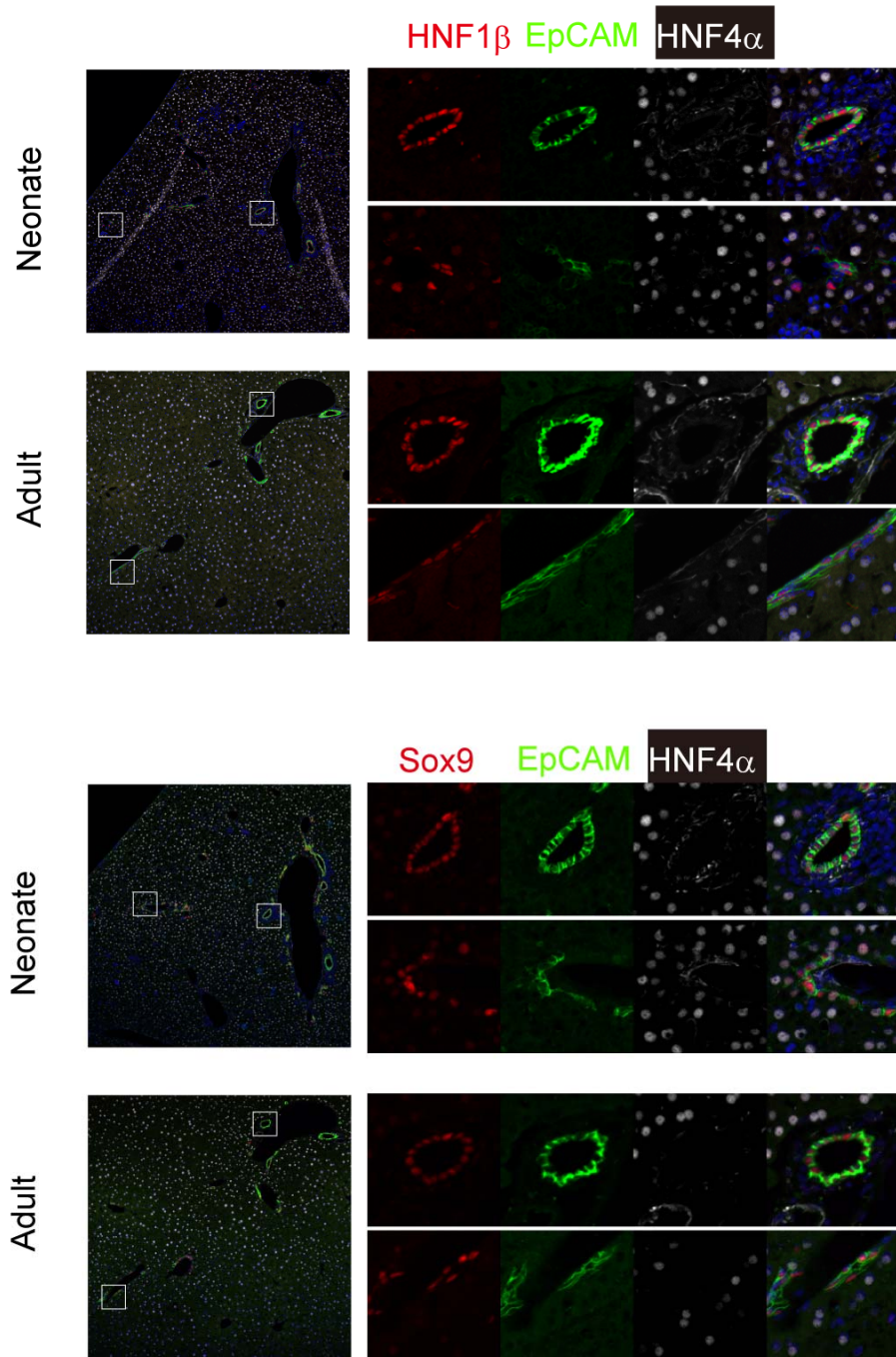


Fig. S2. Neonatal and adult EpCAM⁺ cells are positive for HNF1β and Sox9.

Fig. S3

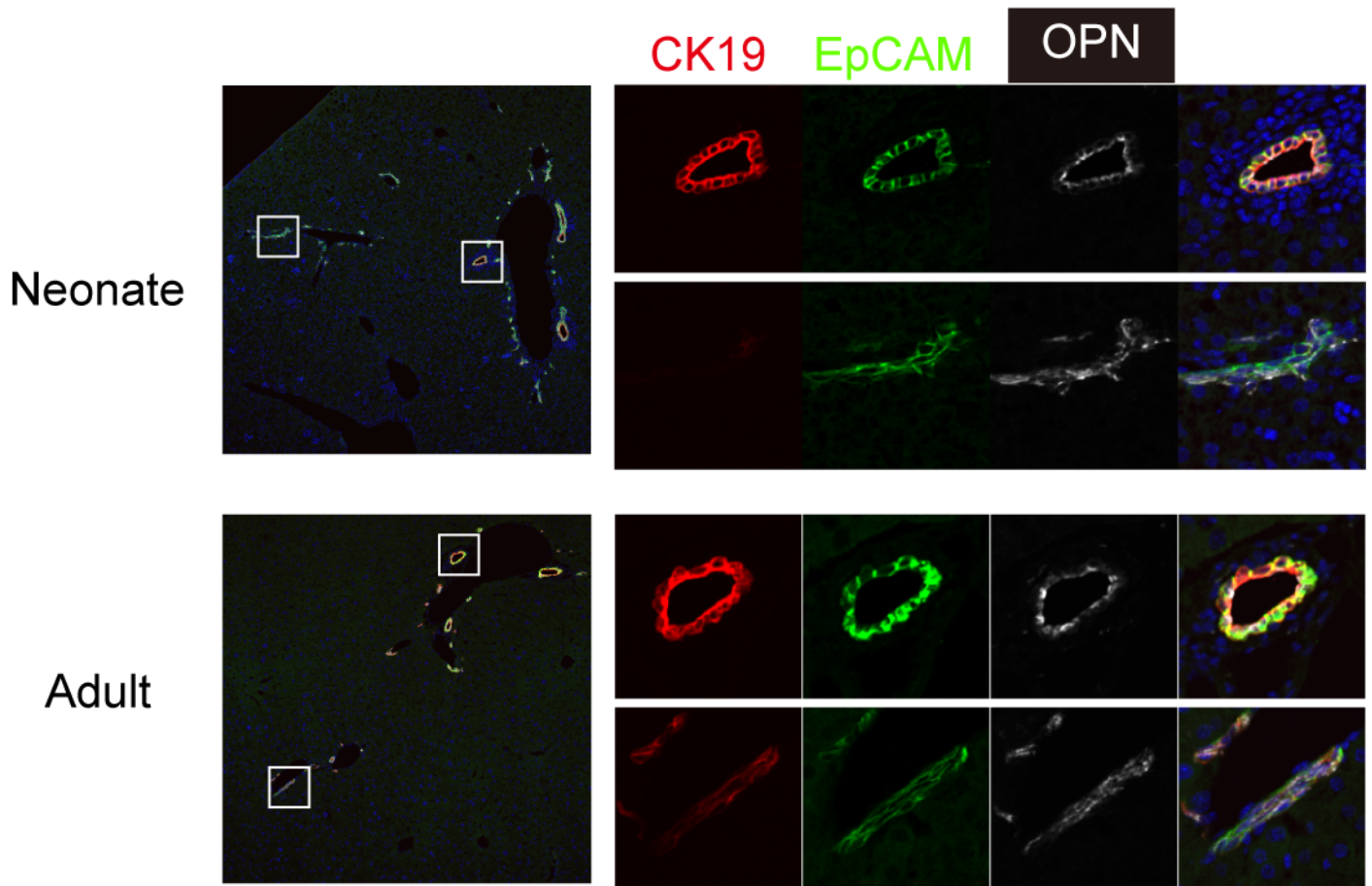


Fig. S3. Neonatal small ductules in the periphery are negative for CK19.

Fig. S4

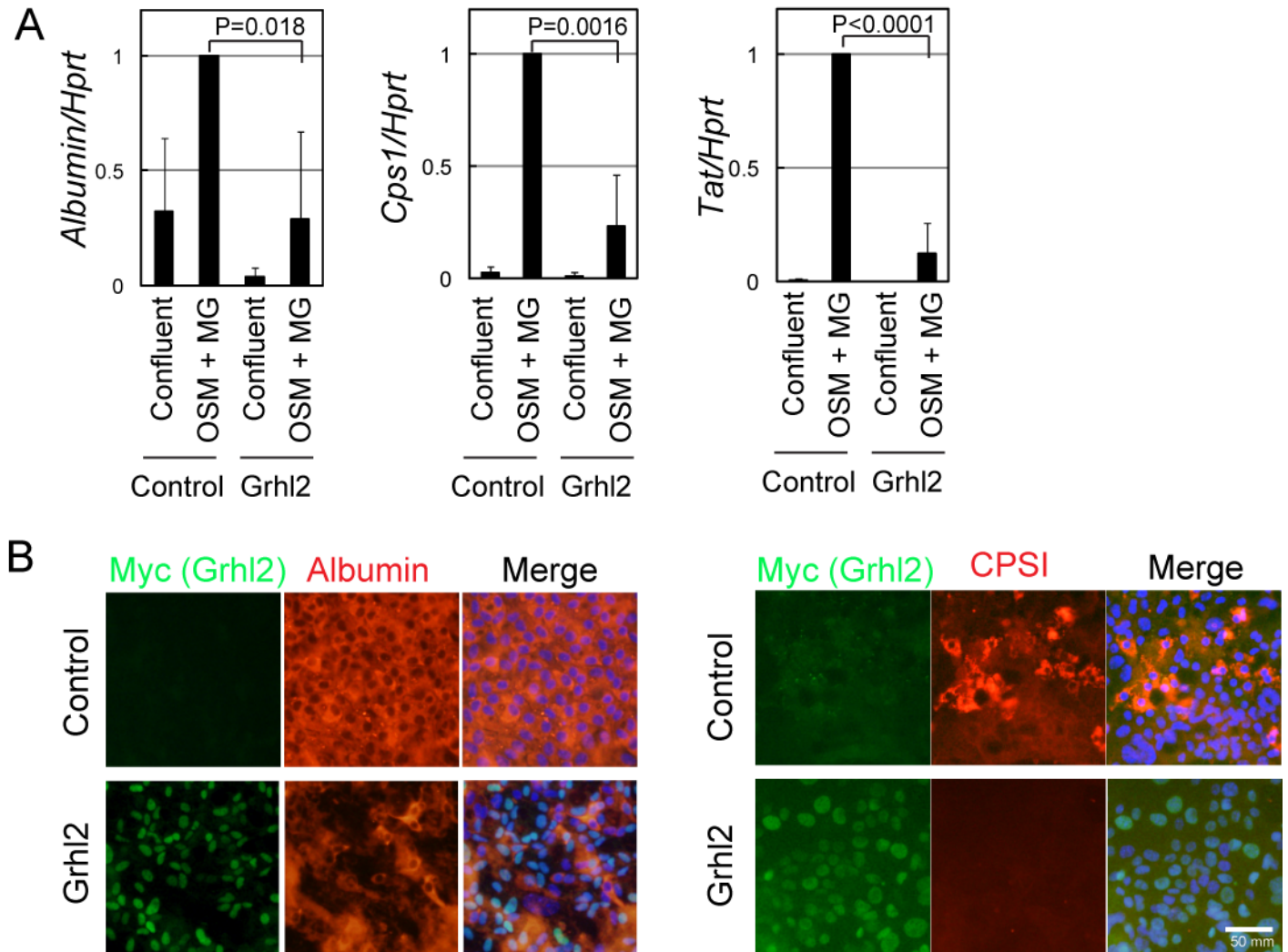


Fig. S4. Grhl2 inhibits hepatocytic differentiation of liver progenitors. HPPL, bipotential liver progenitor cell line, was introduced with Grhl2 by using a retrovirus vector. Hepatocytic differentiation was induced by oncostatin M (OSM) and Matrigel (MG). Grhl2 inhibits induction of *Albumin*, *Cps1*, and *Tat* (A) as well as ALBUMIN and CPSI (B). A bar represents 50 μ m.

Fig. S5.

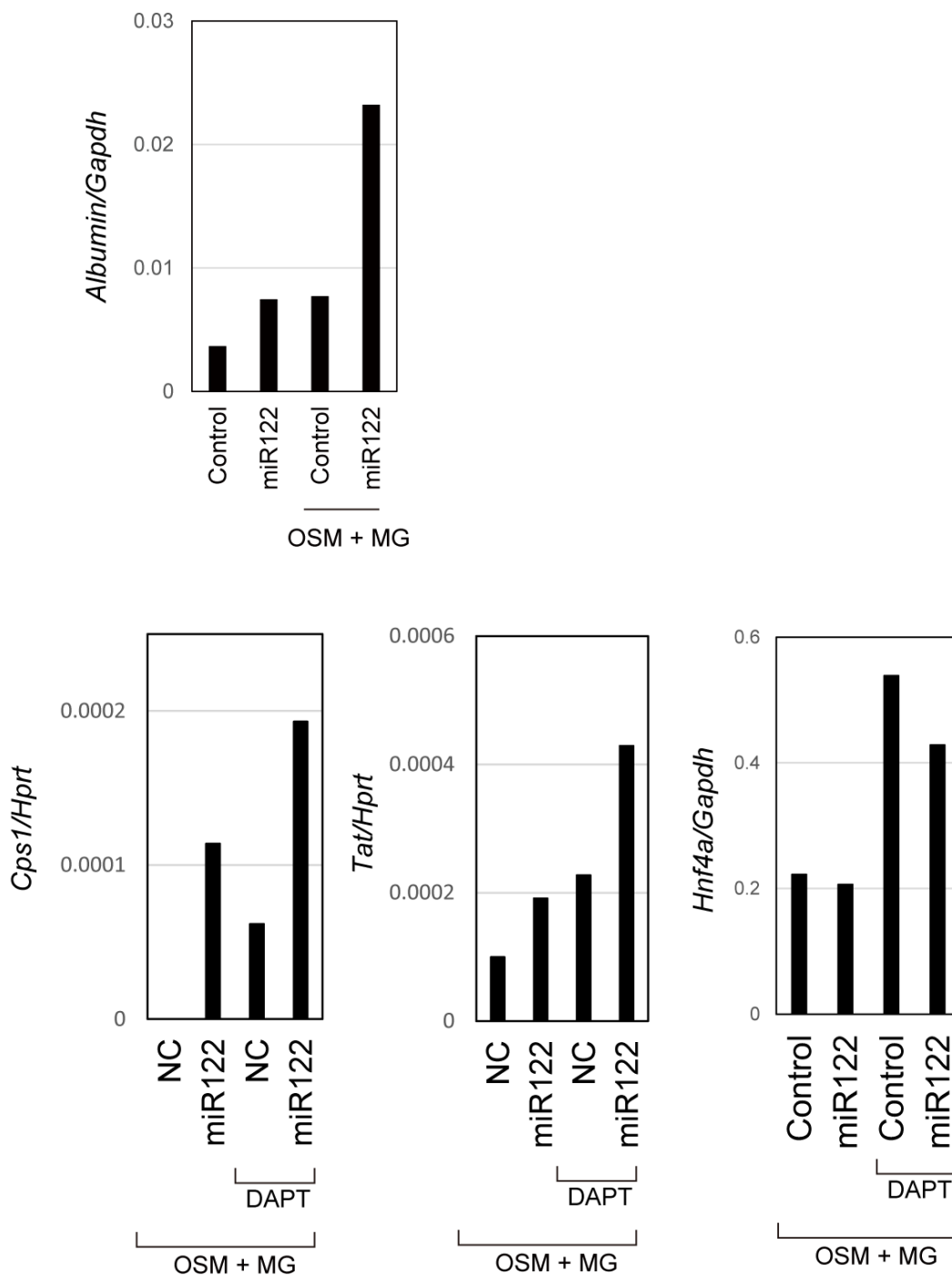


Fig. S5. miR122 promotes hepatocytic differentiation of adult EpCAM⁺ cells. miR122 slightly upregulates hepatocyte markers. EpCAM⁺ cells isolated from adult livers were cultured on gelatin coated dishes. They were transfected with miR122 and then treated with OSM and MG. For inducing *Cps1* and *Tat*, EpCAM⁺ cells were also treated with DAPT, a γ -secretase inhibitor, and potential inhibitor of the Notch signaling pathway.

Fig. S6

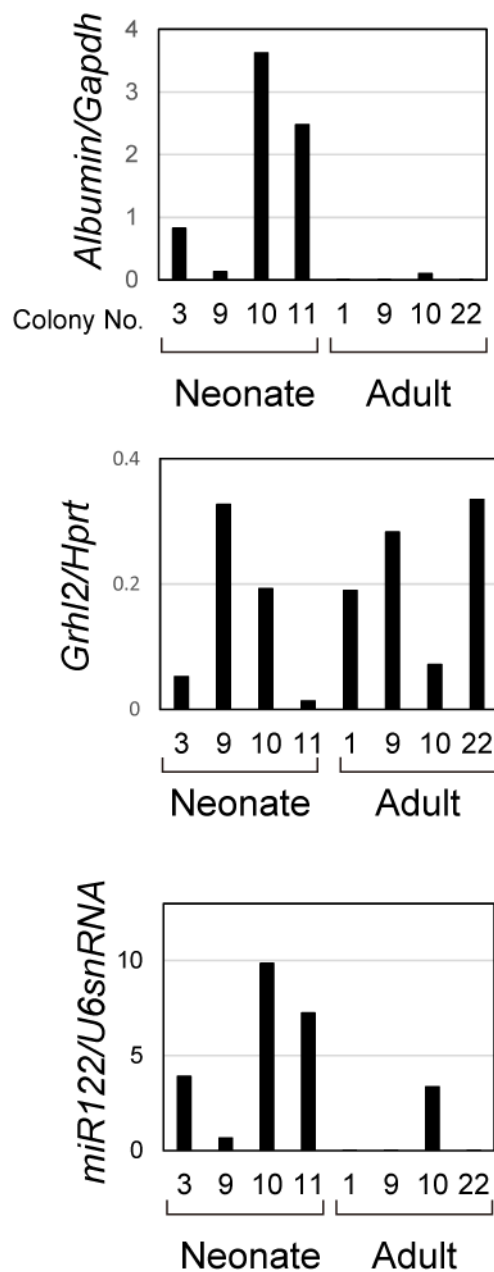


Fig. S6. Expression of Albumin, Grhl2, and miR122 in colonies derived from neonatal and adult EpCAM⁺ cells.

Table S1. Primary Antibodies

Antibody	Company	Cat. Number	Host animal	Method	Dilution
ALB	Bethyl laboratory	A90-234A	goat	IF	1:1000
CD16/32	BD Pharmingen	553141	rat	FACS	1:1000
CD31	Biologend	102501	rat	IF	1:1000
CD31 (APC-conjugated)	BD Pharmingen	551262	rat	FACS	1:1000
CD45 (APC-conjugated)	BD Pharmingen	559864	rat	FACS	1:1000
C/EBP α	SantaCruz Biotechnology Inc.	sc-61	rabbit	IF	1:200
CPSI	Santa Cruz Biotechnology Inc.	sc-10516	goat	IF	1:200
Cytokeratin 19	Tanimizu et al. 2003		rabbit	IF	1:2000
EpCAM	BD Pharmingen	552370	rat	IF	1:500
EpCAM (FITC or APC-conjugated)	Biologend	118208 118213	rat	FACS	1:1000
GFP	MBL	598	rabbit	IF	1:1000
GFP	Nakarai Tesk	GF090R	rat	IF	1:1000
GRHL2	Sigma-Aldrich	HPA004820	rabbit	IF	1:1000
HNF1 β	SantaCruz Biotechnology Inc.	sc-22840	rabbit	IF	1:200
HNF4 α	SantaCruz Biotechnology Inc.	sc-8987	rabbit	IF	1:200
HNF4 α	SantaCruz Biotechnology Inc.	sc-6557	goat	IF	1:200
Myc-tag	Millipore	05-074	mouse	IF	1:1000
Nidogen-1	Millipore	MAB1946	rat	IF	1:1000
Osteopontin	R&D systems	AF808	goat	IF	1:600
SOX9	Millipore	AB5535	rabbit	IF	1:3000
TER119 (APC/Cy7-conjugated)	Biologend	116223	rat	FACS	1:1000

Table S2. Primers used for PCR

Gene name		Sequence
Albumin	Sense	5'-GAA AGC CCA CTG TCT TAG TG-3'
	Antisense	5'-GGG TGT AGC GAA CTA GAA TG-3'
Cyp1a2	Sense	5'-CCCTGCCCTTCAGTGGTACA-3'
	Antisense	5'-AAGCTGTAGAGGTCTGGTCG-3'
Cyp2b10	Sense	5'-GTTGAGCCAACCTTCAAGGAA-3'
	Antisense	5'-AAGAGCTCAAACATCTGGCTG-3'
Cyp2d10	Sense	5'-GATCCCAAGGTGTGGTCCTT-3'
	Antisense	5'-GCAGGAGTATGGGGAACATA-3'
CPSI	Sense	5'-ACT GAG AGA TGC TGA CCC TA-3'
	Antisense	5'-CCT GGA AAT TGG TGA GGA GA-3'
GAPDH	Sense	5'-ACC ACA GTC CAT GCC ATC AC-3'
	Antisense	5'-TCC ACC ACC CTG TTG CTG TA-3'
G6pc	Sense	5'-CCA ACG TAT GGA TTC CGG TG-3'
	Antisense	5'-TCC CAG GTT TTT GAA GAG GC-3'
GFP	Sense	5'-CTGAAGTTCATCTGCACCAC-3'
	Antisense	5'-TTGAAGTTCACCTTGATGCC-3'
Pepck	Sense	TTGATGCCCAAGGCAACTTA
	Antisense	ACGGCCACCAAAGATGATAC
TAT	Sense	5'-GAG GAG TGT GAC AAA TAG GC-3'
	Antisense	5'-AGA GGA CAC TCC TGT GTC AG-3'
Tdo2	Sense	5'-TGAGTAAAGGTGAACGACGAC-3'
	Antisense	5'-AGCCGACTGAGAATCCTGTA-3'

Table S3.

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