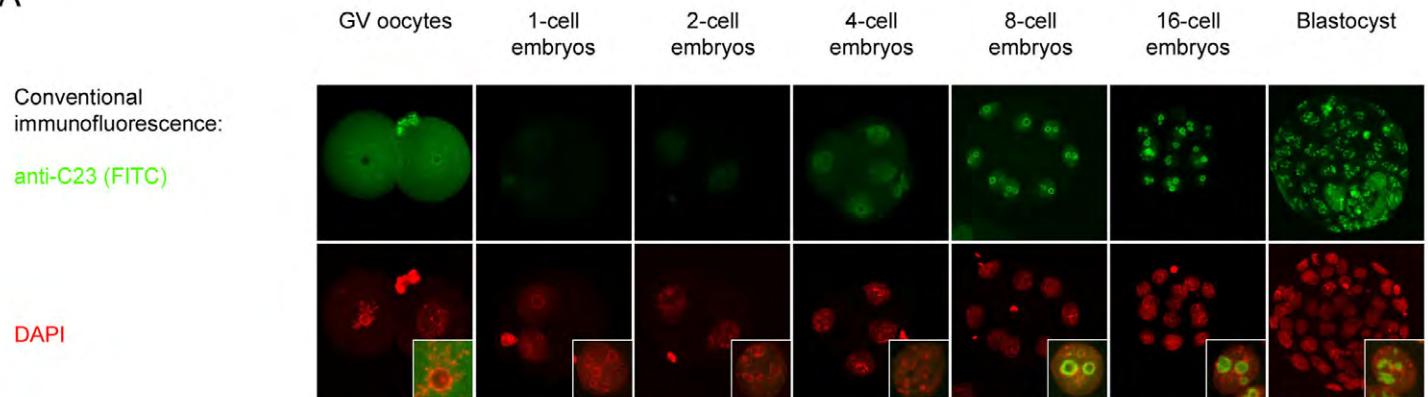


A



B

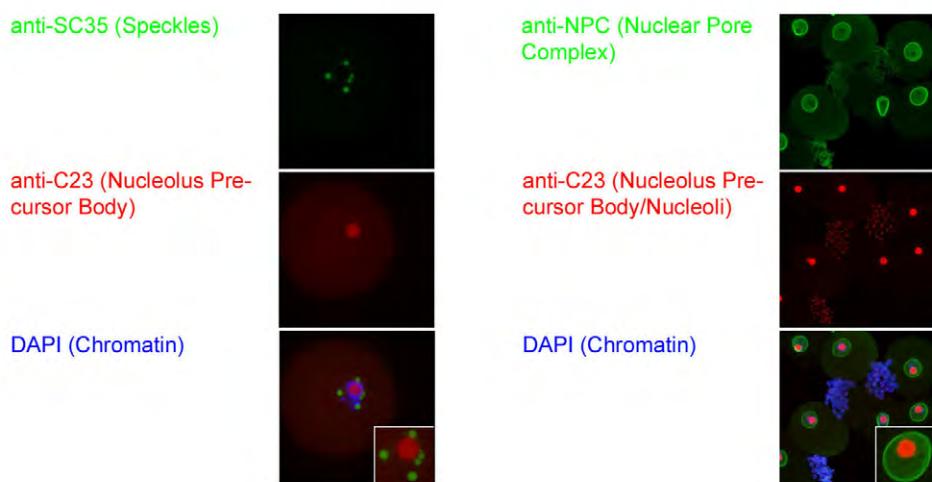


Figure S1. The comparison of NPB labelling strategies by conventional immunofluorescence protocols and antigen retrieval/immunofluorescence protocol. (A) The use of conventional protocols does not allow staining of the inside of NPBs as antibodies are unable to penetrate the dense structure. Typically, positive staining of the nucleolus is obtained from the time of embryonic genome activation onwards (2- to 4-cell embryos). (B) The use of antigen retrieval combined with immunofluorescence results in NPB labelling even in GV stage oocytes. Moreover, this protocol does not cause unspecific reactivity of the NPB to other antibodies (e.g. anti-SC35 or anti-NPC).

Table S1. Primers used for real-time PCR and SNP analysis

Primer	Sequence	Concentration	Location
rRNA_F	GGTGTCCAAGTGTTCATGC	600 nM	5'external transcribed spacer
rRNA_R	CAAGCGAGATAGGAATGTCTTAC	900 nM	5'external transcribed spacer
5ETS_F1	TGTTTCACTTGGTCTGTCTC	600 nM	5'external transcribed spacer
5ETS_R1	TCGACGCCCTACAAGAACAGC	600 nM	5'external transcribed spacer
5ETS_F2	GTCTTCTGGTTCCCTGTGTG	900 nM	5'external transcribed spacer
5ETS_R2	GCTAGAGAAGGAAACTTCTCACTG	600 nM	5'external transcribed spacer
ITS1_F	TCTCGTTCGTCCTGCTGG	900 nM	internal transcribed spacer 1/5.8S rRNA
ITS1_R	GATCCACCGCTAAGAGTCGTATC	900 nM	internal transcribed spacer 1/5.8S rRNA
ITS2_F1	GAGAACGGAGAGAGGTGGTATC	300 nM	internal transcribed spacer 2
ITS2_R1	AGAACCGGAGACGAAGAAGAG	300 nM	internal transcribed spacer 2
ITS2_F2	CGTGTGAGTAAGATCCTCCAC	900 nM	internal transcribed spacer 2/28S rRNA
ITS2_R2	GTTACTGAGGGAATCCTGGTTAG	600 nM	internal transcribed spacer 2/28S rRNA
EGFP_F	CACCATCTTCTCAAGGACG	900 nM	
EGFP_R	GTGGCTGTTGTAGTTGACTC	300 nM	
C23_SNP_F	CACACCAGCTAAGAAAAACATTACAC	300 nM	Exon 2/3
C23_SNP_R	ACTCATCCTCTTCCTCATCATCTTC	300 nM	Exon 2/3
UBF_SNP_F	GAGAAGAAGAAGGCTAAATACAAGG	300 nM	Exon 10/11
UBF_SNP_R	TCACCCGGTCATTCTTGAAG	300 nM	Exon 10/11