

Supplemental figures Legends, tables and figures

Xu et al.,

Maternal BCAS2 protects genomic integrity through RPA in mouse preimplantation embryos

Qianhua Xu^{1,2,7}, Fengchao Wang^{3,7}, Yunlong Xiang¹, Xiaoxin Zhang¹, Zhenao Zhao¹, Zheng Gao^{1,2}, Wenbo Liu¹, Xukun Lu^{1,2}, Yusheng Liu¹, Xing-jiang Yu¹, Haibin Wang¹, Jun Huang⁴, Zhaohong Yi⁵, Shaorong Gao^{6,8}, Lei Li^{1,8}

¹State Key Laboratory of Stem Cell and Reproductive Biology, Institute of Zoology, Chinese Academy of Sciences, Beijing, 100101, China. ²University of Chinese Academy of Sciences, Beijing, 100049, China. ³National Institute of Biological Sciences, Beijing, 102206, China. ⁴Life Sciences Institute, Zhejiang University, Hangzhou, Zhejiang, 310058, China. ⁵College of Biological Science and Engineering, Beijing University of Agriculture, Beijing, 102206, China. ⁶School of Life Sciences and Technology, Tongji University, Shanghai, 200092, China.

Figure S1

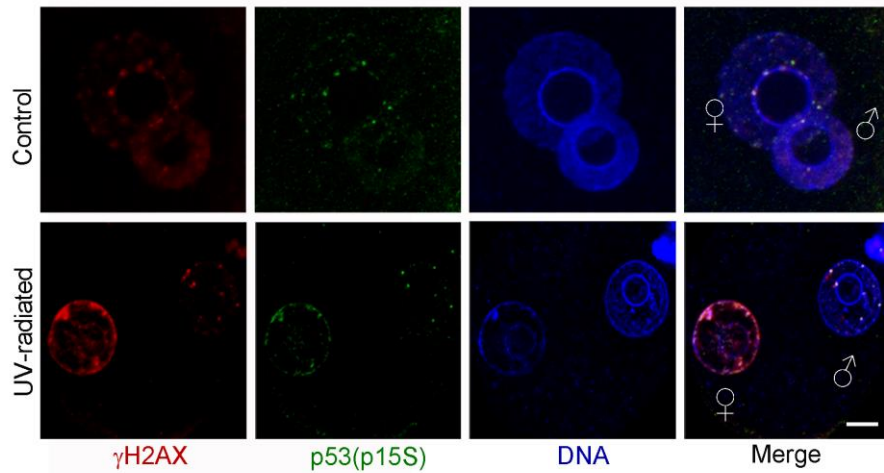


Fig. S1. BCAS2 responses to UV-induced DNA damage. MII oocytes were UV-irradiated and *in vitro* fertilized with normal sperms. Zygotes were cultured until PN4-5 and immunostained with phosphor-p53 (p15S) and γ H2AX. Male- and female pronuclei are indicated with male and female symbols, respectively. Scale bar, 10 μ m.

Figure S2

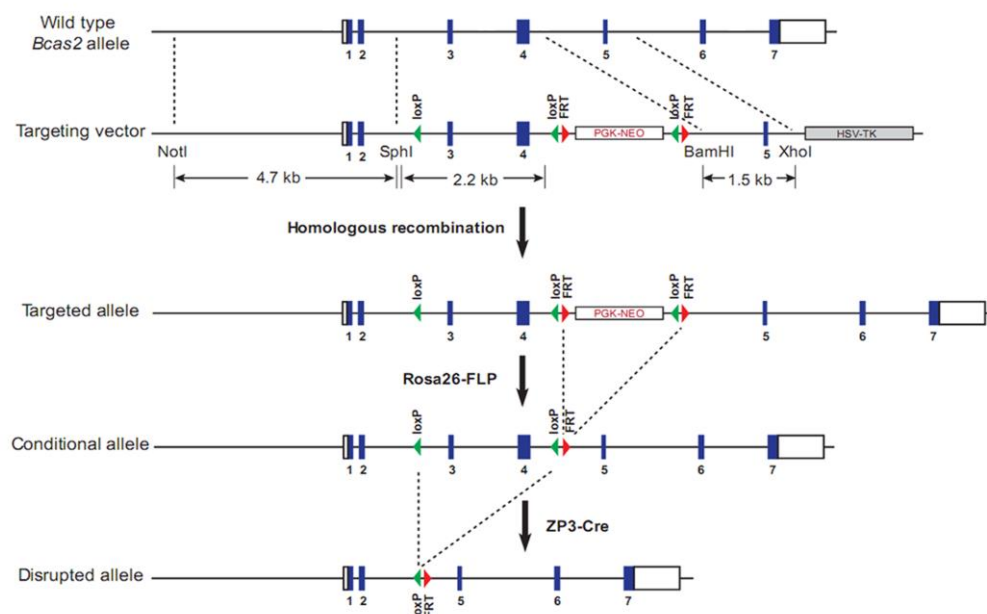


Fig. S2. *Bcas2* targeting strategy and analysis of F2 offspring. A targeting vector was constructed by anchoring LoxP sites around exon 3 and exon 4. HSV-TK expression was used for negative selection, and PGK-NEO expression was used for positive selection. Targeted ES clones were confirmed by sequencing and injected into blastocysts to produce chimeric mice.

Figure S3

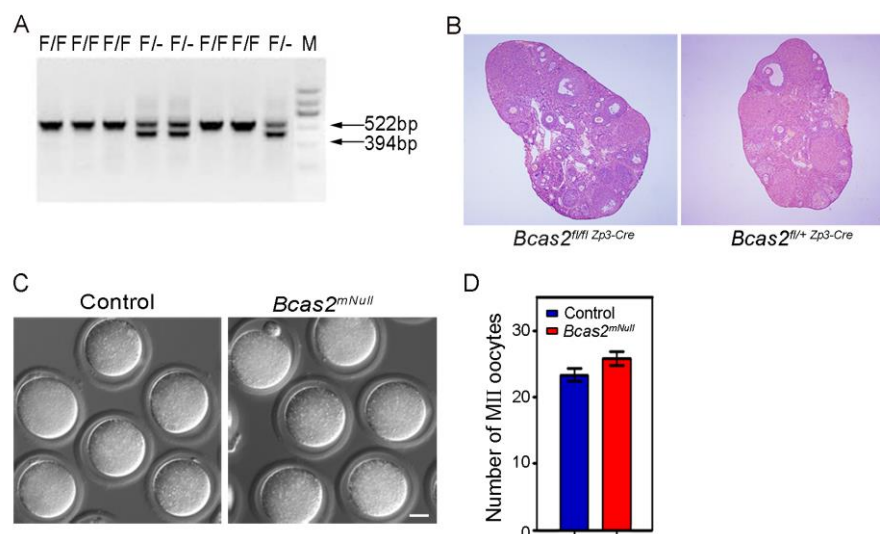


Fig. S3. Characterization of *Bcas2*^{fl/fl} and *Bcas2*^{fl/fl};Zp3-Cre mice. (A) PCR analysis of genomic DNA derived from eight offspring. (B) Ovaries from two-month old *Bcas2*^{fl/+};Zp3-Cre and *Bcas2*^{fl/fl};Zp3-Cre females were fixed and stained with hematoxylin and eosin. Scale bar, 50 μ m. (C and D) MII oocytes were obtained from *Bcas2*^{fl/+};Zp3-Cre and *Bcas2*^{fl/fl};Zp3-Cre females after 13 hours post-hCG. Scale bar, 20 μ m.

Figure S4

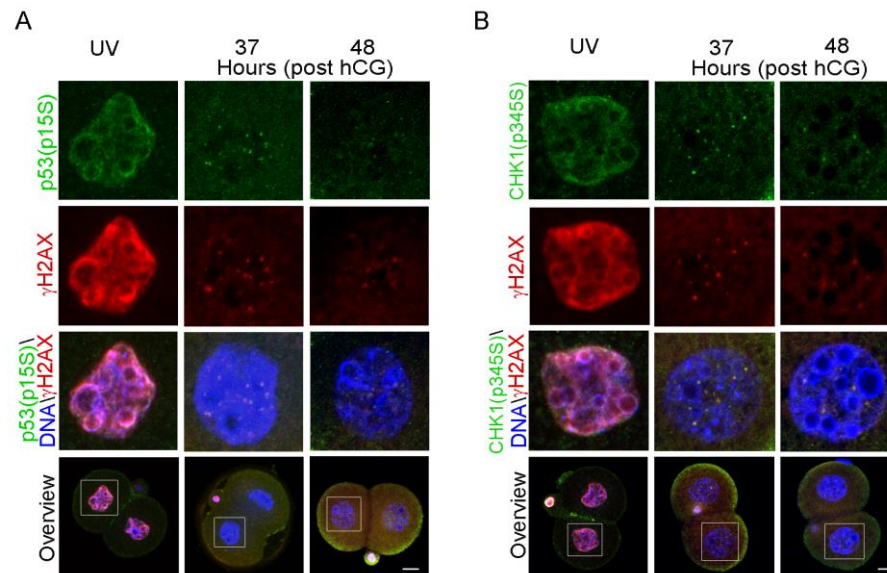


Fig. S4. Checkpoint activation during cell cycle progression in normal two-cell embryos. (A) Normal two-cell embryos at 37 and 48 hrs post-hCG were fixed and stained with γ H2AX and phosphor-p53 (S15) antibodies under TP conditions. (B) Normal two-cell embryos at 37 and 48 hours post-hCG were fixed and stained with γ H2AX and phosphor-pCHK1(S345) antibodies under TP conditions. Normal two-cell embryos irradiated by UV were positive control. Scale bar, 20 μ m.

Figure S5

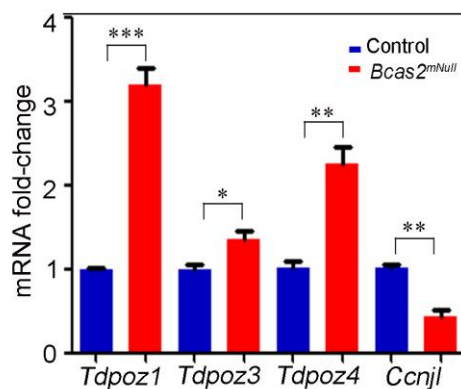


Fig. S5. Zygotic genome active is partially impaired in *Bcas2^{mNull}* two-cell embryos. Levels of *Tdpoz1*, *Tdpoz3*, *Tdpoz4*, *Ccnjl* and *Gm13043* were measured by qRT-PCR with specific primers (Table S 4) in two-cell embryos obtained at 48 hrs post-hCG. The error bars represent the SEM from three independent experiments. *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$.

Figure S6

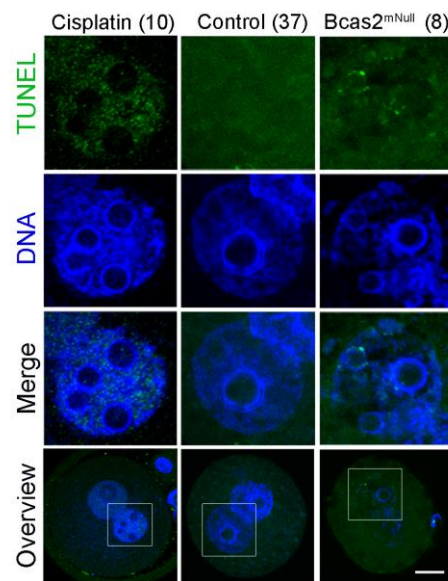


Fig. S6. BCAS2 is required for DNA repair in mouse zygotes. Control and BCAS2^{mNull} zygotes were obtained 30 hrs post-hCG and examined by TUNEL assay. Normal zygotes treated with Casplatin as the positive control.

Figure S7

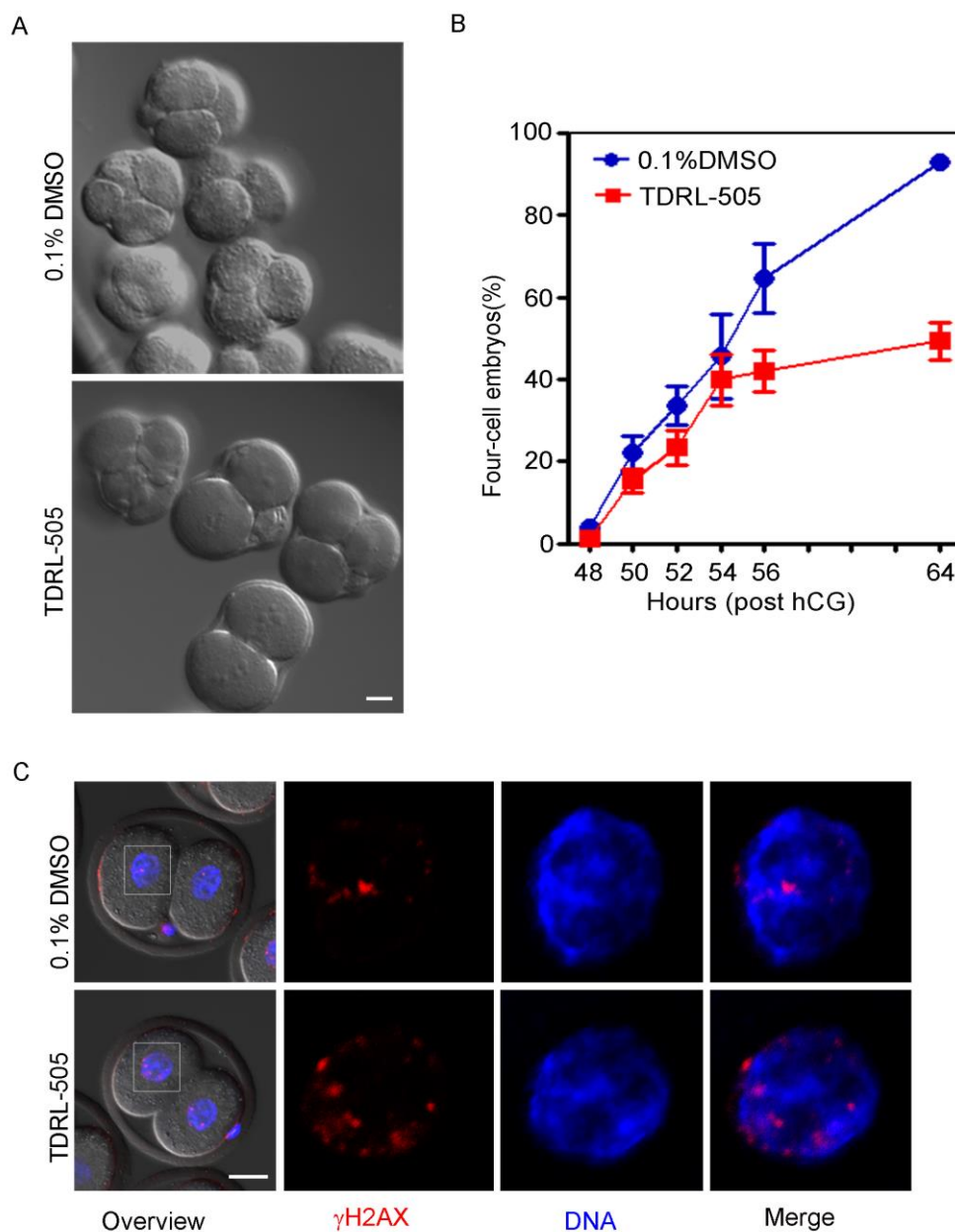


Fig. S7. RPA inhibition induces developmental arrest and increased γ H2AX at two-cell stage. (A) Normal zygotes were recovered 23 hrs post-hCG. Zygotes were treated with either 0.1% DMSO as control (n=65) or 100 μ M TDRL-505 (n=68) for 5 hrs and released. Zygotes were allowed to progress to the four-cell stage. (B) Quantification of (A) developmental rate

during the transition from the two- to four-cell stage. Developmental rates were calculated without the amount of arrested two-cell embryos. Error bars represented SEM. (C) Two-cell embryos from (B) were fixed after 48 hours post-hCG and immunostained with γ H2AX. Scale bar, 20 μ m.

Supplementary Table S1

PCR primers used for gene targeting at Bcas2 locus and genotyping.

Primer	5'→3'
SphI-Loxp-F	ACAGCATGCATAACTTCGTATAGCATACATTATACGAAG TTATGTTTTTAAGTGGTTATTCTACAAGTGC
NotI-R	GAATTCGCGGCCGCTGTAGTTCTGAGACAATCCC
Not-F	GAATTCGCGGCCGCGGTGCCAACGACAGTGTTTTTC
SphI-R:	ACAGCATGCAGTTAGACATGCTTGTTTCAGGC
BamHI-F:	TCGGATCCCAACGAAAGGCTACCTTGAG
XhoI-R:	TCCTCGAGCTAACTGTGAAGGTGTGTCAG
Neo-F:	TATCGCCTTCTTGACGAGTTC
screen-R	TTGGTCCTGCAGTCCAAATC
F-WT	ATTCCAGCAGTTGGTGTGGG
F2-KO	AGGTGTATGAATGCCTGAACAAG
R	CATTGCTGGACAGAAGGTGAG

Supplementary Table S2

Antibodies used in this study:

Name	Dilution	Application
anti-BCAS2 (Protein Tech Group, 10414)	IF	1: 200
	WB	1:600
anti-phosphor-p53 on Ser15 (Cell Signaling, 9284)	IF	1:100
	WB	1:1000
anti- γ H2AX antibody (Cell signaling, 22551)	IF	1: 500
	WB	1:1000
phosphor-RPA2 (Bethyl Laboratories, A300-245A)	IF	1:100
anti- β -Actin monoclonal antibody (ZSGB-BIO, ZM0002)	WB	1: 5000
Alexa 488 donkey anti-rabbit (Jackson, 711545152)	IF	1:1000
Alexa 633 donkey anti-mouse (Jackson, A-21050)	IF	1:200
HRP-conjugated anti-mouse (ZSGB-BIO; 2304)	WB	1:5000
HRP-conjugated anti-rabbit secondary antibodies (ZSGB-BIO; 2301)	WB	1:5000

Abbreviation: IF, immunofluorescence staining; WB, western blotting.

SupplementaryTable S3

Primer sequences for *Bcas2* and *Rpa2* mutations:

Gene	Forward (5'→3')	Reverse (5'→3')
<i>Bcas2</i>	TAGAATTCGCCACCATGG GGGCACGGGCTTGGTAG CCGGAGAGG	TACCGCGGGAAGTCTTGG CGGATGTTTTCTTGTG GCCTCCC
ΔN	GAATTCGCCACCATGGCG GCACGGGCTTGGT	TTGACCGCGGTCATTCTC TCAGCTTAGATC
<i>Rpa2</i>	TGCGGATATCATGTGGAA TAGCGGATTCGA	AGGAGGCGCGCCTCACT CTGCATCTGTAGACT
<i>Rpa2</i> (S4A/S8A)	AGGTGGAGCTGCTGAAG CCTTCGAATCCGCCATTC CACAT	ATGTGGAATGGCGGATTC GAAGGCTTCAGCAGCTCC ACCT
<i>Rpa2</i> (S33A)	CTCGATTTCTTCTCCGCC TGCGCCGGTGTGGGCGA CCCGA	TCGGGTCGCCCACACCG GCGCAGGCGGAGAAGAA ATCGAG
<i>Rpa2</i> (Thr21A)	CCGAAGCCACCGGGGGA CTGCGCGTAGCCGCCTG CTCCGCC	GGCGGAGCAGGCGGCTA CGCGCAGTCCCCCGGTG GCTTCGG

Supplementary Table S4

Primers for qRT-PCR assay:

Gene	Forward (5'→3')	Reverse (5'→3')
<i>Bcas2</i>	5'TCGCTGCTCGACAACCG ATTGAA3'	5'AGCTGCATGTTCTTTCGCTG CCA3'
<i>P21</i>	5'GCAGCCGAGAGGTGTG AGCC3'	5'CGGGACCGAAGAGACAAC GGC3'
<i>Gadd45a</i>	CTGCAGAGCAGAAGACCG AA	TACACGCCGACCGTAATGG
<i>Tdpoz1</i>	TCAGAGAAGGATTACAAG CCCA	GGCTGAGCAAACTAGGTAA ACT
<i>Tdpoz3</i>	CCTGTCAGTTTATCTGGAG TTGC	CAGAAAGCATTGGACATTGG AGA
<i>Tdpoz4</i>	GCCCAAGTGCTAACACCA GA	TCCCACAGCTCCCCTACGT
<i>Ccnj1</i>	TGGCATATCGGGACTCGT TG	ATATCCAAGGGCTGGAGGGT
<i>Gm13043</i>	GCCCCACTCTATCTGTTTT GC	AACGAATCCTCCTCCTATTTA CTG