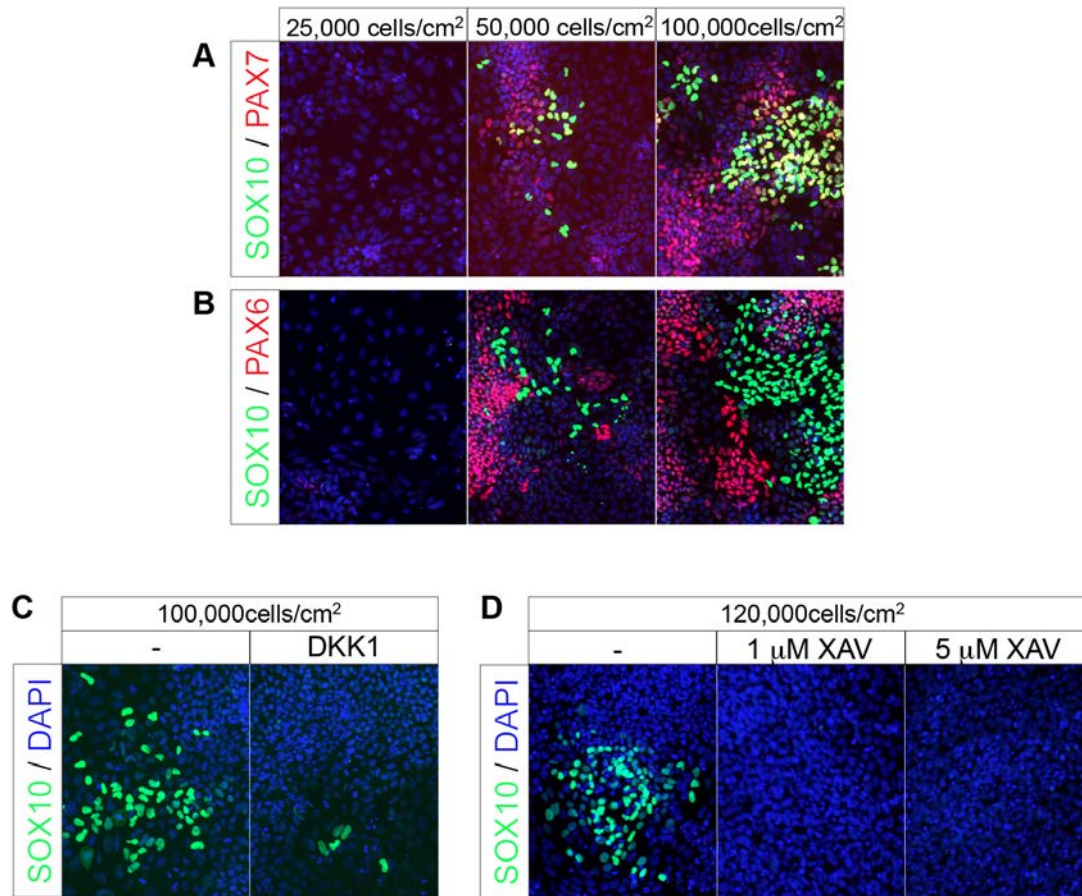
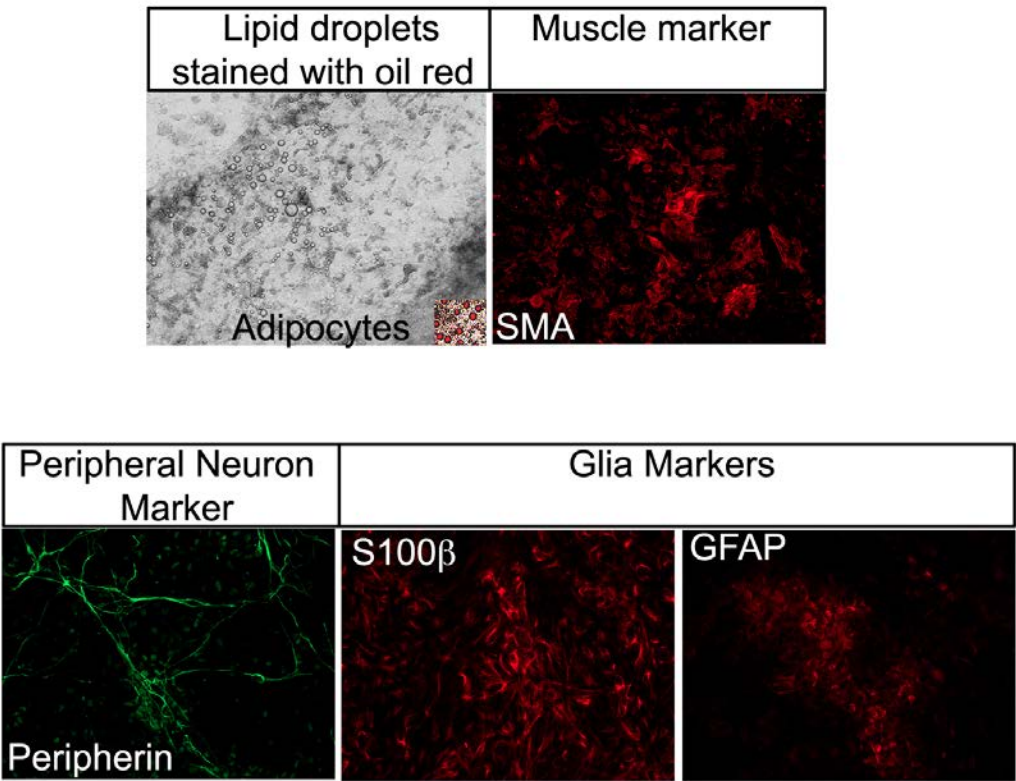


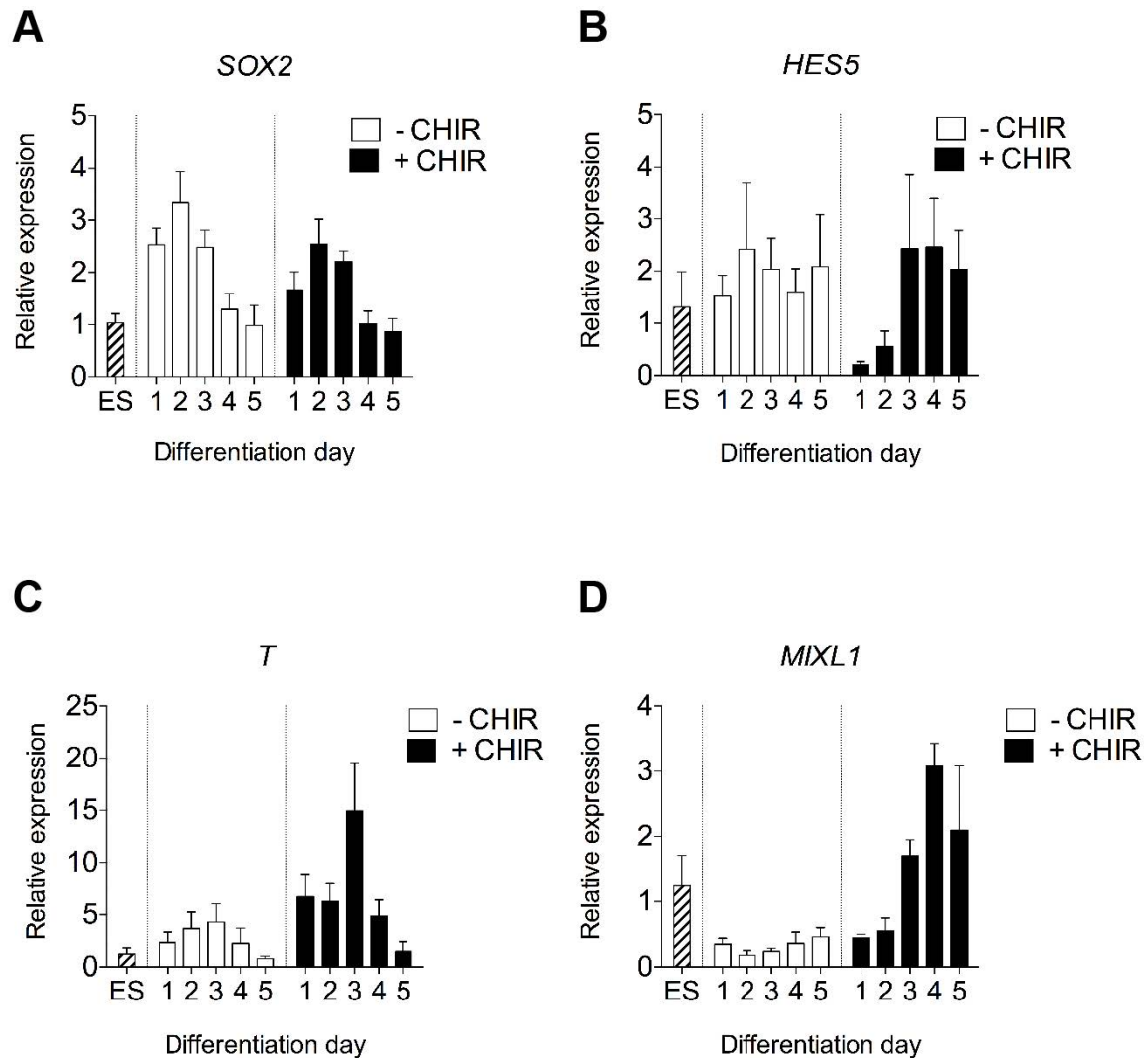
**Supplementary figure 1. CHIR 99021 induces neural crest cells in human Y6 iPSCs.** SOX10 (green), PAX7 (red) and TFAP2 (violet) triple immunostaining of iPSC RIV9 cultured after dissociation under control (No CHIR) or with CHIR for 5 days. SOX10 staining of day 5 Y6 iPSC cultures treated continuously with 3  $\mu$ M CHIR and with or without 300 ng/mL NOGGIN from day 0 to day 1.



**Supplementary figure 2. Spontaneous induction of SOX10<sup>+</sup>, PAX7<sup>+</sup> and PAX6<sup>+</sup> cells in high-density cultures.** A, B. Dissociated hESCs were plated at the indicated densities and cultured for 5 days in the base medium for NC induction without the administration of CHIR. SOX10/PAX7 (A) and SOX10/PAX6 (B) double labeling were performed. C. Day 5 cultures at the indicated initial seeding density were treated with or without 100 ng/mL DKK1 for 5 days and were stained with SOX10 antibody. D. Differentiating hESCs cultured in base differentiation medium without CHIR were treated with or without XAV 939 (XAV) at the indicated concentrations. Day 5 cultures were collected for SOX10 immunofluorescence.



**Supplementary figure 3. Terminal neural crest derivatives derived from WNT3a induced neural crest cells.** Dissociated hESCs were induced for 5 days in Wnt3a (10 ng/ml) prior to being assayed for their ability to produce neural crest derivatives. Assays were specific to the derivatives of interest.



**Supplementary figure 4. Expression of neural and mesoderm associated transcripts during human NC induction.** A-D. Time course real time PCR assays of *SOX2*, *HES5*, *T* and *MIXL1* during human NC induction.

Supplementary table 1. Human primer sequences for real-time PCR of gene markers during ESC differentiation.

Transcript	Forward	Reverse
<i>ACTB</i>	TGAACCCCAAGGCCAACCGC	GACCCCGTCACCGGAGTCCA
<i>AXIN2</i>	AGTCAGCAGAGGGACAGGAA	AGCTCTGAGCCTTCAGCATC
<i>CHD1/E-CADHERIN</i>	GGATGTGCTGGATGTGAATG	CTCAAATCCTCCCTGTCCA
<i>CTNNB1</i>	TCCCACTGGCCTCTGATAAA	AAC TAGTCGTGGAATGGCAC
<i>ΔNp63</i>	GGAAAACAATGCCCAGACTC	GTGGAATACGTCCAGGTGGC
<i>DLX5</i>	ACGCTAGCTCCTACCACCAG	TTTGCCATTACCATTTCTCA
<i>FOXD3</i>	GCATCTGCGAGTTCATCAGC	CGTTGAGTGAGAGGTTGTGG
<i>GATA3</i>	GAGGAGGTGGATGTGCTTTT	CAGGGTAGGGATCCATGAAG
<i>HES5</i>	CTCAGCCCCAAAGAGAAAAA	TAGTCCTGGTGCAGGCTCTT
<i>MIXL1</i>	AGCTGCTGGAGCTCGTCTTC	TGGAAGGATTTCCCACTCTG
<i>MSX1</i>	CTGCACCCTCCGCAAACACA	AGGCTGAGCGAGCTGGAGAA
<i>MSX2</i>	CGGTCAAGTCGGAAAATTCA	GAGGAGCTGGGATGTGGTAA
<i>NANOG</i>	CAAAGGCAAACAACCCACTT	TCTGCTGGAGGCTGAGGTAT
<i>NR2F1</i>	CGAGTACAGCTGCCTCAAAG	TACTGGCTCCTCACGTACTC
<i>OCT4</i>	GAAGGATGTGGTCCGAGTGT	GTGAAGTGAGGGCTCCATA
<i>PAX3</i>	AATTACTCAAGGACGCGGTC	TTCTTCTCGCTTTCCTCTGC
<i>PAX6</i>	CACCTACAGCGCTCTGCCGC	CCCGAGGTGCCATTGGCTG
<i>PAX7</i>	TGACAGCTTCATGAATCCGG	GATGGAGAAGTCAGCCTGTG
<i>SIX1</i>	TCAGCTCCAAGACTCTCTGC	ACAAGCTGCAAAAATGTTCC
<i>SNAI2</i>	CAGACCCTGGTTGCTTCAAG	GAGCCCTCAGATTTGACCTG
<i>SOX2</i>	TCAAGCGGCCCATGAATGCC	AGCCGCTTAGCCTCGTCGAT
<i>SOX10</i>	CTCTGGAGGCTGCTGAA	TGGGCTGGTACTTGTAGTC
<i>T</i>	AGGCTCCCGTCTCCTTCAGCAA	TGGCTGGTGATCATGCGCTGT
<i>TFAP2A</i>	GATCCTCGCAGGGACTACAG	TACCCGGGTCTTCTACATGC
<i>ZIC1</i>	GTCCTACACGCATCCCAGTT	GCGATAAGGAGCTTGTGGTC
<i>ZIC3</i>	AAGTCTTTCAAGGCGAAGTA	GTCACAGCCTTCAAATTCAC

Supplementary table 2. Short-hairpin RNA sequences and information for knockdown experiments in hESCs.

Transcripts	ID	RNA consortium Clone ID	Target sequence	Match region	Knockdown efficiency*
<i>CTNNB1</i>	shRNA1	TRCN0000003844	CGCATGGAAGAAATAGTTGAA	CDS	62%
<b><i>CTNNB1</i></b>	<b>shRNA2</b>	<b>TRCN0000314921</b>	<b>TCTAACCTCACTTGCAATAAT</b>	<b>CDS</b>	<b>82%</b>
<i>CTNNB1</i>	shRNA3	TRCN0000010824	CCATTGTTTGTGCAGCTGCTT	CDS	46%
<i>PAX6</i>	shRNA1	TRCN0000016123	GCAAGAATACAGGTATGGTTT	CDS	74%
<i>PAX6</i>	shRNA2	TRCN0000016125	CGTCCATCTTTGCTTGGGAAA	CDS	68%
<b><i>PAX6</i></b>	<b>shRNA3</b>	<b>TRCN0000246162</b>	<b>ATACGCACTGTTGGTACAATT</b>	<b>3UTR</b>	<b>97%</b>

\*Knockdown efficiency was measured by real-time PCR in undifferentiated hESCs and differentiated ESC cultures at day 5 for *CTNNB1* and *PAX6* respectively. The species with the highest knockdown were bolded and selected for functional analyses.



Supplementary table 3. Antibodies used for immunofluorescence.

Protein/Gene	Source	Catalog no.	Antibody type	Dilution
PAX7	Developmental hybridoma bank	Pax7	Mouse IgG1	1:50
PAX6	Developmental hybridoma bank	Pax6	Mouse IgG1	1:100
TFAP2A	Developmental hybridoma bank	3B5	Mouse IgG2b	1:50
ISL1	Developmental hybridoma bank	Isl1/2	Mouse IgG2b	1:100
SOX10	Vivian Lee	-	Rabbit	1:3000
POU5F1 (OCT4)	Santa Cruz Biotechnology	SC-5279	Mouse IgG2b	1:250
T (BRACHYURY)	R&D Systems	AF2085	Goat	1:200
HuC/D	Molecular probe	A-21271	Mouse IgG2b	1:300
S100 $\beta$	Thermo-Scientific	MA1-2505	Mouse IgG1	1:100
MITF	R&D Systems	AF5769	Goat	1:50
PRPH	Millipore	AB1530	Rabbit	1:200
TUJ1	Covance	MMS-435P-250	Mouse IgG2a	1:2000
GFAP	Covance	SMI-22R	mouse IgG2b	1:500