Supplemental information:

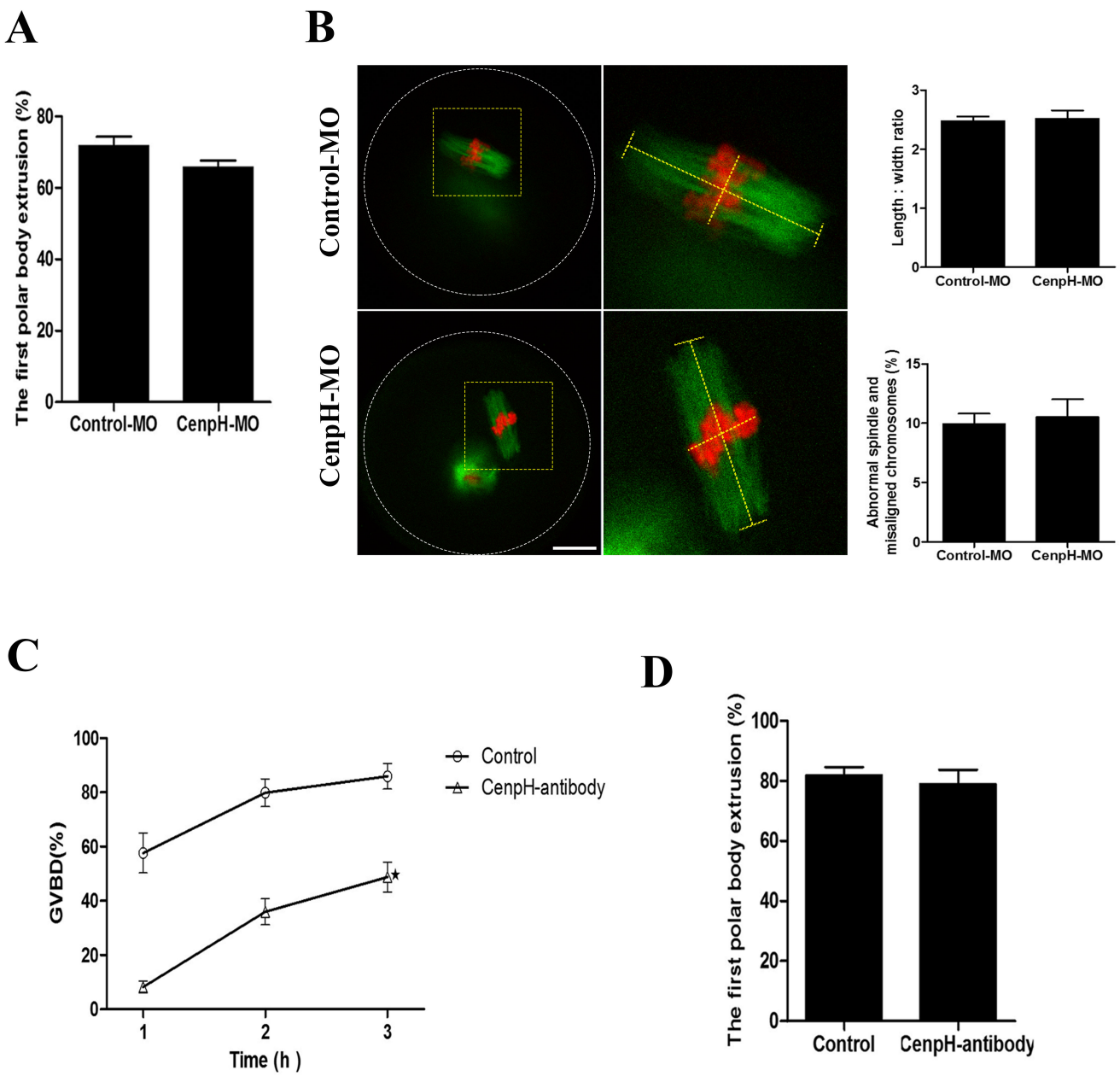

Figure S1. CenpH is not required for polar body extrusion and MII spindle morphology. (A and B) About 37% of injected CenpH MO oocytes underwent GVBD by 3 h following release from IBMX, and these oocytes were used. (A) The percentage of first polar body extrusion at 14 h following release from IBMX in control-MO and CenpH-MO oocytes. (B) Confocal images of control-MO and CenpH-MO oocytes immunostained for DNA, and microtubules (α-tubulin) at 14h. Graph showing length: width ratios; abnormal spindle and misaligned chromosomes for control-MO and CenpH-MO oocytes at 14 h (n=20 and n=18). (C) CenpH antibody was microinjected into GV stage oocytes. The percentages of GVBD at 1, 2 and 3 h following microinjection of CenpH antibody in control and CenpH-antibody oocytes. (D)CenpH antibody was microinjected into GVBD stage oocytes. The percentage of first polar body extrusion at 14 h following microinjection of CenpH antibody in control and CenpH-antibody injected oocytes. (E) CenpH-MO was injected into GV stage oocytes followed by 20 h incubation in IBMX. Subsequently, high-concentration of cyclin B1 mRNA was injected into these GV oocytes followed by 2h incubation in IBMX. The cyclin B1 protein levels of control-MO oocytes and CenpH-MO-cyclin B1-GFP oocytes were assessed by Western blot. (F) CenpH-MO was injected into GV stage oocytes followed by a 24 h incubation in IBMX to deplete the protein. The CenpH protein levels of control-MO oocytes and CenpH-MO oocytes (GV or GVBD) by 2 h following release from IBMX were assessed. Data are mean ± SEM.* significantly different ($p < 0.05$). Scale bars: 20 μm. The total numbers of analysed oocytes are indicated (n).