

Figure S1: β-cell specific expression of dnVegfaa at embryonic or post-embryonic stages results in islet disruption. A-B) Pancreatic endocrine cells (green) in 56 hpf *Tg(NeuroD:GFP)*; *Tg(ins:DsRed)* and *Tg(NeuroD:GFP)*; *Tg(ins:dnvegfaa)* embryos. C-D) Confocal projection images of the pancreatic islet in 56 hpf *Tg(ins:DsRed)* and *Tg(ins:dnvegfaa)* animals showing Glucagon-positive α-cells (cyan). E) Number of β-cells in 48 hpf *Tg(ins:dnvegfaa)* and *Tg(ins:DsRed)* animals; mean ± SEM, n = 6-7 animals. P values from t-tests are presented. F) Induction of dnVegfaa expression in *TgBAC(pdx1:CreERT2)*; *Tg(ins:loxP-GFP-STOP-loxP-mTomato-P2A-dnvegfaa)* animals with 10 μ M Tamoxifen treatment starting at 72 hpf and imaging at 120 hpf. G-H) β-cells in 120 hpf *TgBAC(pdx1:CreERT2)*; *Tg(ins:loxP-GFP-STOP-loxP-mTomato-P2A-dnvegfaa)* larvae treated with DMSO (vehicle) or Tamoxifen. Yellow dotted lines outline the spread of the β-cells in the primary islet. Maximum intensity projections are presented; A, anterior; D, dorsal.

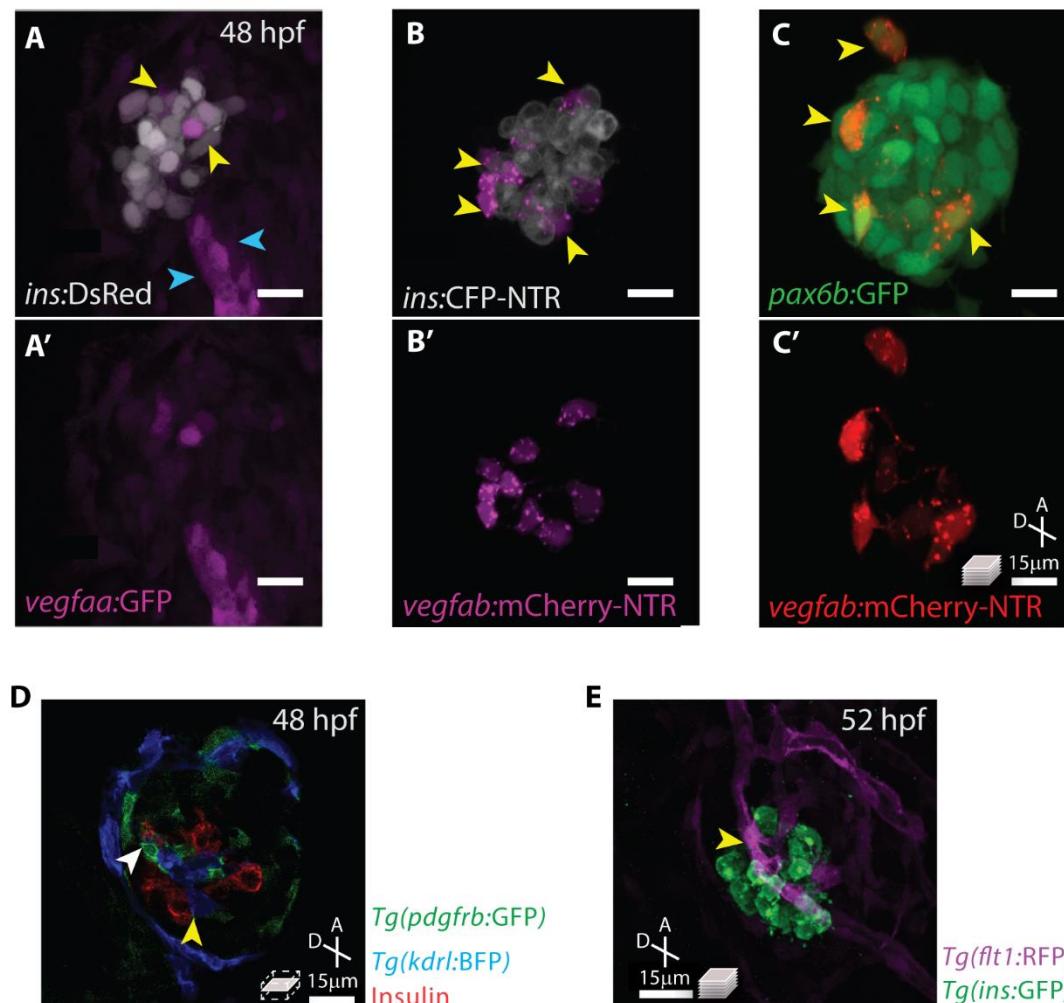


Figure S2: *vegfaa* and *vegfab* are expressed in and around the pancreatic endocrine cells. A) β-cells (white) in 48 hpf *TgBAC(vegfaa:GFP); Tg(ins:DsRed)* embryos. Yellow arrowheads point to *vegfaa* expressing β-cells. Blue arrowheads point to *vegfaa* expressing cells close to the islet that likely belong to non-endocrine lineages. B) β-cells (white) in 48 hpf *TgBAC(vegfab:gal4ff); Tg(UAS-E1b:Nfsb-mCherry); Tg(ins:CFP-NTR)* embryos. *TgBAC(vegfab:gal4ff); Tg(UAS-E1b:NfsB-mCherry)* is referred to as *Tg(vegfab:mCherry-NTR)*. Yellow arrowheads point to *vegfab* expressing cells around β-cells. C) Pancreatic endocrine cells (green) in 48 hpf *Tg(vegfab:mCherryNTR); Tg(pax6b:GFP)* embryos. Yellow arrowheads point to *vegfab* expressing endocrine cells. Maximum intensity projections are presented. D) Confocal plane of the primary pancreatic islet in a 48 hpf *Tg(kdrl:BFP); Tg(pdgfrb:GFP)* embryo showing β-cells (red). Yellow arrowhead points to a *kdrl* expressing endothelial cell in contact with a β-cell. White arrowhead points to a *pdgfrb* expressing cell in contact with a β-cell. E) Confocal projection image of the primary pancreatic islet in a 52 hpf *Tg(ins:GFP); Tg(flt1:RFP)* embryo showing β-cells (green). Yellow arrowhead points to a *flt1* expressing blood vessel in the islet. ; A, anterior; D, dorsal.

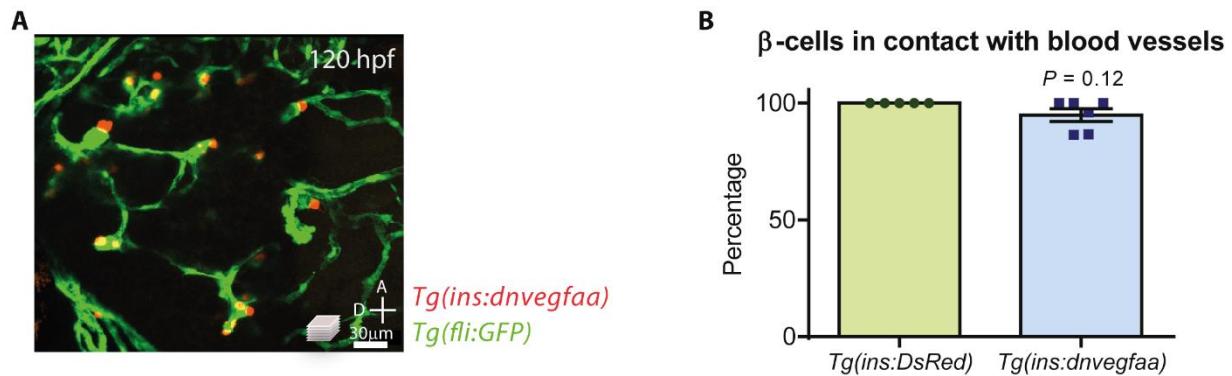


Figure S3: β -cells in *Tg(ins:dnvegfaa)* animals are in contact with blood vessels. A) Confocal projection image of the pancreatic islet in a 120 hpf *Tg(ins:dnvegfaa);Tg(fli:GFP)* animal showing β -cells (red). Maximum intensity projection is presented. B) Percentage of β -cells in contact with blood vessels in 120 hpf *Tg(ins:DsRed)* and *Tg(ins:dnvegfaa)* animals; mean \pm SEM, n = 6-7 animals. P values from t-tests are presented.

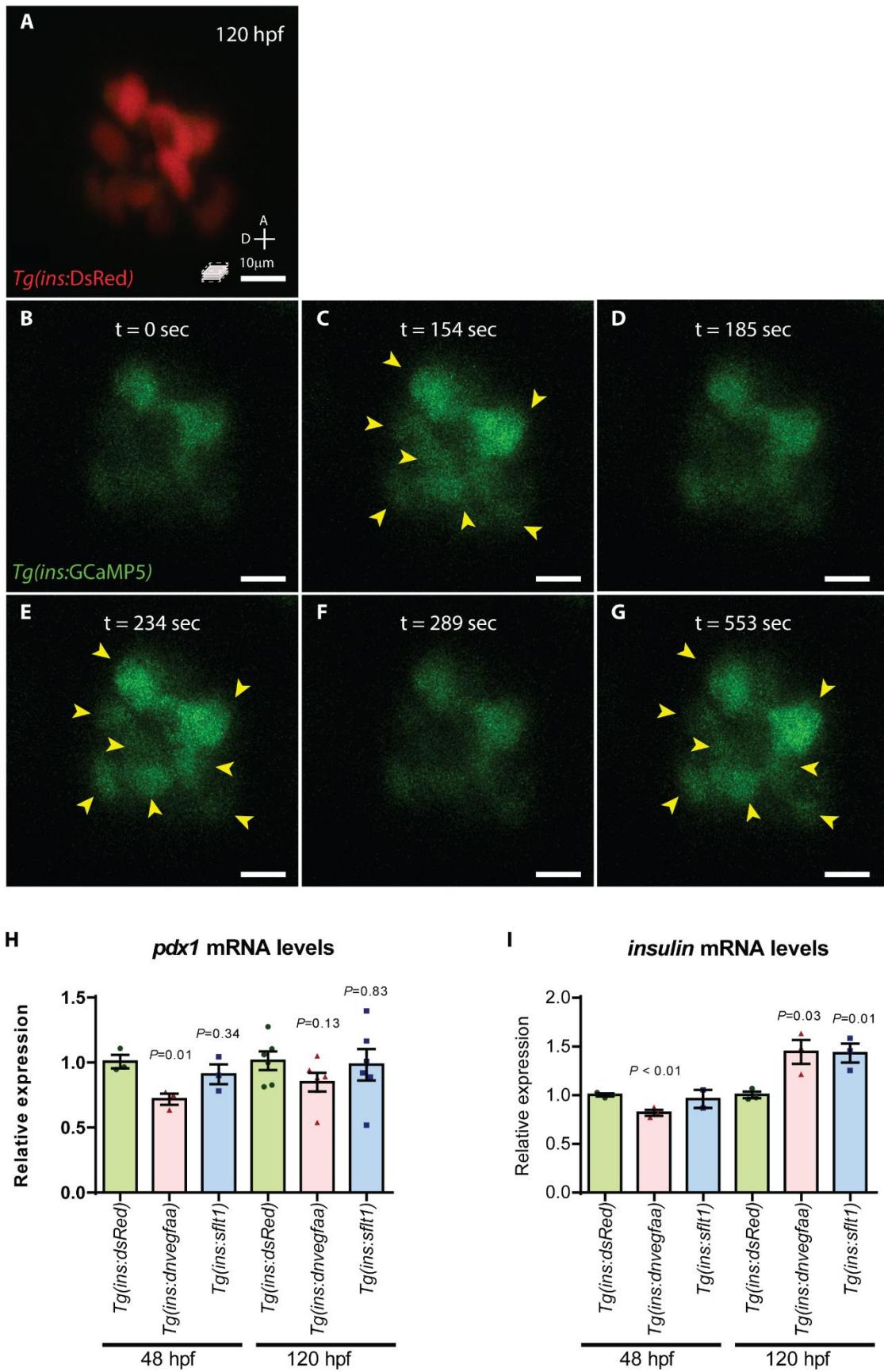


Figure S4: Live imaging shows dynamics of calcium signaling in β -cells. A) Pancreatic β -cells (red) in 120 hpf *Tg(ins:DsRed); Tg(ins:GCaMP5)* animals. B-G) Calcium responsive fluorescence signal in 120 hpf *Tg(ins:DsRed); Tg(ins:GCaMP5)* animals, at the indicated time point, from live confocal imaging. Yellow arrowheads point to β -cells showing an observable increase in fluorescence signal intensity. Maximum intensity projections of planes in the central 30 μm region of the islet are shown. A, anterior; D, dorsal. H-I) *pdx1* (H) and *ins* (I) mRNA levels in 48 and 120 hpf animals; 3-6 replicates, 10 animals per replicate, mean \pm SEM. *P* values from t-tests are presented.

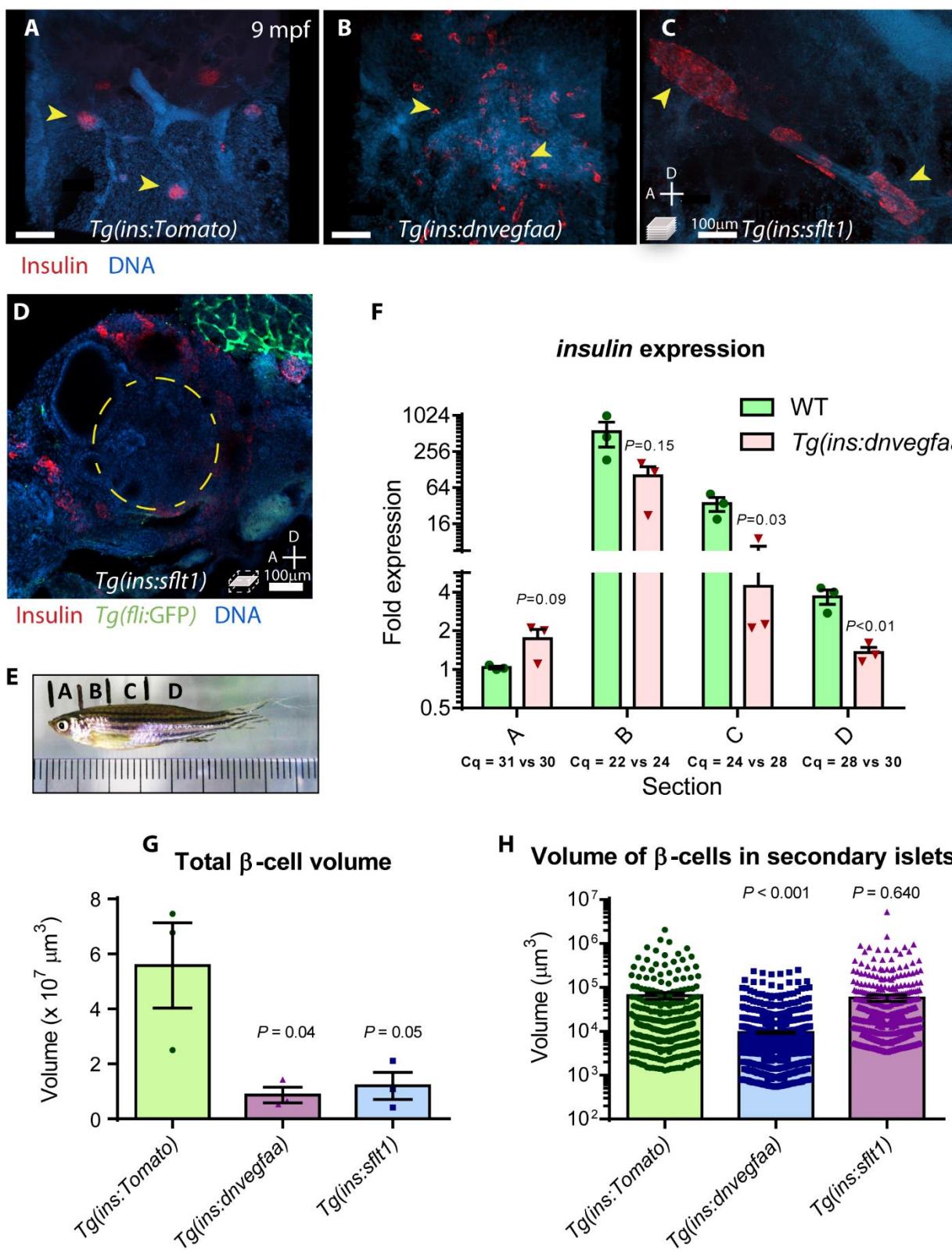
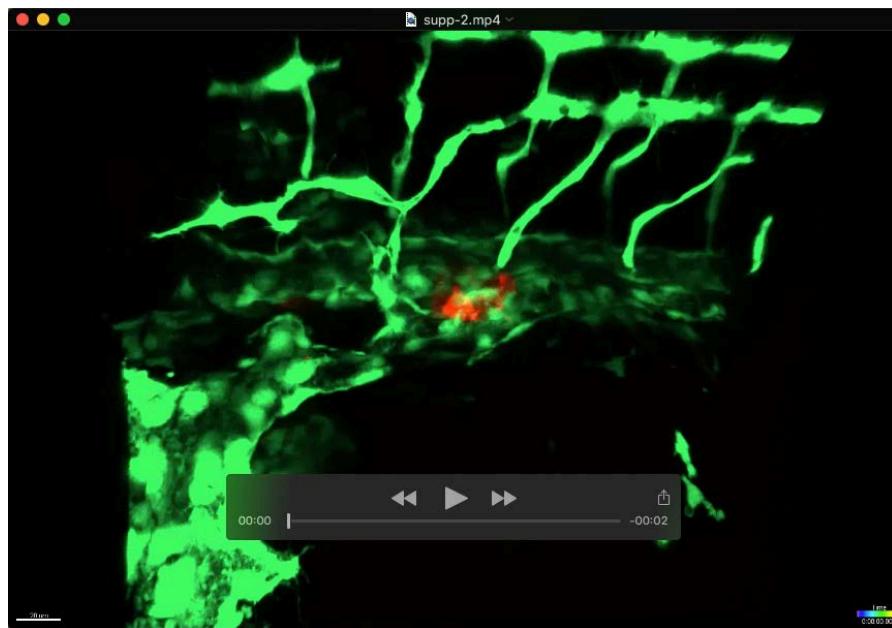


Figure S5: β-cell specific expression of dnVegfaa or sFlt1 causes defects in secondary islet formation.

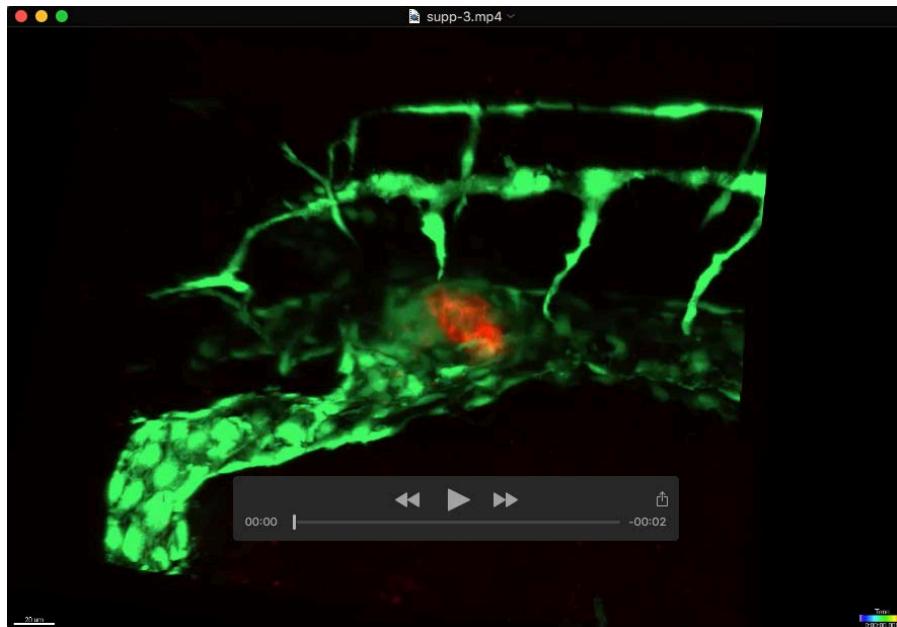
A-C) Wholemount immunostaining for β-cells (Insulin) along the pancreatic ducts of 9 mpf zebrafish after CLARITY-based tissue clearing. Yellow arrowheads point to β-cell clusters. D) Wholemount immunostaining for β-cells (Insulin) in the primary islet of a 9 mpf *Tg(ins:sflt1); Tg(fli:GFP)* zebrafish after CLARITY-based tissue clearing. Yellow dotted line outlines the nuclei in the core of the islet. E) Adult zebrafish divided up into four sections, as marked, to obtain RNA and quantify *insulin* expression in each section separately; section A - head, heart, eyes; section B - anterior gastrointestinal (GI) tract; section C - posterior GI tract; section D - lower trunk and tail regions. F) *insulin* mRNA levels (qPCR) from each of the four sections along with Ct values. G) Volume of Insulin antibody immunostaining (β-cells) in adult zebrafish pancreas, computed using surface construction in Imaris. H) Volume of Insulin antibody immunostaining (β-cells) in secondary islets, computed using surface construction in Imaris. Data points greater than $5.0 \times 10^5 \mu\text{m}^3$ in *Tg(ins:Tomato)* and *Tg(ins:sflt1)* animals correspond to the largest secondary islet clusters located close to the primary islet (Figure 4), mean ± SEM, n = 3 animals, 100 to 560 secondary islets per animal, P values from t-tests are presented.

	Ensembl Gene ID	Gene	Normalized Read Count (β -cells)	% of pancreatic cells with expression
Ligands	ENSDARG00000035350	<i>ins</i>	4324351	99.76
	ENSDARG00000103542	<i>vegfaa</i>	4101	6.64
	ENSDARG00000034700	<i>vegfab</i>	11588	13.72
	ENSDARG00000069640	<i>vegfc</i>	0	0.4
	ENSDARG00000092480	<i>vegfbb</i>	0	ND
	ENSDARG00000077588	<i>pdgfc</i>	147	0.56
	ENSDARG00000058424	<i>pdgfab</i>	420	1.61
	ENSDARG00000055505	<i>pdgfaa</i>	194	1.05
	ENSDARG00000077677	<i>pdgfd</i>	442	ND
	ENSDARG00000038139	<i>pdgfb</i>	0	0.93
Receptors	ENSDARG00000019371	<i>flt1</i>	1	1.01
	ENSDARG00000017321	<i>kdr</i>	0	0.08
	ENSDARG00000105215	<i>kdrl</i>	16	1.97
	ENSDARG00000015717	<i>flt4</i>	1	0.64
	ENSDARG00000070494	<i>pdgfra</i>	2	0.68
	ENSDARG00000100897	<i>pdgfrb</i>	0	ND
	ENSDARG00000006456	<i>pdgfrl</i>	755	0.60
	ENSDARG00000027290	<i>nrp1b</i>	58	0.32
	ENSDARG00000071865	<i>nrp1a</i>	139	2.01
	ENSDARG00000096546	<i>nrp2a</i>	175	0.40
	ENSDARG00000038446	<i>nrp2b</i>	0	0.20

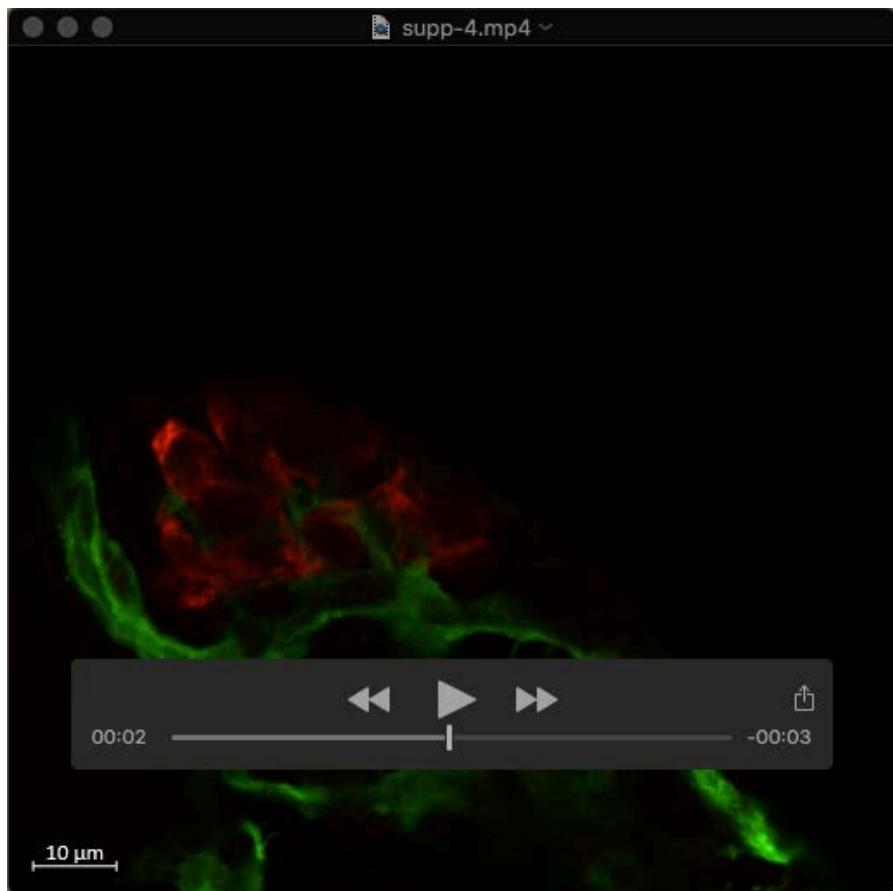
Table S1: Pancreatic β -cell expression of Vegf family ligand and receptor genes. Normalized read counts (Tarifeño-Saldivia et al., 2017) and percentage of pancreatic cells (Salem et al., 2019) expressing different Vegf family ligand and receptor genes; ND, not detected.



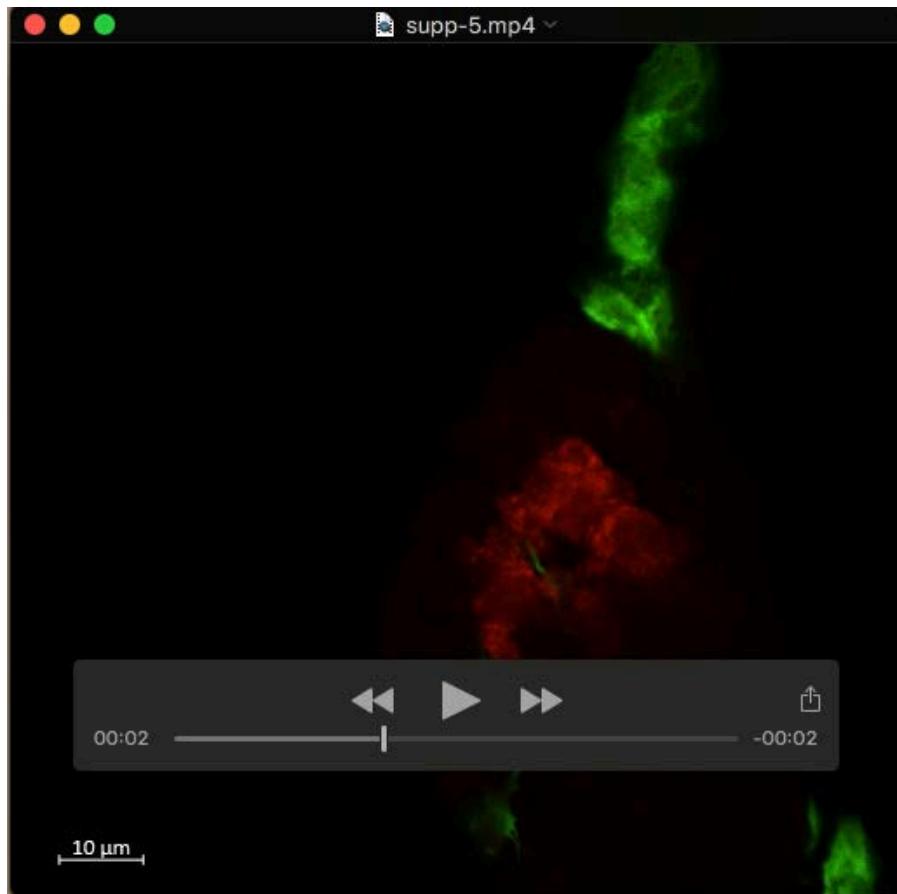
Movie 1: Time-lapse imaging shows collective migration of β -cells and their interaction with the vasculature in wild-type embryos. Confocal imaging of *Tg(ins:Tomato); Tg(fli:GFP)* zebrafish from 34 to 48 hpf at 60 min time intervals. Movie corresponds to Figure 1e.



Movie 2: Time-lapse imaging shows dispersion of β -cells in *Tg(ins:dnvegfaa)* embryos. Confocal imaging of *Tg(ins:dnvegfaa); Tg(fli:GFP)* zebrafish from 34 to 48 hpf at 60 min time intervals. β -cells are marked in red. Movie corresponds to Figure 1f.

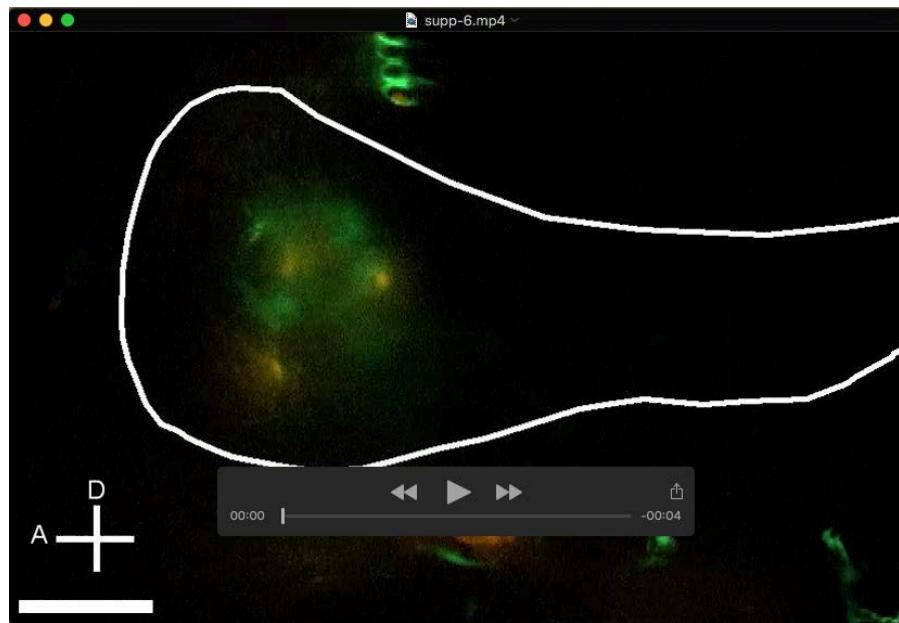


Movie 3: Wild-type pancreatic islets contain intra-islet vessels. Progressive planes in a Z-stack from confocal imaging of pancreatic islets in 48 hpf *Tg(ins:DsRed); Tg(fli:GFP)* zebrafish, with immunostaining for Insulin (red) and GFP. Movie corresponds to Figure 2c.

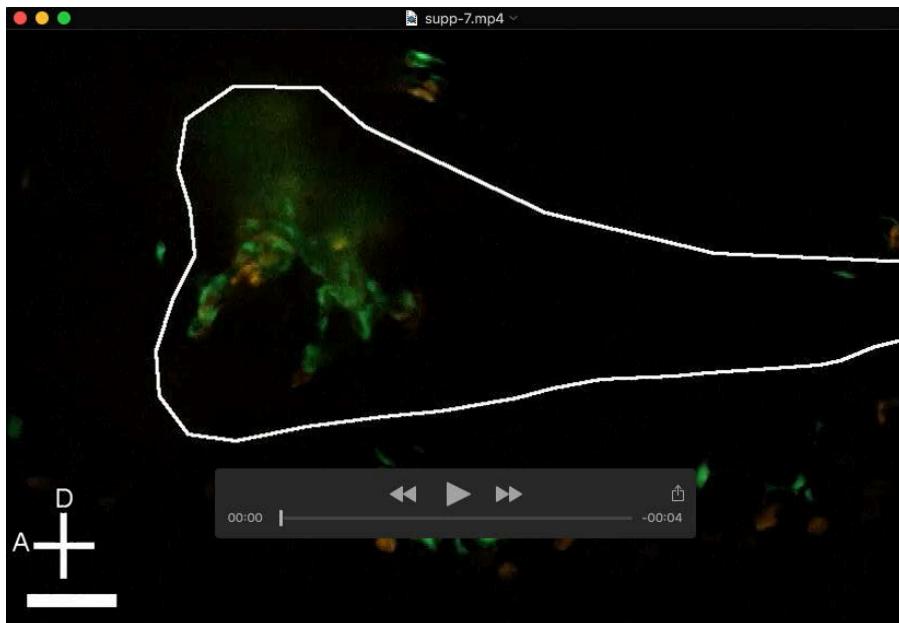


Movie 4: β -cell-specific sFlt1 expression results in markedly reduced intra-islet vasculature.

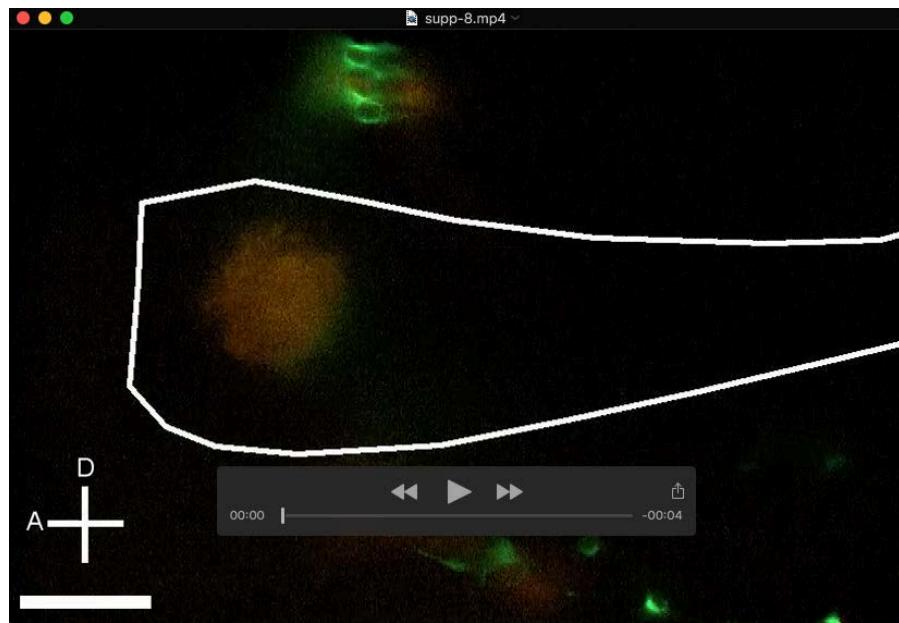
Progressive planes in a Z-stack from confocal imaging of pancreatic islets in 48 hpf *Tg(ins:sflt1); Tg(fli:GFP)* zebrafish, after immunostaining for Insulin (red) and GFP. Movie corresponds to Figure 2j.



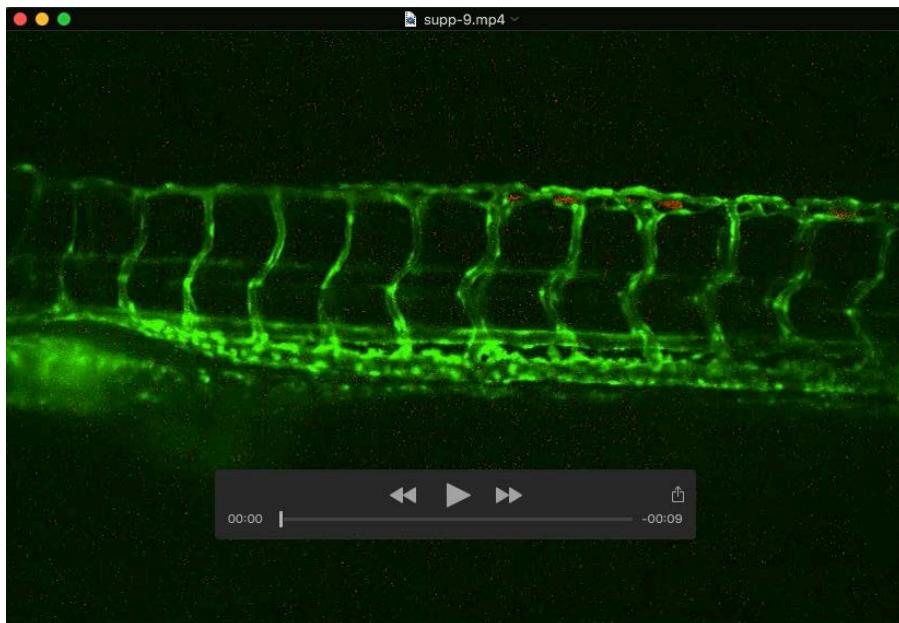
Movie 5: Live imaging shows circulating red blood cells in wild-type larval pancreas. Spinning disk confocal imaging of the pancreatic islet in 120 hpf *Tg(gata1:DsRed); Tg(fli:GFP)* zebrafish, showing circulating red blood cells (red). Pancreas is outlined in white. Scale bar, 40 μm .



Movie 6: Live imaging shows circulating red blood cells and disrupted vasculature in *Tg(ins:dnvegfaa)* pancreas. Spinning disk confocal imaging of the pancreatic islet in 120 hpf *Tg(ins:dnvegfaa); Tg(gata1:DsRed); Tg(fli:GFP)* zebrafish. Pancreas is outlined in white. Scale bar, 40 μm .



Movie 7: Live imaging shows lack of vasculature and circulation in *Tg(ins:sflt1)* pancreas. Spinning disk confocal imaging of the pancreatic islet in 120 hpf *Tg(ins:sflt1); Tg(gata1:DsRed); Tg(fli:GFP)* zebrafish. Faint red signal in the anterior region of the pancreas (outlined in white) is from β -cells. Scale bar, 40 μ m.



Movie 8: Live imaging shows circulating β -cell in *Tg(ins:dnvegfaa)* larvae. Spinning disk confocal imaging of 72 hpf *Tg(ins:dnvegfaa); Tg(fli:GFP)* zebrafish. Arrowhead points to the β -cells circulating in the trunk region.