

Figure S1

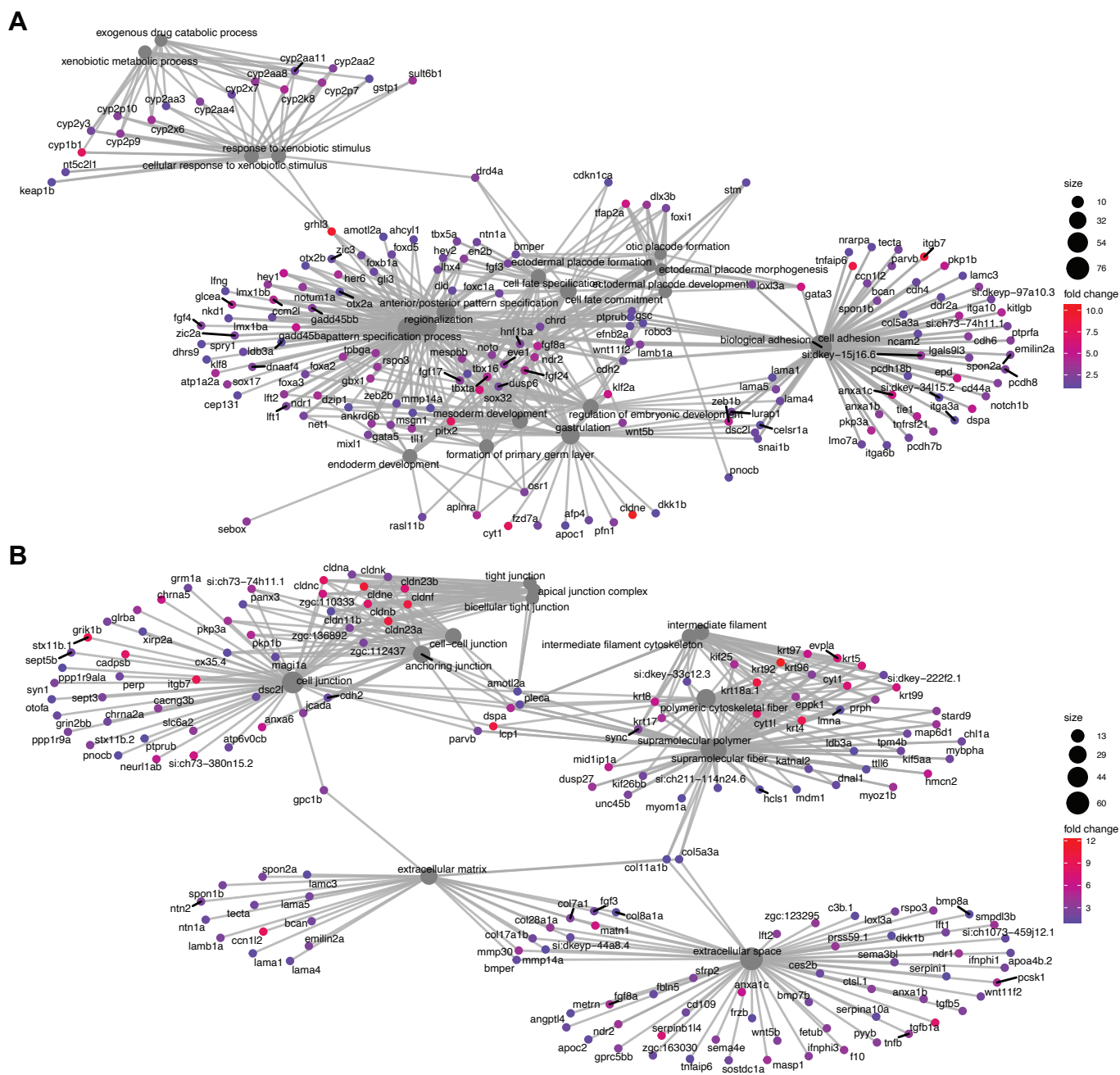


Figure S1. Gene ontology (GO) gene-concept network analysis of RNA-seq data.

(A) *irf6*^{-/-} embryos have perturbations in biological processes such as cell adhesion, gastrulation, mesoderm, ectoderm and endoderm development, and response to xenobiotic stimulus. (B) *irf6*^{-/-} embryos also have perturbations of various cellular compartments including cell junctions, cytoskeletal elements and extracellular space. Grey nodes show GO terms, colored nodes show individual genes from the RNA-seq dataset, and edges connect genes to one or more associated GO terms. Colored nodes show relative enrichment (measured by fold-change) of genes in wild type samples relative to *irf6*^{-/-} embryos. Maps were generated using the enrichplot package in R.

Figure S2. GO Enrichment analysis of the wt vs. *mz-irf6*^{-/-} RNA-seq dataset.

Nodes show GO terms associated with enriched genes in the dataset. Edges and arrows show previously annotated relationships between GO terms. Developmental processes such as ectodermal development and otic placode formation are enriched in the analysis, which involve the recruitment of neural crest cell progenitors also involved in palate development. Color represents q-value, with red having a lower q-value and blue having a higher q-value. Q-values were computed using FDR with the Benjamini-Hochberg correction. Data were visualized using the enrichGO and goplot packages in R.

Figure S3. KEGG Graph of Cell Adhesion Molecules. KEGG pathway analysis that maps enriched genes in our dataset to a database containing cellular pathway information. Colored nodes show increased expression of genes in red and decreased expression in green for wild type fish compared to control. Many cell-adhesion molecular interactions are disrupted in the *irf6*^{-/-} embryos. Data were generated using the enrichKEGG and pathview packages in R.

Figure S4

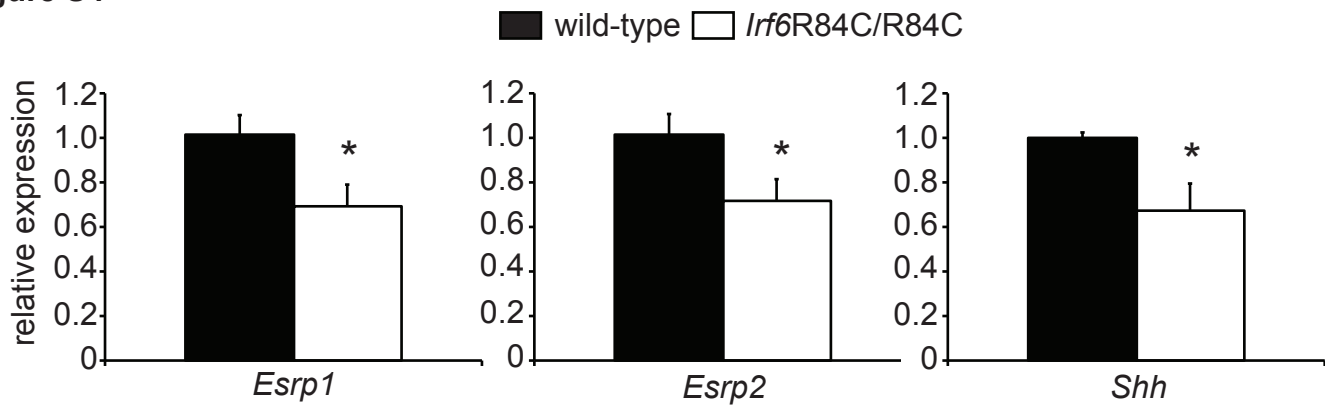


Figure S4. Decreased expression of *Esrp1* and *Esrp2* in *lrf6*^{R84C} mutant mouse embryos. *lrf6*^{R84C/WT} mice were intercrossed, and embryos were collected at E11.5. Embryos were individually genotyped and heads of *lrf6*^{WT/WT} and *lrf6*^{R54C/R84C} embryos were utilized for RNA isolation and RT-qPCR. Expression of *Esrp1*, *Esrp2* and *Shh* were significantly decreased in the *lrf6*^{R54C/R84C} embryos relative to wild type. Expression levels were normalized to *TBP* expression. n=5. Students t-test, p<0.05.

Figure S5

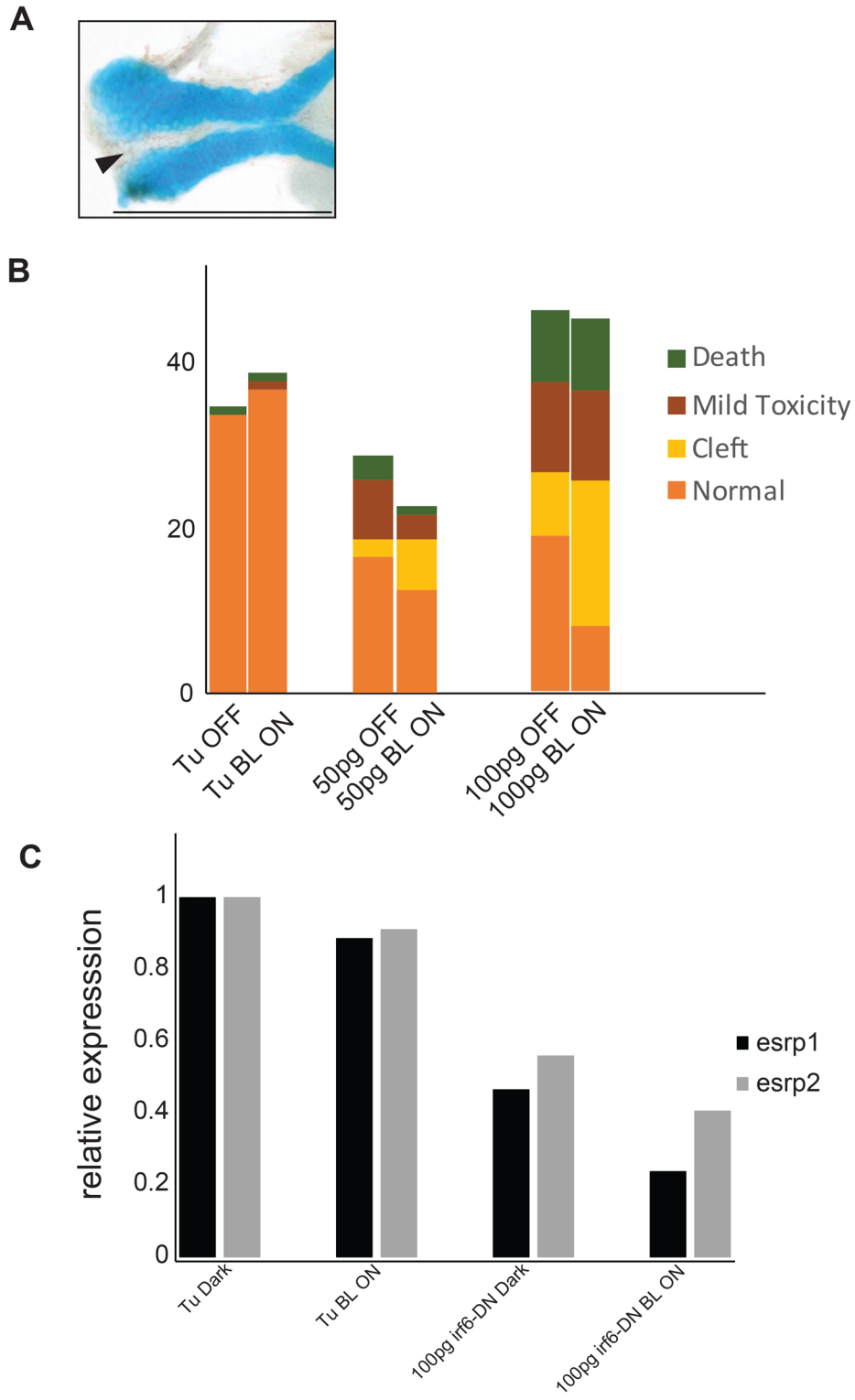


Figure S5. Optogenetic disruption of *irf6*. (A) Cleft ANC phenotype observed in *irf6*-DN embryos exposed to blue light at 10 hpf. (B) Distribution of observed phenotype in wild type (uninjected) vs. *irf6*-DN injected embryos at 50pg/50pg or 100pg/100pg doses of poly-A tailed EL222-VP16 mRNA and (C120)-*irf6*-DN plasmid grown in the dark or exposed to blue light at 10 hpf. Bar chart height represents absolute number of embryos. (C) qPCR gene expression analysis for *esrp1* and *esrp2*, showing approximately two-fold and five-fold reduction in *esrp1* expression in embryos injected with the optogenetic *irf6*-DN system grown in the dark, or under blue light, respectively. *esrp2* follows a similar trend. Scale bars: 150um.

Figure S6

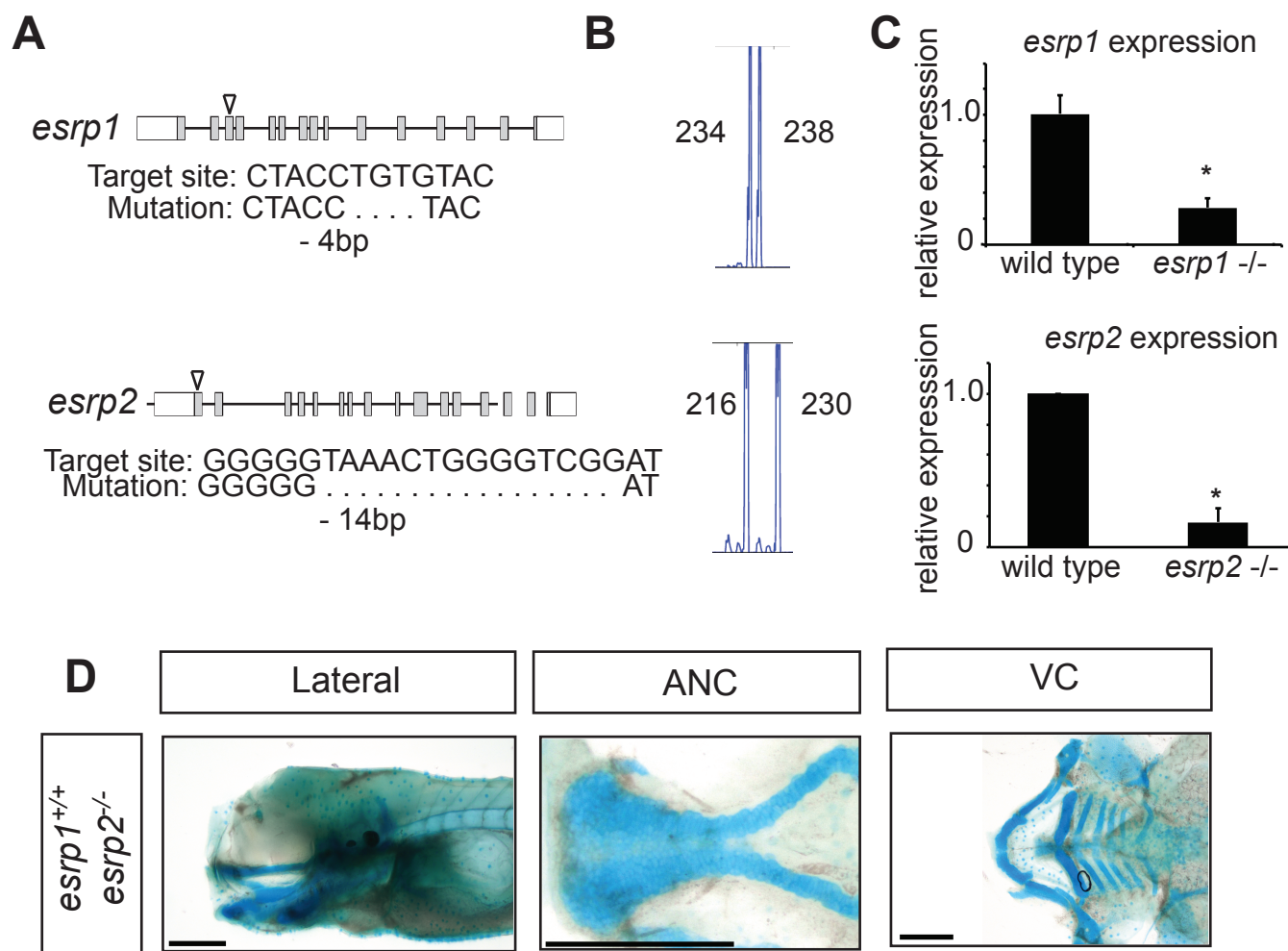
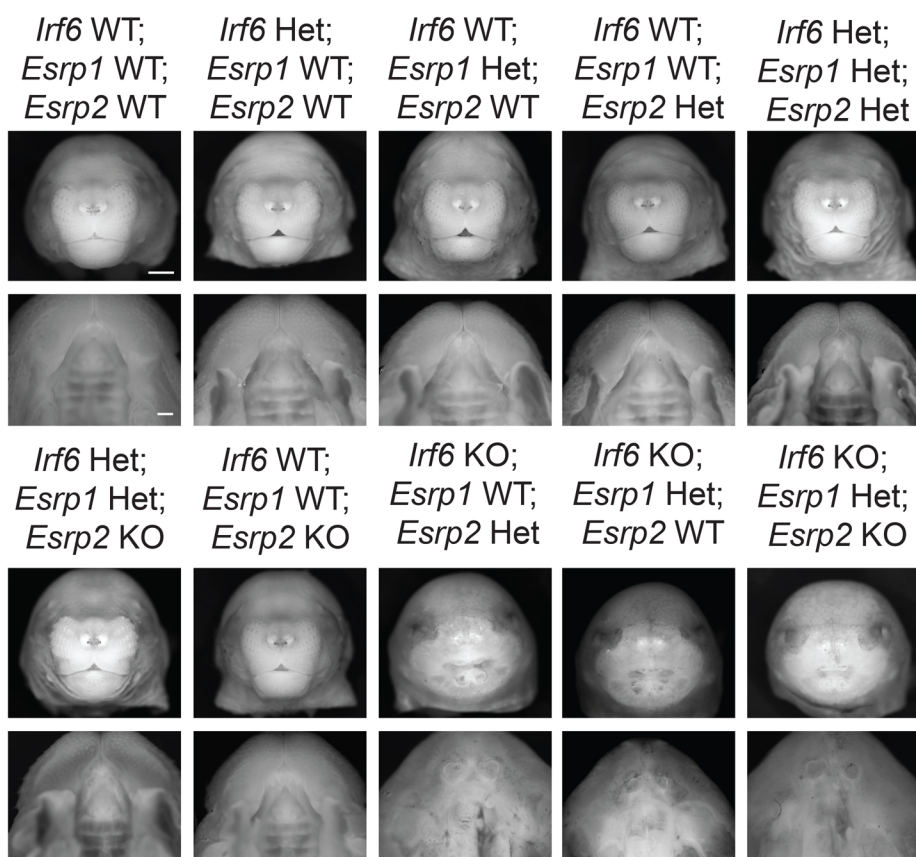


Figure S6. Generation of *esrp1* and *esrp2* CRISPR/Cas9 gene disruption in zebrafish. (A) Schematic representations of *esrp1* and *esrp2* exons, positions of target site (arrow), and sequences of mutations. (B) Micro-satellite results of genotyping PCR showing a size shift consistent with the observed mutation. (C) Expression of the CRISPR/Cas9 targeted gene is decreased relative to wild type, as measured by qPCR. $n=3,4$. $p<0.01$ (D) *esrp1*^{+/+}; *esrp2*^{-/-} fish embryos at 4dpf exhibit normal craniofacial development of the anterior neurocranium and ventral cartilages. Scale bars: 200 μ m.

Figure S8

Figure S7. Representative images of *Irf6*^{R84C/WT}; *Esrp1*^{+/-}; *Esrp2*^{+/-} triple

heterozygote in-crosses. Mice compound heterozygous for *Irf6*^{R84C}, *Esrp1* and *Esrp2* were generated by breeding *Irf6*^{R84C/+} with *Esrp1*^{+/-}; *Esrp2*^{-/-} mice. The triple heterozygotes were then inter-crossed and embryos were collected at E18.5.

Representative frontal and oral images of embryos displaying no craniofacial phenotype in single heterozygotes, triple heterozygotes or *Irf6*^{R84C/+}; *Esrp1*^{+/-}; *Esrp2*^{-/-}. Further, *Esrp1/2* alleles did not appear to modify *Irf6*^{R84C/R84C} phenotype, however epithelial adhesions hampered a thorough observation. Scale bar: 100 μ m.

Table S1. *Irf6*, *Esrp1* and *Esrp2* genotypes interact to produce non-Mendelian embryo ratios.

[Click here to Download Table S1](#)

Table S2. *Irf6*, *Esrp1* and *Esrp2* genotypes interact to produce non-Mendelian embryo ratios

<i>Irf6</i>	<i>Esrp1</i>	<i>Esrp2</i>	probability	Expected	Observed
WT	WT	WT	1/64	1.2	3
WT	WT	Het	1/32	2.5	4
WT	WT	KO	1/64	1.2	1
WT	Het	WT	1/32	2.5	2
WT	Het	Het	1/16	4.9	3
WT	Het	KO	1/32	2.5	4
WT	KO	WT	1/64	1.2	0
WT	KO	Het	1/32	2.5	1
WT	KO	KO	1/64	1.2	1
Het	WT	WT	1/32	2.5	5
Het	WT	Het	1/16	4.9	4
Het	WT	KO	1/32	2.5	3
Het	Het	WT	1/16	4.9	10
Het	Het	Het	1/8	9.9	8
Het	Het	KO	1/16	4.9	2
Het	KO	WT	1/32	2.5	2
Het	KO	Het	1/16	4.9	6
Het	KO	KO	1/32	2.5	3
KO	WT	WT	1/64	1.2	1
KO	WT	Het	1/32	2.5	2
KO	WT	KO	1/64	1.2	0
KO	Het	WT	1/32	2.5	5
KO	Het	Het	1/16	4.9	4
KO	Het	KO	1/32	2.5	5
KO	KO	WT	1/64	1.2	0
KO	KO	Het	1/32	2.5	0
KO	KO	KO	1/64	1.2	0

Irf6^{R84C/WT}; *Esrp1*^{+/-}; *Esrp2*^{+/-} triple heterozygous mice were in-crossed and embryos were collected between E12.5 and E21. Table 1 is a subset of expected number of embryos based on Mendelian genetics versus the observed number of viable embryos. The *Irf6*^{R84C/R84C}; *Esrp1*^{-/-} genotype appears to be lethal prior to E12.5 as approximately 5 embryos were expected but zero embryos were observed. A total of 79 embryos were collected from 9 different litters.