

Table S1: *C. elegans* strains used in this study

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Table S2: *C. elegans* transgenes used in this study

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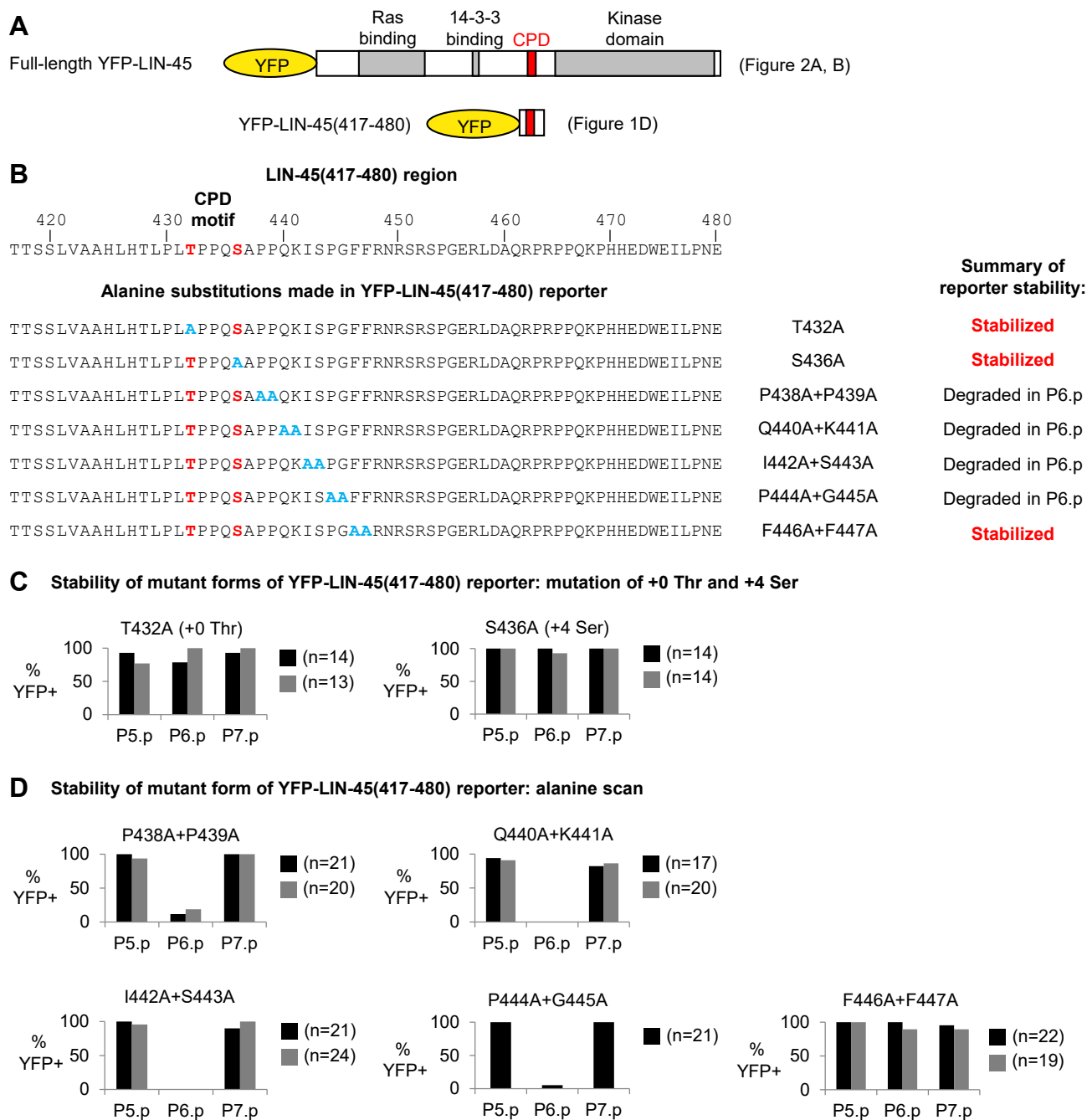


Figure S1. YFP-tagged LIN-45 protein reporters. **A)** Schematic of full-length YFP-LIN-45 and truncated YFP-LIN-45(417-480) proteins. **B)** Amino acid sequence of LIN-45(417-480) region. The +0 Thr and +4 Ser CPD residues are shown in red. In mutant forms tested, alanine substitutions are shown in blue. **C-D)** Stability of mutant forms of the truncated YFP-LIN-45(417-480) reporter. With exception of the P444A+G445A form, two independent strains were scored for each mutant form tested. Percentage of VPCs with visible YFP fluorescence, and number larvae scored (*n*) is shown for each strain. (C) Mutations at the +0 Thr or +4 Ser abolish degradation in P6.p. (D) Mutations at F446 and F447 abolish degradation in P6.p.

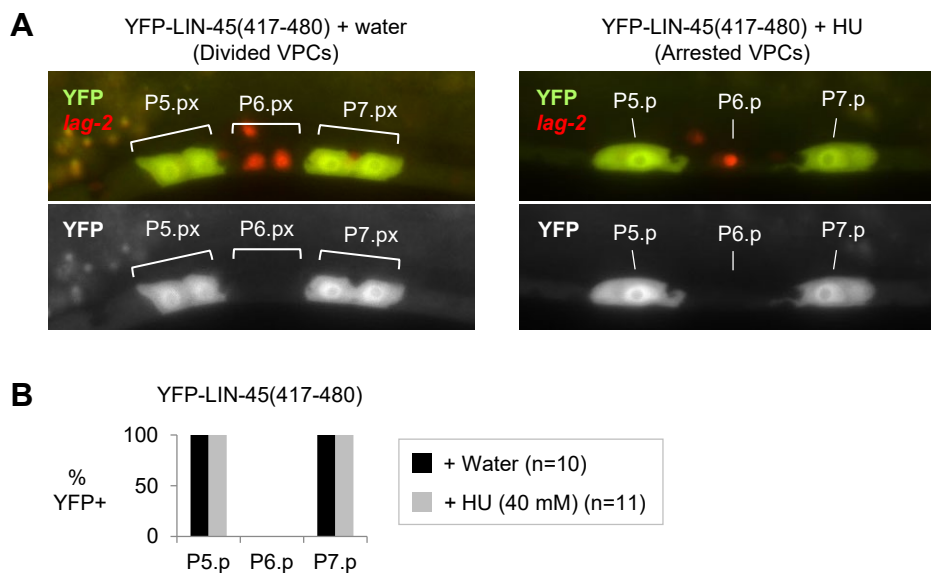


Figure S2. Blockade of S phase does not prevent degradation of the truncated LIN-45(417-480) reporter. Larvae were picked during L2 lethargus based on their lack of locomotion and pharyngeal pumping, transferred to plates with either water (control) or 40 mM hydroxyurea (HU), and grown for 6 hours. At that time, VPCs of all larvae on control plates had divided. For 11/12 larvae grown on HU plates, the VPCs did not divide. We excluded the larva in which VPCs had divided from our analysis. **A)** YFP-tagged LIN-45(417-480) (green) and *arls222[lag-2p::2xNLS-tagRFP]* (red) proteins in representative images of L3 stage larvae. Top, merged image; bottom, YFP-LIN-45(417-480) alone in grayscale. For the water control, VPCs had divided, and the locations of daughters P5.px, P6.px, and P7.px are indicated with brackets. For the HU-treated larva, undivided VPCs P5.p, P6.p, and P7.p are indicated. **B)** Percentage of VPCs positive for YFP-LIN-45(417-480) in either the water control or HU-treated larvae, and the number of larvae scored (n) is shown.

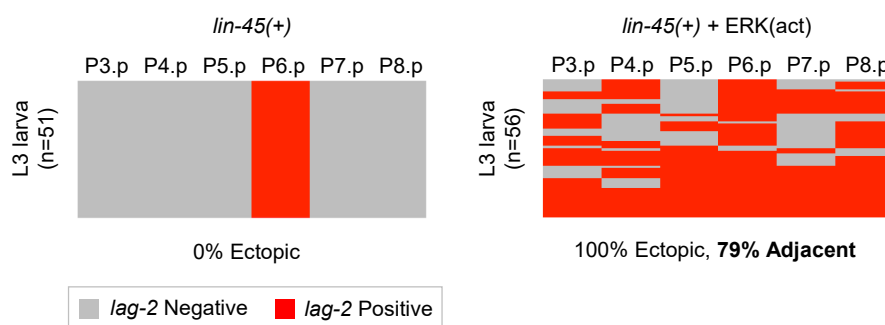


Figure S3. *lag-2* transcription is induced by ERK(act). The *gals37* transgene, referred to here as ERK(act), leads to highly active MPK-1/ERK and causes ectopic expression of a transcriptional reporter for *lag-2*, *arls222[lag-2p::2xNLS-tagRFP]*, a direct target of the EGFR-Ras-Raf-ERK signaling pathway and marker of 1° fate. Expression was scored in L3 stage larvae. Each panel represents the *lag-2* positive (red) or negative (gray) status in individual VPCs. Each larva is represented in a row, and VPC is represented in a column. The number of L3 larvae scored (n) is shown at left of each panel. Based on ectopic *lag-2* expression, larvae were categorized as displaying either alternating or adjacent 1° fate. The percentage of larvae that displayed any ectopic 1° fate (% Ectopic), and adjacent 1° fate (% Adjacent) is indicated at each panel.

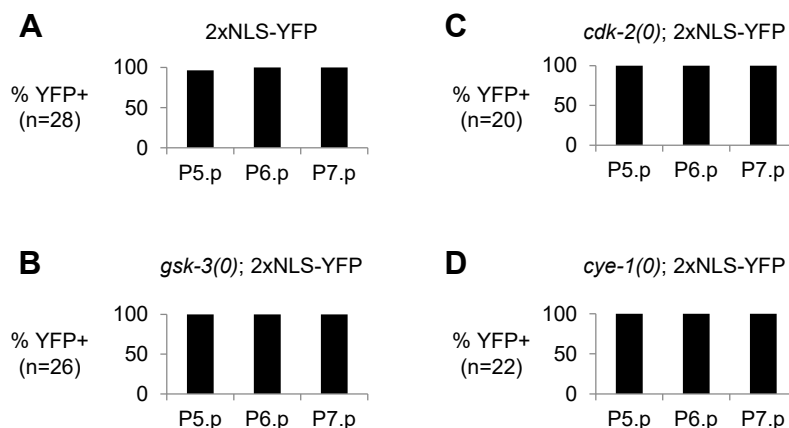


Figure S4. Expression of *lin-31p* is unaffected in *gsk-3*, *cdk-2*, and *cye-1* mutants. In wild-type L3 stage larvae, the *lin-31* promoter (*lin-31p*) is expressed approximately uniformly in P5.p, P6.p, and P7.p. Activity of *lin-31p* was assessed using *arTi88[lin-31p::2xNLS-YFP]*, which drives a nuclear-localized YFP. **A)** Percentage of VPCs positive for 2xNLS-YFP in otherwise wild-type larvae, and the number of larvae score (n) is shown. **B-D)** To confirm that the *lin-31p* expression system used to drive YFP-LIN-45 reporters throughout this work is active in the genotypes examined, *arTi88[lin-31p::2xNLS-YFP]*, was scored in *gsk-3*, *cdk-2*, and *cye-1* null mutants. Percentage of VPCs positive for 2xNLS-YFP, and the number of larvae scored (n) is shown for (B) *gsk-3(0)* larvae, (C) *cdk-2(0)* larvae, and (D) *cye-1(0)* larvae.