

Fig. S1. Cre recombinase activity hindlimb-forming LPM in the the RetRV5Cre transgenic deleter line. LacZ staining of Cre expressing cells at E9.75 (n=8).right-side lateral views of whole embryos. (B,D)Transverse (A,C)cryosections through the hindlimb regions of embryos. A, E9.5 Rosa26RlacZ; RetRV5Cre embryo. LacZ activity is detected in the nascent hindlimb bud. B, Transverse cryosection through the hindlimb region of an E9.5 Rosa26RlacZ; RetRV5Cre embryo. LacZ activity is additionally present in the hindgut diverticulum (black arrowhead) and anterior to the vitelline artery (black asterisk). C, E10.5 Rosa26RlacZ; RetRV5Cre embryo, LacZ activity is detectable throughout the hindlimb buds. **D**, Transverse cryosection through the hindlimbs of an E10.5 Rosa26RlacZ; RetRV5Cre embryo, in which LacZ activity is not detectable in the ectoderm. LacZ activity is visible in the mesoderm of the hindgut (black arrowhead) and the dorsal mesentery (black asterisk). NT-Neural tube, S-somite, PSM-presomitic mesoderm, HL-hindlimb E, Graphical representation of Q-PCR results to analyse the extent of deletion of the Tbx4 conditional allele in 2 separate E10.5 Tbx4lox/lox; RetRV5Cre hindlimbs. An estimated 96% of conditional Tbx4 allele has been excised in Tbx4lox/lox; RetRV5Cre mutant hindlimbs.

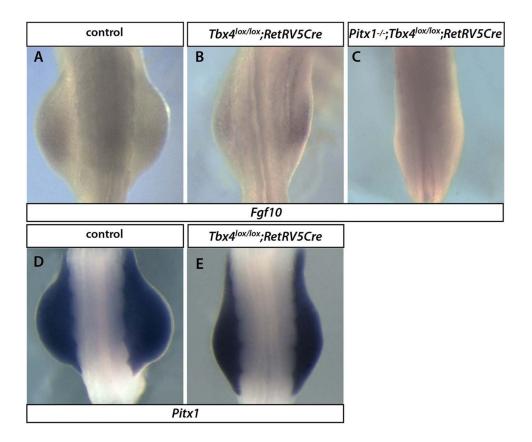
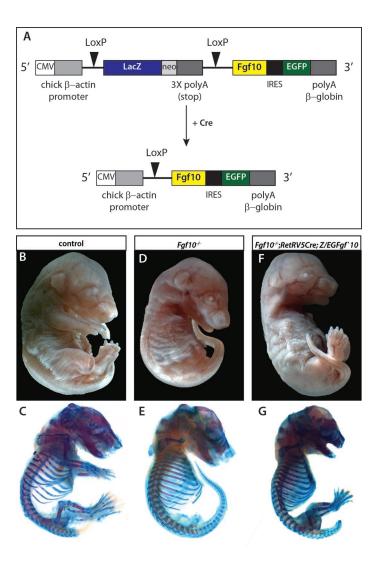
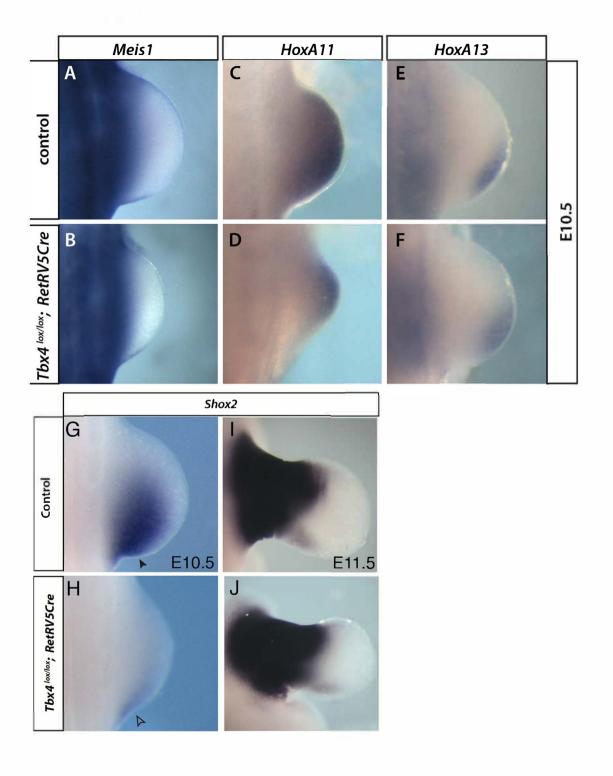


Fig. S2. Fgf10 expression is regulated by dual inputs from Tbx4 and Pitx1. Whole mount in situ hybridisation showing Fgf10 expression in E10.25 hindlimbs of A, control (n=8), B, Tbx4^{lox/lox};RetRV5Cre (n=3) C, Pitx1^{-/-};Tbx4^{lox/lox};RetRV5Cre (n=2) D,E Pitx1



Suppl. Figure3

Fig. S3. The Z/EGFgf10 transgene is able to fully rescue hindlimb outgrowth in Fgf10^{-/-} embryos when activated by the RetRV5Cre line. A, Cre recombinase recombines the LoxP flanked cassette to enable transgenic Fgf10 transcription in the Z/EGFgf10 construct. The Z/ EGFgf10 construct was produced using the Z/EG transgenic backbone (Novak et al. 2000) and contains a β-Geo cassette, flanked by two LoxP sites (black triangles) downstream of the chick β-actin promoter (light grey rectangle) and the CMV enhancer (white rectangle). The β-Geo cassette contains LacZ sequence (blue rectangle) and a neomycin resistance gene (neo), as well as a 3x polyadenylation sites (dark grey) that serve as a transcriptional stop signal. Located 3' of the β -Geo cassette is the Fgf10 cDNA (yellow rectangle) and an internal ribosome entry site (IRES, black rectangle), eGFP (green rectangle) and β-globin polyadenylation signal (dark grey rectangle). In the absence of Cre recombinase, CAGGS drives transcription of β -Geo. Following cre excision, the β -Geo cassette is removed and the Fgf10/IRES/eGFP cassette comes under the control of the CAGGS promoter. **B,D,F**, E17.5 embryos and C,E,G, complementary skeletal preparation of B,C control (n=1), D,E Fgf10^{-/-} (n=1) and F,G Fgf10-/-; RetRV5Cre; Z/EGFgf10 rescued embryos (n=2). Hindlimb formation is completely rescued in the Fgf10 mutant by activation of the Fgf10 transgene.



control (n=3) (I) and $Tbx4^{lox/lox}$; RetRV5Cre mutant (n=2) (G).

Fig. S4. *Meis1*, *Hoxa11* and *Hoxa13* are expressed in *Tbx4* mutant hindlimb buds. A-F Whole mount *in situ* hybridisation of E10.5 hindlimb buds. Dorsal views. Expression of the proximal segment (stylopod) marker *Meis1* (n=3) (A) is expressed in the *Tbx4* lox/lox/*RetRV5Cre* mutant (n=3) (B). Medial segment (zeugopod) marker *Hoxa11* (n=3) (C) is expressed in the *Tbx4* mutant (n=2) (D). At E10.5 *Hoxa11* is expressed both medially and distally. Distal segment (autopod) marker *Hoxa13* (n=3) (E) is expressed in the *Tbx4* mutant (n=3) (F). *Shox2* is downregulated in the *Tbx4* mutant at E10.5 Whole mount *in situ* hybridisation of E10.5 control (n=4) (G) and *Tbx4* lox/lox/, *RetRV5Cre* mutant (n=3) (H). Robust expression in control (solid arrow) contrasts with reduced expression in the *Tbx4* conditional mutant (hollow arrow). *Shox2* expression is restricted to the proximal limb bud at E11.5 in

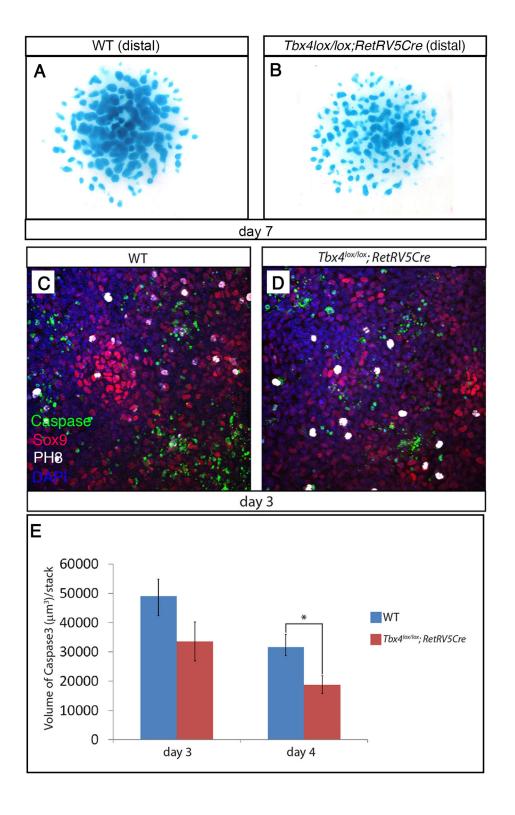
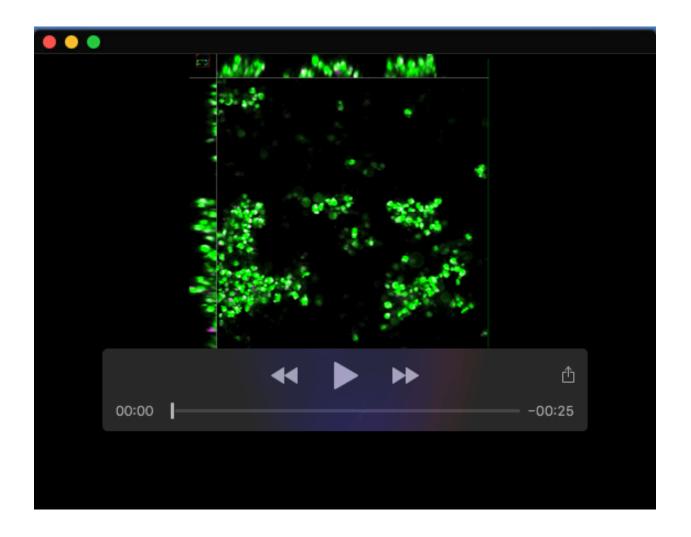
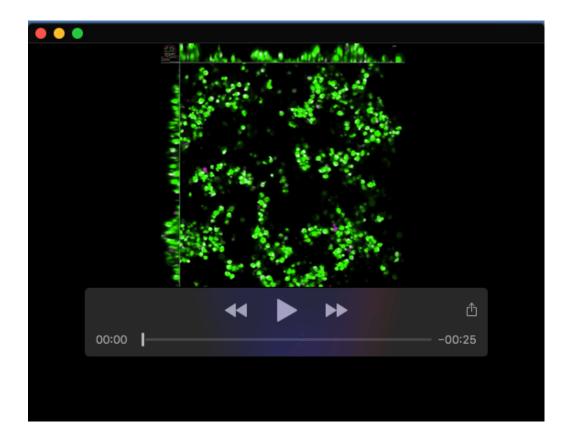


Fig. S5. Cell death is not responsible for the decreased number chondroprogenitors detected over time. A,B Alcian Blue staining of cartilage nodule formation after 7 days of culture in micromass cultures established from distal cells of (A) Wild type hindlimbs (n=6), (B) Tbx4lox/lox; RetRV5Cre hindlimbs (n=6). C,D Extended focus confocal stacks of immunostained proximal micromass culture after 3 days culture. Anti-Caspase (green), anti-Sox9 (red), phospho-histone H3 (white) and nuclear staining (DAPI-blue) of control (C) and Tbx4lox/lox; RetRV5Cre (D) proximal micromass. (E) Histogram showing mean \pm s.e.m. volume in μ m³ of Caspase 3 staining measured in a stack at 20x magnification of control (blue) and Tbx4lox/lox;RetRV5Cre (red) proximal micromass after 3 and 4 days of culture. No statistical difference is observed between the two conditions after Student t-test (P>0.05); n=5. Standard errors are shown.



Movie 1. Time lapse movie of a control micromass culture stained with Cell Tracker dye. xyz views of Z-scan stacks imaged every 5 minutes during an 84hr culture. 20X magnification.



Movie 2. Time lapse movie of a *Tbx4*^{lox/lox}; *RetRV5Cre* micromass culture stained with Cell Tracker dye. xyz views of Z-scan stacks imaged every 5 minutes during an 84hr culture. 20X magnification.