

Figure S1. All KIF2A isoforms have diffused localization to cytoplasm and nucleus in Neuro2A cells. Neuro2A cells transfected with plasmids expressing pEGFPC1-tagged KIF2A isoforms (KIF2A.1, KIF2A.2 and KIF2A.3) 2 days after plating, fixed with 4% PFA 2 days after transfection and immunostained with anti-GFP (green). Anti-αTubulin visualized microtubules (red) and DAPI stained DNA. Scale bar, 10 µm.



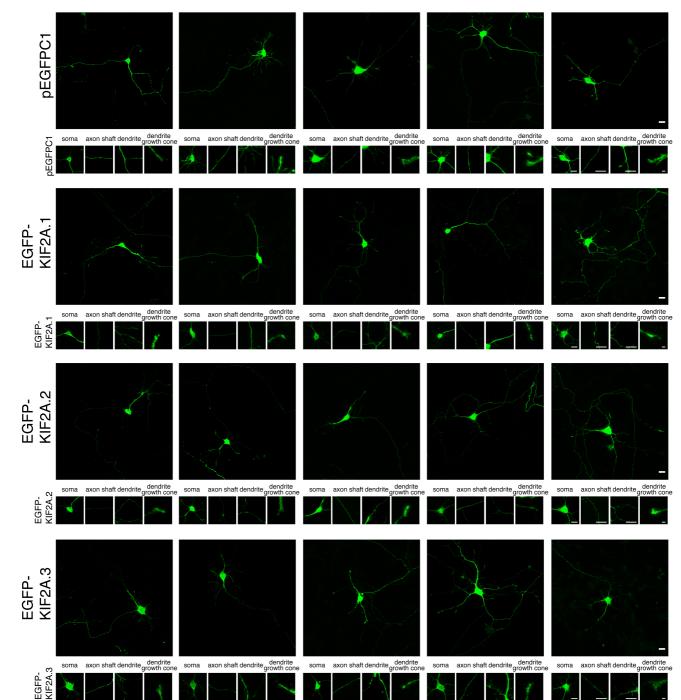


Figure S2. All KIF2A isoforms are localized to soma, axon and dendrites in primary cortical neurons. E14.5 primary cortical neurons were transfected with pEGFPC1 backbone or one of the EGFP expressing KIF2A isoforms at 2 DIV, fixed and immunostained against EGFP at 4 DIV. Digital zoom was applied to visualize localization of KIF2A isoforms in soma, axon shaft, dendrite and dendrite growth cone of each transfected neuron. 5 representative images for each condition were shown in the figure. Scale bar, 10 μ m; scale bar for dendrite growth cone, 1 μ m.

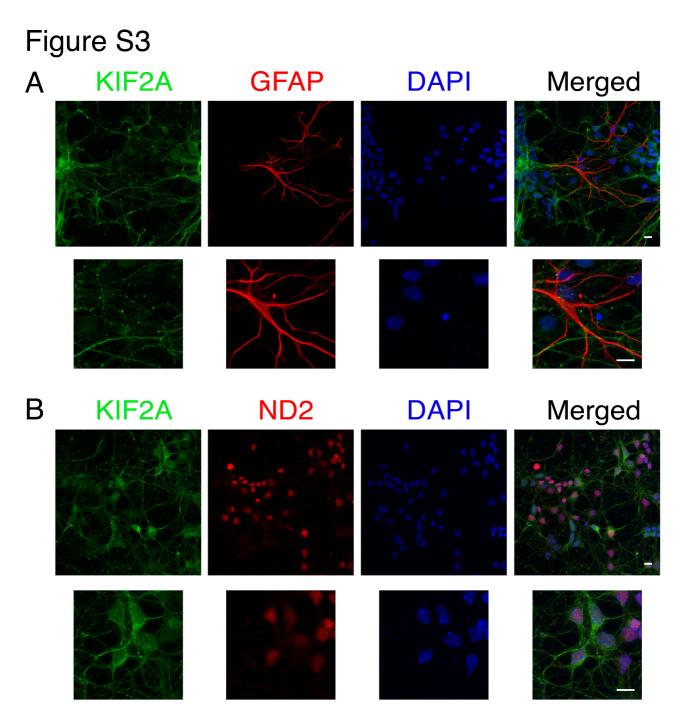


Figure S3. KIF2A is expressed predominantly by neurons but not by glia cells. A and B. E14.5 primary cortical culture was fixed at 4 DIV and stained with KIF2A (green) either anti-GFAP antibody (red) to stain glia cells (A) or anti-NEUROD2 antibody (red) to label neurons (B). Digital zoom was applied to visualize localization of KIF2A. Scale bar, 10 μm.

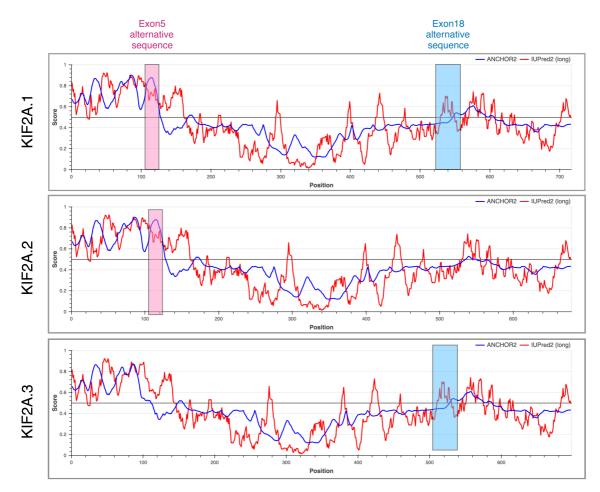


Figure S4. Measure of disorder in protein regions of individual KIF2A isoforms. A measure of protein disorder is plotted as a function of amino acid sequence of individual KIF2A isoforms. The IUPred2 and ANCHOR2 algorithms are used (Dosztányi, 2018). Alternative sequences are labeled in pink (exon5) and blue (exon18).

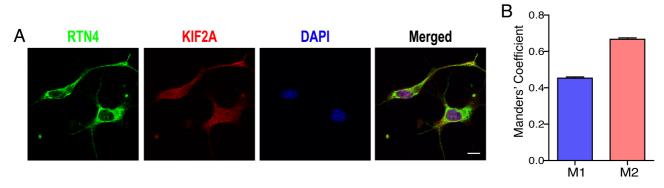


Figure S5. KIF2A and RTN4 are partially co-localized in Neuro2A cells. A. Immunofluorescence staining of RTN4 (green), KIF2A (red), DNA (blue) in Neuro2A cells. Scale bar, 10µm. **B.** Quantification of co-localization of KIF2A with RTN4A using Manders' coefficient (Dunn et al., 2011; Manders et al., 1993; Zinchuk et al., 2008). M1: Fraction of KIF2A fluorescence overlapped with RTN4 fluorescence. M2: Fraction of RNT4 fluorescence overlapped with KIF2A fluorescence. n=219, data is represented as bar graphs. Lines represent S.E.M.

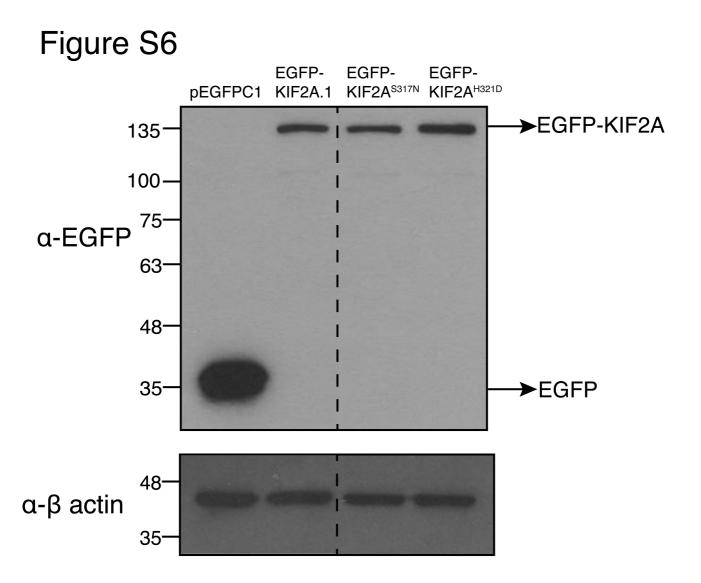


Figure S6. KIF2A mutants (*Kif2a^{S317N}* or *Kif2a^{H321D}*) expressions are similar with wildtype KIF2A and mutants do not affect protein instability. Immunoblotting with anti-EGFP demonstrating the levels of EGFP-tagged KIF2A isoforms and KIF2A mutants. B-ACTIN is used as a loading control.

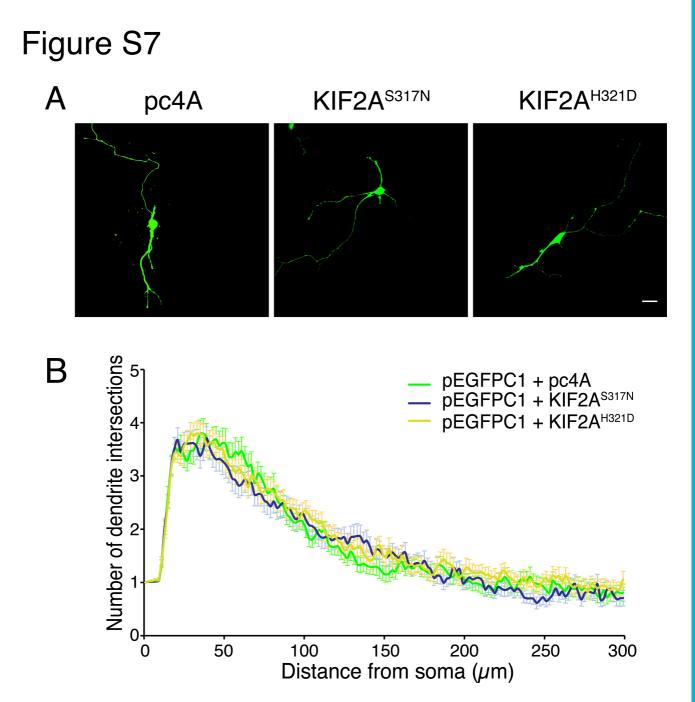
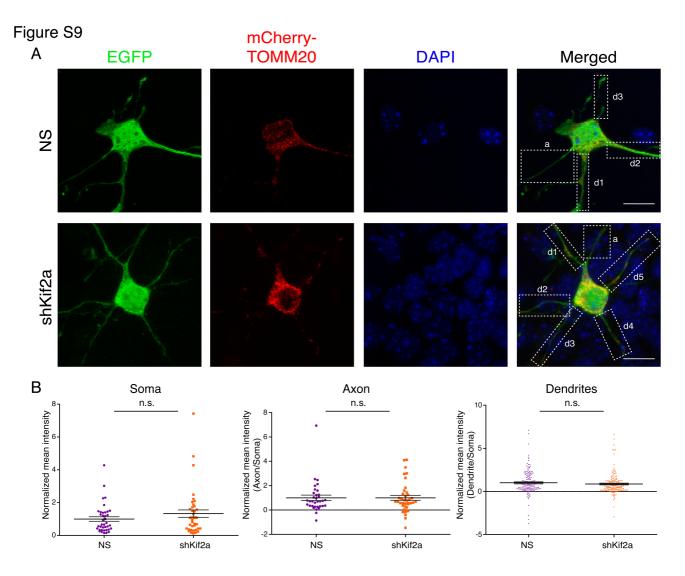


Figure S7. Expression *Kif2a*^{S317N} or *Kif2a*^{H321D} in wild-type neurons do not affect dendrite arborization. A. Representative images of E14.5 primary cortical neurons co-transfected with pEGFPC1 along with *Kif2a* mutants. Scale bar, 10 μ m. B. Dendrite development was quantified by Sholl analysis (Ferreira et al., 2014; Schindelin et al., 2012). n=70 for each condition derived from three separate neuronal cultures. Bars represent S.E.M. Unpaired two-tailed *t* test determined the p value. p<0.05.

	Soma			Dendrite			Axon		
	mCherry- TOMM20	EGFP	Colocalization	mCherry- TOMM20	EGFP	Colocalizatio	mCherry- TOMM20	EGFP	Colocalization
pEGFPC1							1		
EGFP- KIF2A.1				1					1 and
EGFP- KIF2A.2			L.	1	X	-			
EGFP- KIF2A.3				K			1	/	1 Com
EGFP- KIF2A ^{S317N}	\bigcirc				financial and the second	Contraction In the Contraction of the Contraction o			
EGFP- KIF2A ^{H321D}	Q			À	J				

Figure S8. Co-localization of different KIF2A isoforms and *Kif2a* patient mutants with mitochondria in soma, axon and dendrites of primary cortical neurons. Magnified images of primary cortical neurons co-transfected with pEGFPC1-tagged three *Kif2a* isoforms and *Kif2a* disease mutants (*Kif2a*^{S317N} or *Kif2a*^{H321D}) along with mCherry-TOMM20-N-10. Co-localization (white) of KIF2A (EGFP, blue) with mitochondria (mCherry, red) in soma, axon and dendrite was shown in separate images by co-localization threshold tool in ImageJ. Scale bar, 1 μ m.



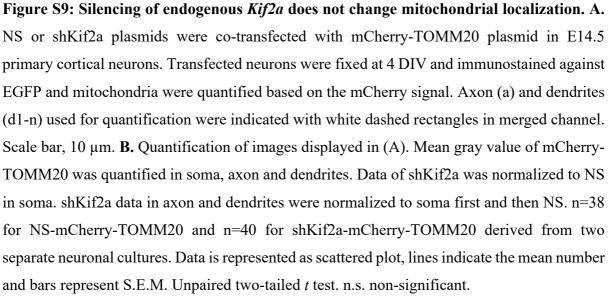


Table S1. Peptide Spectra Match Counts for BioID-mass spectrometry hits for individual KIF2A isoforms

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- Dosztányi, Z. (2018). Prediction of protein disorder based on IUPred. 27(1), 331-340. doi:10.1002/pro.3334
- Dunn, K. W., Kamocka, M. M., & McDonald, J. H. (2011). A practical guide to evaluating colocalization in biological microscopy. *300*(4), C723-C742. doi:10.1152/ajpcell.00462.2010
- Ferreira, T. A., Blackman, A. V., Oyrer, J., Jayabal, S., Chung, A. J., Watt, A. J., Sjöström, P. J., & van Meyel, D. J. (2014). Neuronal morphometry directly from bitmap images. *Nature Methods*, 11(10), 982-984. doi:10.1038/nmeth.3125
- Manders, E. M. M., Verbeek, F. J., & Aten, J. A. (1993). Measurement of co-localization of objects in dual-colour confocal images. *169*(3), 375-382. doi:10.1111/j.1365-2818.1993.tb03313.x
- Schindelin, J., Arganda-Carreras, I., Frise, E., Kaynig, V., Longair, M., Pietzsch, T., Preibisch, S.,
 Rueden, C., Saalfeld, S., Schmid, B., Tinevez, J.-Y., White, D. J., Hartenstein, V., Eliceiri, K.,
 Tomancak, P., & Cardona, A. (2012). Fiji: an open-source platform for biological-image
 analysis. *Nature Methods*, 9(7), 676-682. doi:10.1038/nmeth.2019
- Zinchuk, V., & Zinchuk, O. (2008). Quantitative Colocalization Analysis of Confocal Fluorescence Microscopy Images. *39*(1), 4.19.11-14.19.16. doi:10.1002/0471143030.cb0419s39