

Fig. S1. Characteristic F-actin patterns in diverse epithelial organs. (A) F-actin (TexasRed-phalloidin) images of explant cultures of epithelial organs. Yellow dashed lines present the outline of epithelial buds. Yellow arrowheads present regions of high actin signals. (B) F-actin intensity of single cells of epithelial buds. (C) Percentage of epithelial cells in the different F-actin intensity. n = 50 cells per group. (D) Phase contrast (upper) and TexasRed-phalloidin (lower) images of developing epiSMG cultures for 48 h. (E) TexasRed-phalloidin and immunolabelled phosphorylated myosin (pMyosin) images of epiSMG culture. The white arrow indicates the region use for the intensity plot (panel F). (F) Intensity plot of F-actin and phosphorylated myosin signals. (G) The images of TexasRed-phalloidin (magenta), pMyosin (green) and DAPI (blue) of sectioned SMG cultures (4 µm thickness). (H) Intensity of pMyosin signals in apical and basal part of peripheral epithelial cells in panel G (n = 20 cells). Paired t-test. Mean \pm STD. (I) Morphological changes in mouse embryonic SMG (upper) and epiSMG (lower) cultures upon 100 nM cytochalasin D. Scale bars: 50 µm.

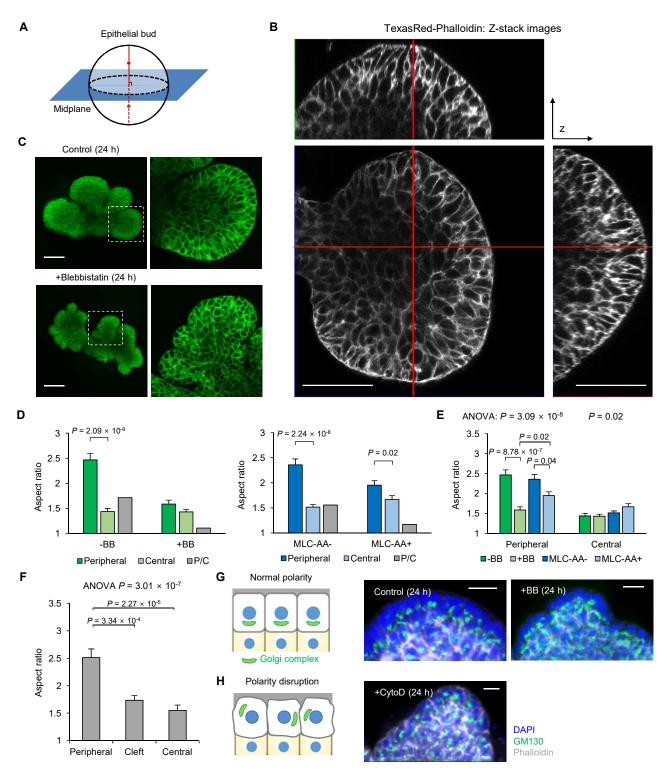


Fig. S2. Changes in diverse cell parameters upon perturbation of actomyosin contractility (A) Schematic representation of the optically sectioned imaging plane for single cell anlaysis. (B) Z-stack images of TexasRed-phalloidin in epiSMG culture (E13 + 24 h). 1 μ m section, z: 71.00 μ m. (C) Immunolabelled images of E-cadherin (green) in E13 eSMG culture (left) and blebbistatin-treated (right) cultures. White dashed regions indicate the enlarged area. (D and E) Aspect ratio of cells in peripheral (P) and central (C) region of epithelial buds. Mean ± sem. n = 30 cells (-BB and +BB) and 34 cells (MLC-AA- and MLC-AA+). (F) Aspect ratio of cells in different regions of epithelial buds. Mean ± sem. n = 20 cells per group. (G) Apically polarized location of the Golgi complex (immunolabelled by GM130) in control and blebbistatin-treated eSMG cultures (E13). (H) Disrupted polarization of the Golgi complex in eSMG cultures treated with low-concentration cytochalasin D. Scale bars: 50 µm (B, C), 20 µm (G, H)

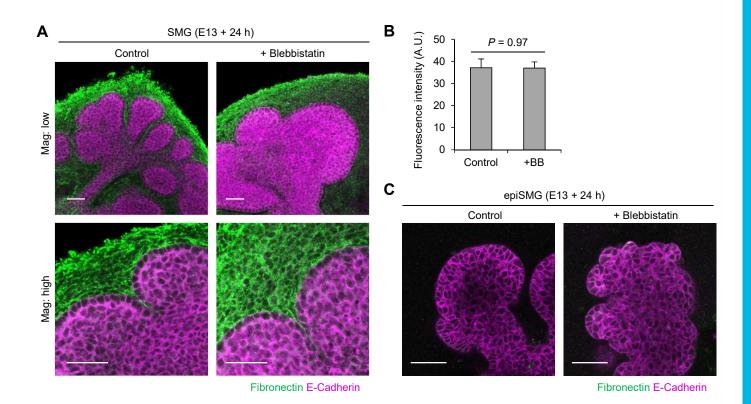


Fig. S3. Fibronectin patterns of SMG upon blebbistatin treatment. (A) Immunolabeled signals of fibronectin (green) and E-cadherin (magenta) of developing SMG cultures. (B) Quantified results of fluorescence signals of fibronectin of developing SMG cultures. Mean \pm sem. n = 12 regions per group. (C) Immunolabeled signals of fibronectin (green) and E-cadherin (magenta) of developing epiSMG cultures. Scale bars: 50 µm.

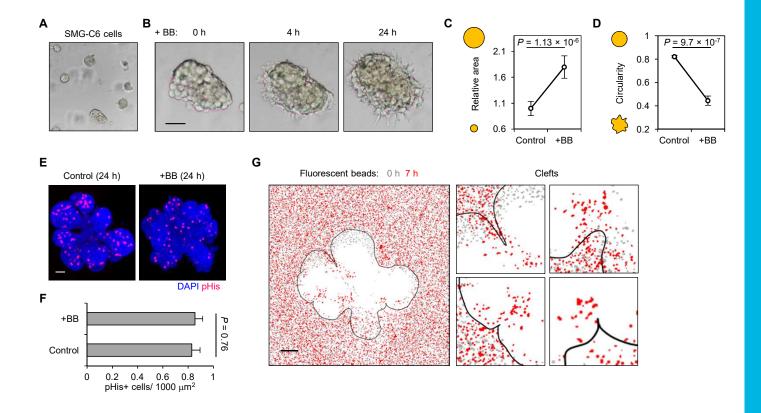
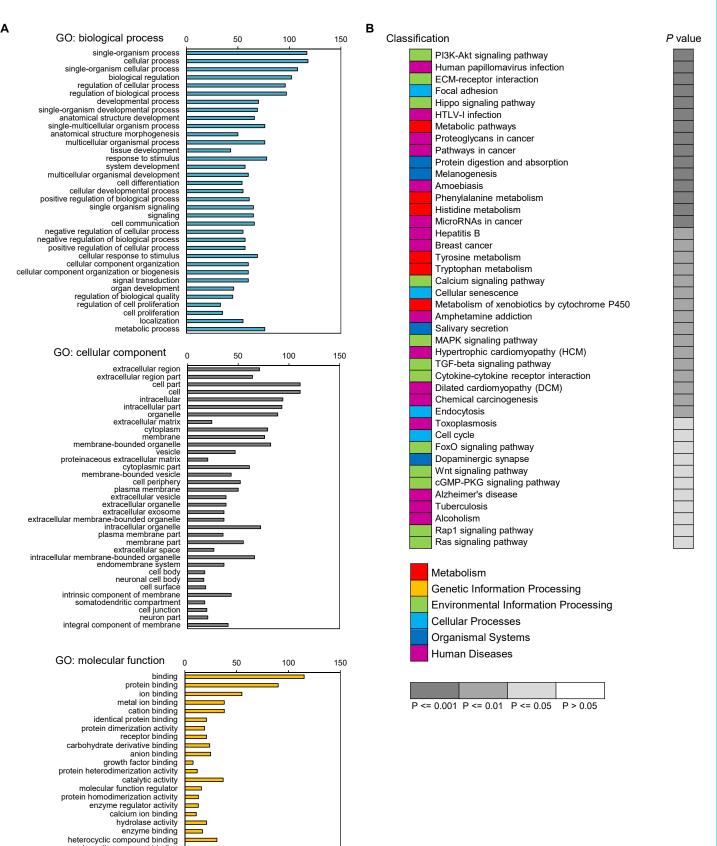


Fig. S4. The mechanism of force transmission in branching morphogenesis (A) Phase contrast image of salisphere culture using the SMG-C6 cell line (rat SMG acinar cell line). (B) Time-dependent morphological changes of salisphere cultures. (C and D) Quantified value of the relative area (C) and circularity (D) of salisphere cultures. Mean \pm sem. n = 12 cultures per group. (E) Immunolabelled phosphorylated histone H3 signals (pHis, red) in developing eSMG cultures (E 13) without (Control) or with blebbistatin treatment (+BB). (F) Quantified pHis+ cell density (per 1,000 µm³) in developing eSMG cultures (E 13). Mean \pm sem. n = 5 cultures per group. (G) The location changes in fluorescent beads around developing epiSMG culture. Right subpanels show enlarged pericleft regions. Scale bars: 50 µm.

organic cyclic compound binding cytoskeletal protein binding small molecule binding cell adhesion molecule binding

glycosaminoglycan binding protein tyrosine kinase activity heparin binding purine ribonucleotide binding

purine nucleotide binding enzyme activator activity ribonucleotide binding transmembrane RTK activity substrate-specific channel activity



Development • Supplementary information

Fig. S5. RNA-seq data analysis (A) The top 35 gene ontology annotations with high significance (Control versus Blebbistatin, P < 0.005). The horizontal axis indicates DEG count. (B) KEGG enrichment map score (based on *P* value). The lists are arranged by descending significance (ascending *P* values). The data can be accessed at NCBI GEO accession number GSE137928.

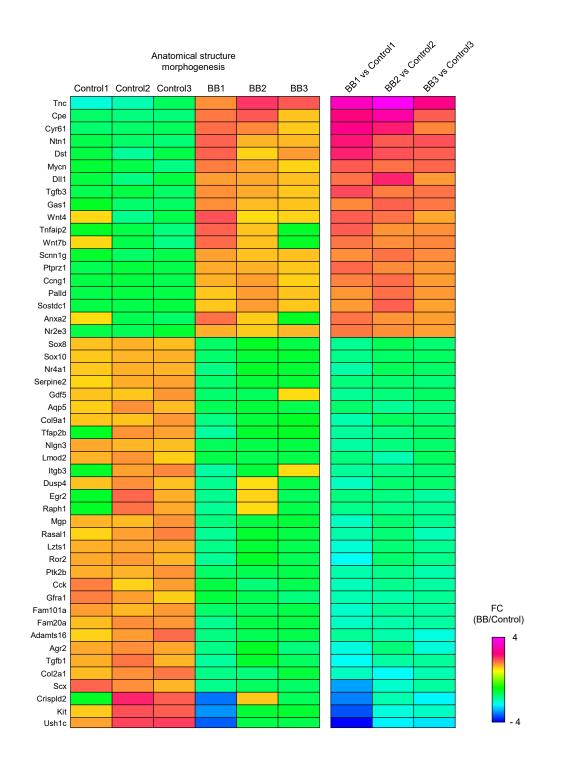


Fig. S6. Heatmap showing total DEGs in the anatomical structure morphogenesis annotation. DEGs are arranged by descending fold-change (FC).

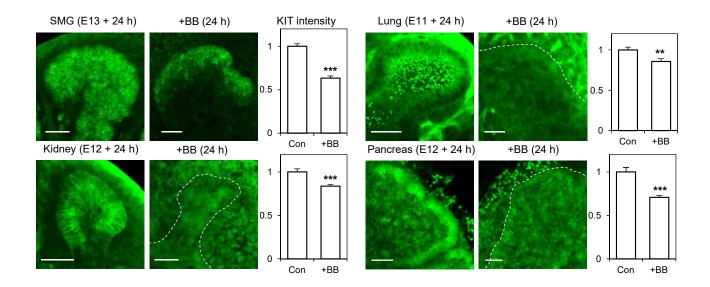


Fig. S7. Immunolabelled images of KIT expression in epithelial organs (images) and quantified intensity of KIT signals (bar graphs). Mean \pm sem. n = 15 buds per group. White dashed regions represent the outline of epithelial region. *, P < 0.05; **, P < 0.01; ***, P < 0.001.

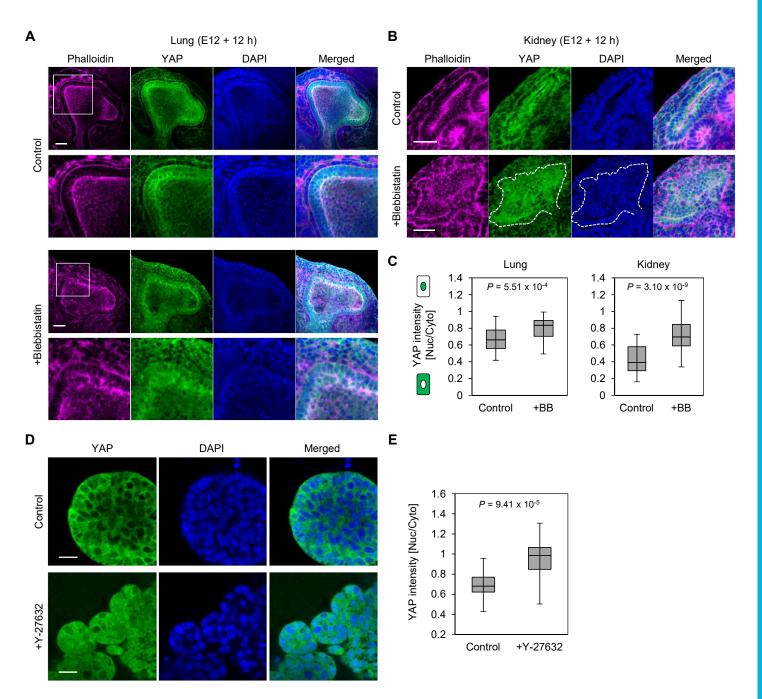


Fig. S8. YAP localization changes upon the perturbation of actomyosin contractility. (A and B) Immunolabelled images of phalloidin (magenta), YAP (green), DAPI (blue) in lung (A) and kidney (B) explant cultures in the control and blebbistatin-treated groups. (C) Relative intensity of nuclear and cytoplasmic YAP signals in control (n = 35 cells) and blebbistatin (n = 35 cells)-treated lung and kidney cultures. Median \pm max/min. (D) Immunolabelled images of YAP (green) patterns and DAPI (blue) signals in control and Y-27632-treated E13 SMG cultures (24 h). (E) Relative intensity of nuclear and cytoplasmic YAP signals in control (n = 20 cells)-treated SMG cultures. Median \pm max/min. Scale bars: 50 µm (A, B), 20 µm (D).

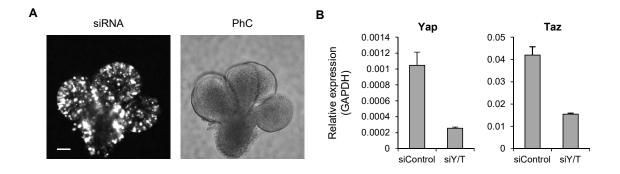
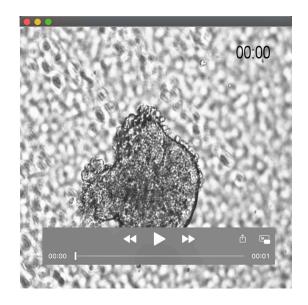
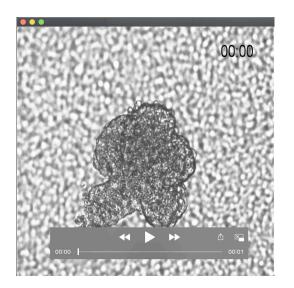


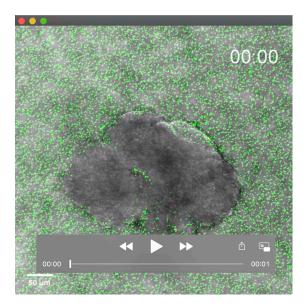
Fig. S9. Regulation of YAP/TAZ expression in epiSMG cultures. (A) The expression pattern of siRNA conjugated with fluorescent dye in E13 eSMG cultures. The transfected cultures were incubated for 24 h. PhC: phase contrast. (B) Nested qPCR results of the changes in Yap and Taz expression in eSMG cultures cotransfected with siYAP and siTAZ (siY/T). Mean \pm sem. n = 3. Scale bar: 50 µm.



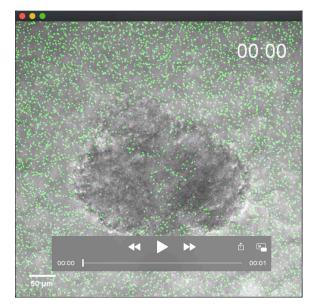
Movie 1. Branching morphogenesis of eSMG culture. Time-lapse phase contrast images of the initial branching process of eSMG cultures (E13). The images were acquired for 16 h with 10-min intervals.



Movie 2. Branching morphogenesis of eSMG culture treated with blebbistatin. Time-lapse phase contrast images of the initial branching process of eSMG cultures (E13) treated with 50 µM blebbistatin. The images were acquired for 16 h with 10-min intervals.



Movie 3. 3-D traction force analysis (Control). Time-lapse images of the initial branching process of eSMG cultures (E13). The isolated epithelial rudiments were embedded in Matrigel-DMEM/F-12 mixture containing 5% fluorescent microbeads (2.0 µm diameter). The images were acquired for 7 h with 10-min intervals.



Movie 4. 3-D traction force analysis (+Blebbistatin). Time-lapse images of the initial branching process of eSMG cultures (E13) treated with 50 µM blebbistatin. The isolated epithelial rudiments were embedded in Matrigel-DMEM/F-12 mixture containing 5% fluorescent microbeads (2.0 µm diameter). The images were acquired for 7 h with 10-min intervals.

Table S1. Lists of all DEGs along with their FC values. Differentially expressed transcripts are satisfied with |fold change| \geq 1.5 and independent t-test raw *p* <0.05.

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