

Fig. S1 (related to Figs. 1, 2, 3). Wnt/β-catenin signaling is activated in retinal ECs and mural cells, but not in astrocytes. A) Analyses of a P14 WT retinal single cell RNAseq database (see Results and Materials and Methods) to explore expression profiles of downstream Wnt/β-catenin targets (*Lef1*, *Apcdd1*, *Axin2*) in endothelial cells (ECs), pericytes (PCs) and astrocytes. B-E) P10 WT retinal flatmounts were stained for Lef1 and Pdgfrβ (A= artery, V= vein, C= capillary). Solid arrowheads point to Lef1+ mural cells. Dotted bar graph shows number of Lef1+ mural cells in unit length for each vessel type (n=4 animals). F-K) Fluorescence *in situ* hybridization of P10 WT retinal sections with antisense probe against *Apcdd1* mRNA followed by immunostaining for Caveolin-1 or Pdgfrβ proteins. Solid arrowheads and empty arrowheads point to localization of *Apcdd1* mRNA inside and outside the corresponding cells marker, respectively. Student's t-test: **p<0.02, NS: not significant. Graphs show mean +/- SEM. Scale bars: B-D = 41 μm.

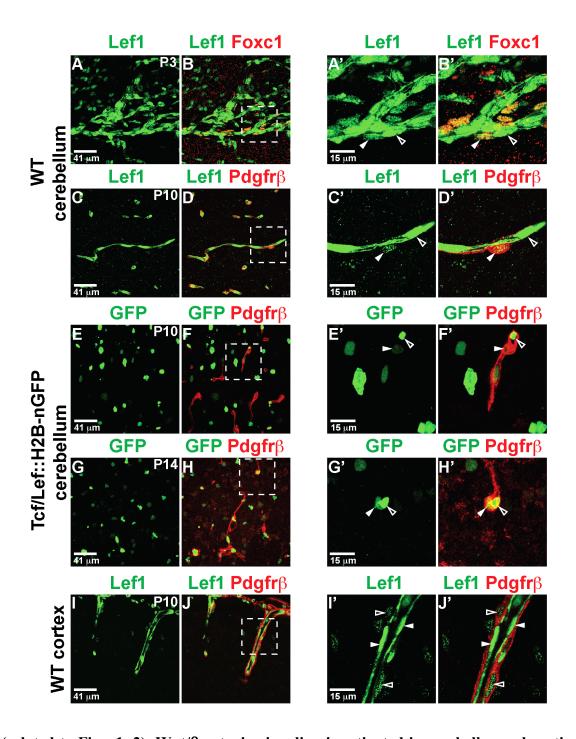


Fig. S2 (related to Figs. 1, 2). Wnt/β-catenin signaling is activated in cerebellar and cortical mural cells. A-B') P3 WT cerebellar sections were stained for Lef1 and Foxc1. C-D') P10 WT cerebellar sections were stained for Lef1 and Pdgfrb. E-H') P10 and P14 Tcf/Lef::H2B-nGFP cerebellar sections were stained for GFP and Pdgfrb. In all images empty arrowheads point to Lef1⁺ or GFP⁺ ECs, whereas solid arrowheads point to Lef1⁺ or GFP⁺ mural cells. Scale bars: A-J = 41 μm; A'-J' = 15 μm.

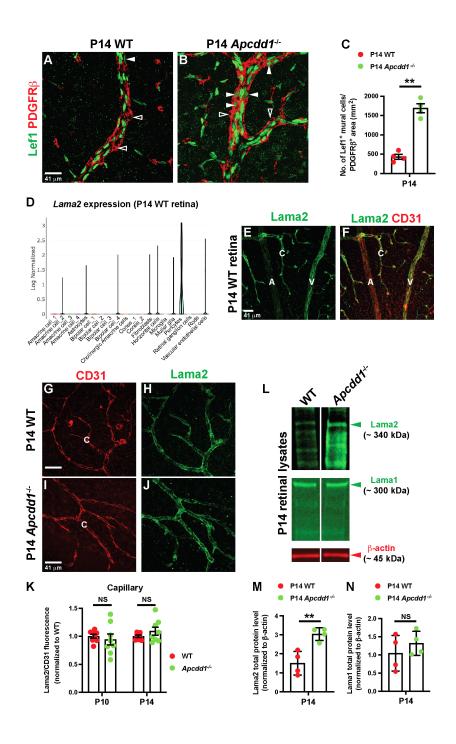


Fig. S3 (related to Fig. 3). Lama2 (*Lama2*) is predominantly expressed in retinal PCs, but not in ECs, and is upregulated in the *Apcdd1*-/- retina. A-C) P14 WT and *Apcdd1*-/- retinal flat-mounts were stained

for Lef1 and Pdgfrβ. Solid arrowheads point to Lef1⁺ mural cells, empty arrowheads point to Lef1⁻ mural cells. Dotted bar graph shows the number of Lef1⁺ mural cells (n=4 animals/genotype). **D)** Expression profile of *Lama2* mRNA in P14 WT retinal cells from the published retina single cell RNAseq database. **E, F)** P14 WT retinal flat mounts were stained for CD31 and Lama2 proteins. A= artery, V= vein, C= capillary. **G-K)** P14 WT and *Apcdd1*^{-/-} retinal flat-mounts were labelled for Lama2 and CD31, showing retinal capillaries (C). Dotted bar graphs show the ratio of Lama2/CD31 M.F.I in capillaries normalized to the WT average values at P10 and P14 (8 fields of capillary network from n=4 mice/genotype). **L)** P14 WT and *Apcdd1*^{-/-} whole retinal lysates were probed for Lama1 or Lama2 (green bands) and β-actin (red bands) by western blot. **M, N)** Quantification of Lama1 and Lama2 protein levels (normalized to β-actin) by western blotting (n=4 samples/group). Student's t-test: **p<0.02, NS= not significant. Graphs show mean+/- SEM (C, K) or mean +/- SD (M, N). Scale bars: 41 μm.

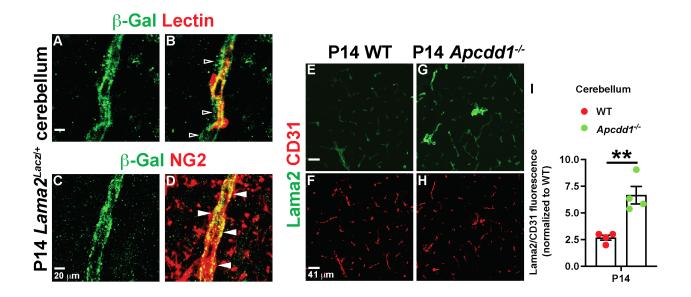


Fig. S4 (related to Fig. 3). Mural cell-derived Lama2 expression and deposition are upregulated in the *Apcdd1*-/- cerebellar vascular basement membrane. A-D) P14 $Lama2^{Lacz/+}$ cerebellar sections were stained for β-galactosidase (β-Gal) and either Lectin (A-B) or NG2 (C-D). Empty arrowheads point to β-Gal- ECs. Solid arrowheads point to β-Gal+ mural cells. E-I) P14 WT and Apcdd1-/- cerebellar sections were immunolabelled for Lama2 and CD31. Dotted bar graph shows the ratio of Lama2/CD31 M.F.I in, normalized to the WT values (n=4 mice/genotype); Students' t-test: **p<0.02. Graph shows mean+/-SEM. Scale bars: A-D= 20 μm; E-H= 41 μm.

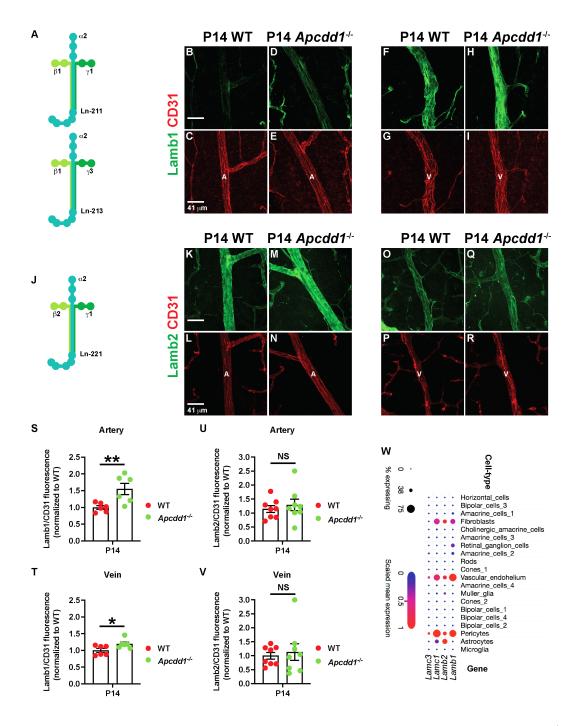


Fig. S5 (related to Fig. 3). Laminin-211 is likely the most affected isoform in the *Apcdd1*^{-/-} retina. A) Schematic diagram of two Laminin heterotrimers (211 and 213) containing α 2 and β 1. B-I) P14 WT and *Apcdd1*^{-/-} retinal flat-mounts were labelled for Lamb1 and CD31. J) Schematic diagram of one α 2- and β 2-

containing Laminin heterotrimer (221). **K-R)** P14 WT and *Apcdd1*-- retinal flat-mounts were labelled for Lamb2 and CD31. **S, T)** Dotted bar graphs show the ratio of Lamb1/CD31 M.F.I in arteries and veins at P14 normalized to the WT average values (6 arteries or veins analyzed from n=3 mice/group). **U, V)** Dotted bar graphs show the ratio of Lamb2/CD31 M.F.I in arteries and veins at P14 normalized to the WT average values (8 arteries and veins analyzed from n=4 mice/group). **W)** Analyses of the published P14 WT retinal single cell RNAseq database (see Results and Materials and Methods sections) to explore the expression profiles of *Lamb1*, *Lamb1*, *Lamc1* and *Lamc3*. A= artery, V= vein. Student's t-test: *p<0.05, **p<0.02, NS= not significant. Graphs show mean +/- SEM. Scale bars: 41 μm.

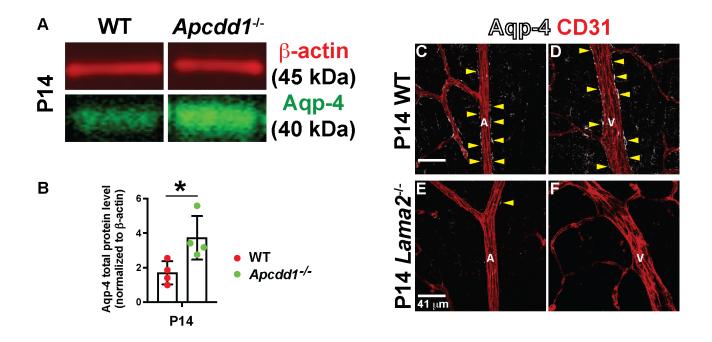


Fig. S6 (related to Fig. 6). Increased Aqp-4 in the *Apcdd1*^{-/-} retina and decreased Aqp-4⁺ astrocyte endfeet polarization in the *Lama2*^{-/-} retina. A) P14 WT and *Apcdd1*^{-/-} whole retinal lysates were probed for Aqp-4 (green bands) and β-actin (red bands) by western blot. B) Quantification of Lama1 and Lama2 protein levels (normalized to β-actin) by western blotting (n=4 samples/group). Student's t-test: *p<0.05. Graphs show mean +/- SD. C-F) P14 WT and *Lama2*^{-/-} retinal flat-mounts were labelled for Aqp-4 and CD31. Yellow arrowheads point at Aqp-4⁺ astrocyte endfeet around retinal blood vessels. A= arteries and V= veins. Scale bars: 41 μm.

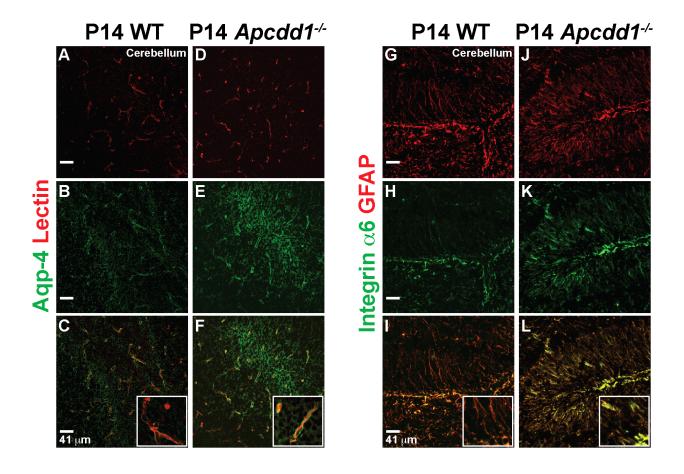


Fig. S7 (related to Figs. 6 and 8). Increased expression of Aqp4 in astrocyte endfeet and Integrin α6 in *Apcdd1*-/- astrocytes in the cerebellum. A-F) P14 WT and *Apcdd1*-/- cerebellar sections were labelled for Aqp-4 and Lectin. Insets show higher magnification images of single vessels. **G-L**) P14 WT and *Apcdd1*-/- cerebellar sections were labelled for Integrin α6 and GFAP. Insets show higher magnification images. Scale bar: 41 μm.

Table S1 (related to Figs. 2, 3, 6 and 8). List of significantly differentially expressed genes between wild-type and *Apcdd1*-/- retinas at P10 and P14. Negative values mean that the gene is lower in *Apcdd1*-/- compared to wild-type retinas, and positive values mean that the gene is higher in *Apcdd1*-/- compared to wild-type retinas.

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Table S2 (related to Fig. 2). List of significant differentially expressed putative endothelial genes between wild-type and *Apcdd1*^{-/-} retinas at P10 and P14. The genes are ranked by log₂Fold Change from the lowest to the highest value. Negative values mean that the gene is lower in *Apcdd1*^{-/-} compared to wild-type retinas, and positive values mean that the gene is higher in *Apcdd1*^{-/-} compared to wild-type retinas.

Click here to download Table S2

Table S3 (related to Figs. 2 and 3). List of significant differentially expressed putative pericyte genes between wild-type and *Apcdd1*-/- retinas at P10 and P14. The genes are listed by log₂Fold Change from the lowest to the highest value. Negative values mean that the gene is lower in *Apcdd1*-/- compared to wild-type retinas, and positive values mean that the gene is higher in *Apcdd1*-/- compared to wild-type retinas.

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Table S4 (related to Figs. 2, 3 and 8). List of significant differentially expressed extracellular matrix genes (ECM) between wild-type and *Apcdd1*-/- retinas at P10 and P14. The genes are listed by log₂Fold Change from the lowest to the highest value. Negative values mean that the gene is lower in *Apcdd1*-/- compared to wild-type retinas, and positive values mean that the gene is higher in *Apcdd1*-/- compared to wild-type retinas.

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Table S5 (related to Fig. 6). List of significant differentially expressed astrocyte maturity genes between wild-type and *Apcdd1*-/- retinas at P10 and P14. The genes are listed by log₂Fold Change from the lowest to the highest value. Negative values mean that the gene is lower in *Apcdd1*-/- compared to wild-type retinas, and positive values mean that the gene is higher in *Apcdd1*-/- compared to wild-type retinas.

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