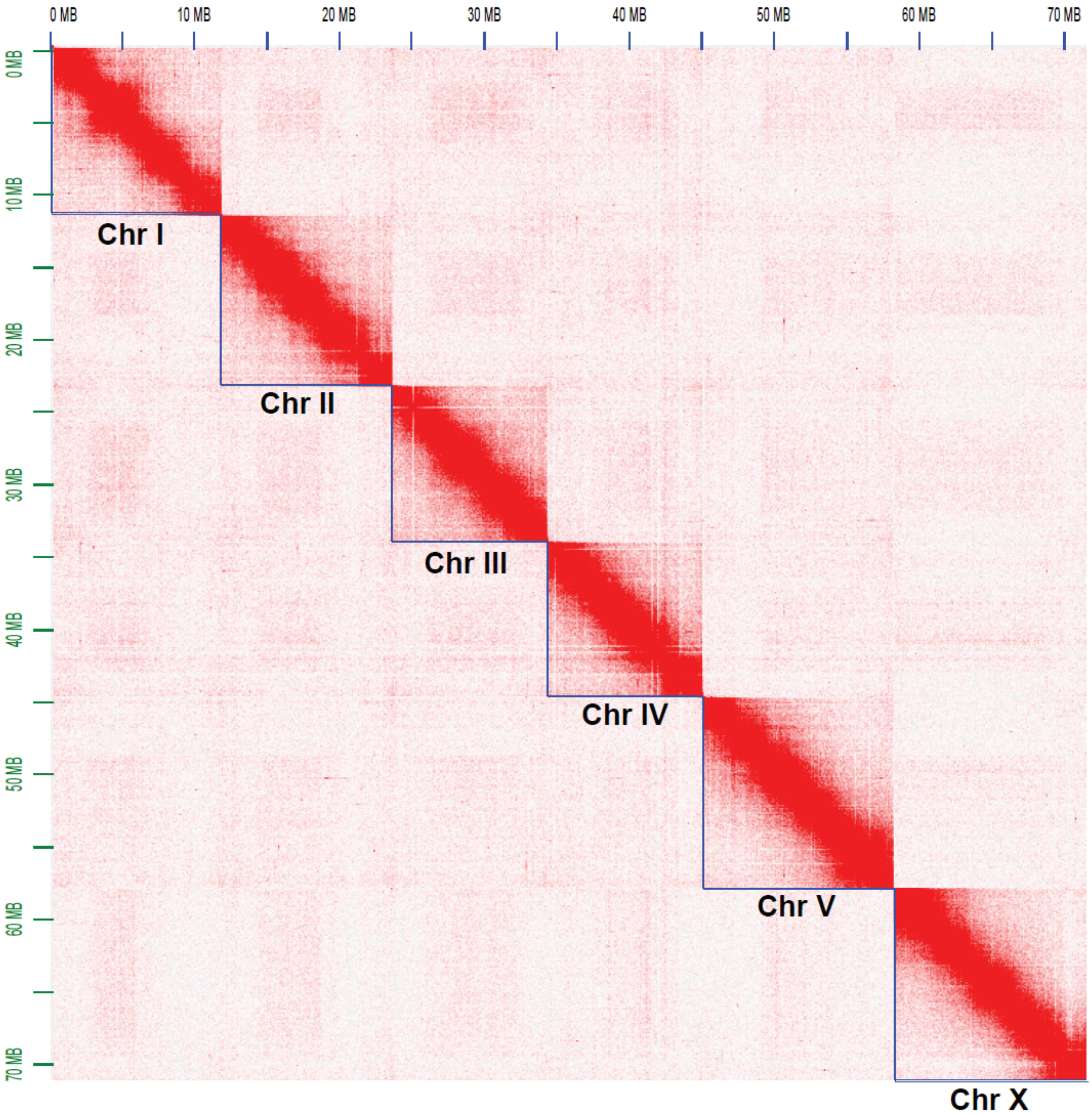

Fig. S1. Hi-C map for *C. angaria* PS1010 sequence indicating linkage groups.

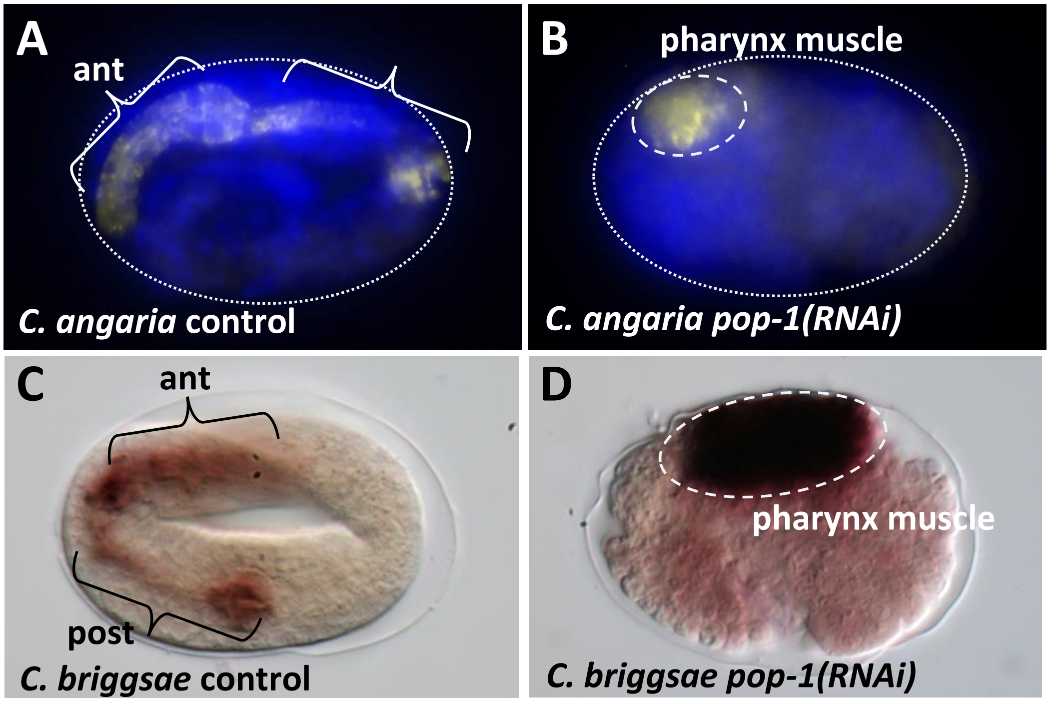

Fig. S2. smiFISH and *in situ* hybridization to detect *myo-2*. (A) Representative PS1010 control 3x stage embryo stained with DAPI (blue) and *Can-myo-2* smiFISH probes (yellow). Staining detects the anterior and posterior pharynx (ant ph and post ph, respectively). 10/10 embryos showed similar staining. (B) Representative arrested *Can-pop-1(RNAi)* embryo stained similarly as in (A). Ectopic *Can-myo-2* expression was not apparent in 20/20 embryos. Because *Can-pop-1(RNAi)* embryos do not elongate, proper morphogenesis of the pharynx does not occur. (C,D) Images from a prior work where ectopic *Cbri-myo-2* tissue was detected in *Cbri-pop-1(RNAi)* using an older *in situ* hybridization protocol (Lin et al., 2009). (D) Extra *Cbri-myo-2* expression is seen because of an E to MS transformation, resulting in extra pharynx muscle. Embryos are approximately 50 μm long.

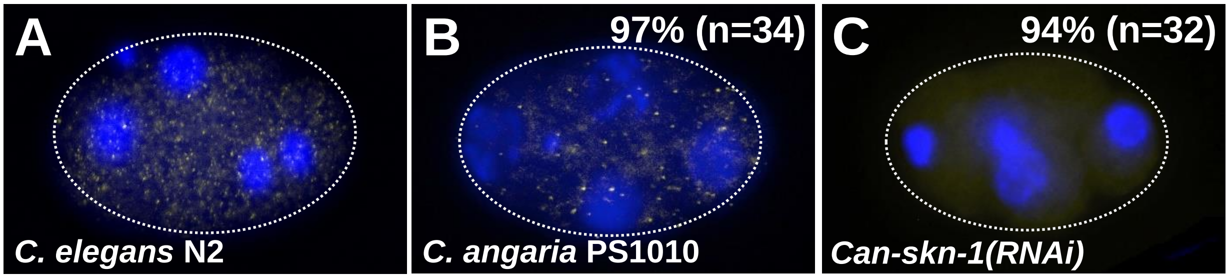

Fig. S3. Expression of *Cel-skn-1* and control for effectiveness of *Can-skn-1(RNAi).* (A) Expression of *C. elegans skn-1* by smiFISH. (B) *Can-skn-1* mRNA was detected in 97% (n=34) of controls. (C) *Can-skn-1* mRNA was absent in 94% (n=32) of *Can-skn-1(RNAi)* embryos.

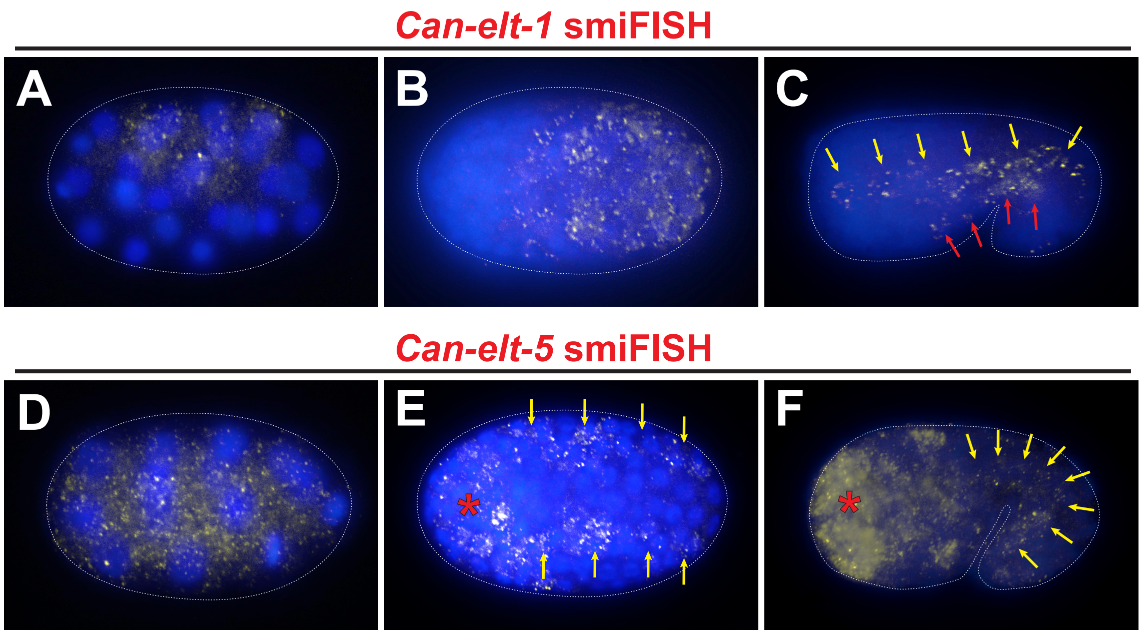

Fig. S4. Expression of *Can-elt-1* (A-C) and *Can-elt-5* (D-F) by smiFISH. (A) Expres-sion of *Can-elt-1* was seen in dorsal hypodermal precursors at gastrulation stage. (B) Dorsal aspect showing widespread expression in hypodermal precursors at late one-fold stage. (C) Expression in lateral seam (yellow arrows) and ventral hypodermis (red arrows) at 1.3-fold stage. (D) Widespread expression of *Can-elt-5* in a 16-cell stage embryo. (E) Dorsal aspect showing expression in anterior cells (*) and lateral hypoder-mal cells (yellow arrows) of late one-fold stage. (F) Anterior expression (*) and lateral hypodermal expression (yellow arrows) in 1.5-fold stage. Expression of *Can-elt-1* and *Can-elt-5* is similar to their orthologous genes in *C. elegans* (Page et al., 1997; Koh and Rothman, 2001). Anterior is to the left and dorsal is up in (A), (C), (D), (F).

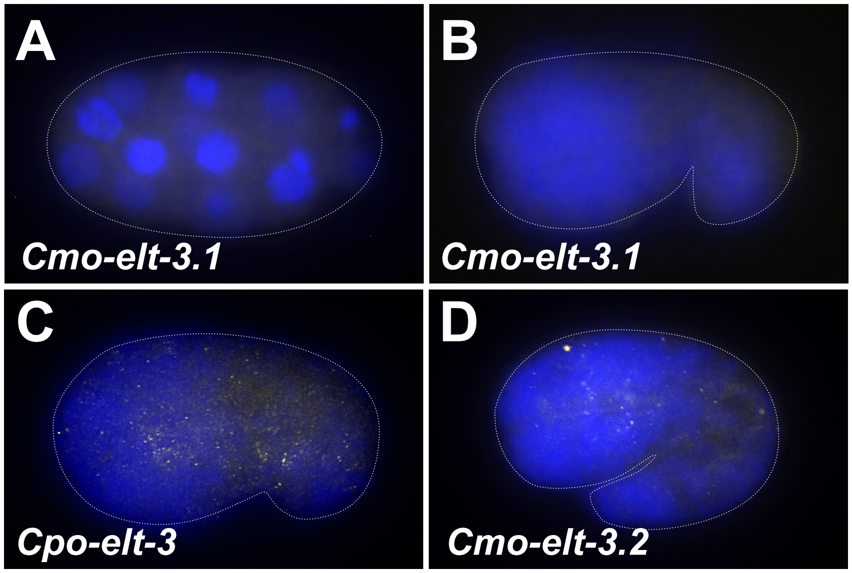

Fig. S5. Absence of embryonic expression of *Cmo-elt-3.1*, and hypodermal expression of *Cpo-elt-3* and *Cmo-elt-3.2*. (A) No expression of *Cmo-elt-3.1* was seen in early embryos (2E stage shown here). (B) No hypodermal expression was seen in later stages (comma stage shown here). (C) Expression of *Cpo-elt-3* in hypodermis of a bean stage embryo. (D) Expression of *Cmo-elt-3.2* in 1.7x stage embryo consistent with hypodermal cells.

Table S1. *C. angaria* genome sequence statistics

	C. angaria (this study)	***C. angaria*** (Wormbase; PRJNA51225.WS285)
Assembly size (Mb)	71.4	106.0
Number of scaffolds	6	34621
Average (kb)	11,895	3
Largest scaffold (kb)	13,323	868
Gaps (kbp)	6	11,442
GC (%)	33.7	35.7

Table S2. Tests of synergy of elt-3(gk121) with mutants in the C. elegans gut gene network Percentage of embryos containing gut and average number of gut nuclei made. An integrated *elt-2::GFP* reporter transgene, *wIs84*, was used to mark both the presence of gut as well as to count the number of gut nuclei at the end of embryogenesis (Owraghi et al., 2010). In pairwise combinations, none of the comparisons of mutant; *elt-3(+)* with mutant; *elt-3(gk121)* in gut nucleus number is significant (p>0.05 for all, *t*-test). Single *elt-7* null mutants make gut 100% of the time (McGhee et al., 2007). We did not test *elt-7(tm840); wIs84* alone. However, a comparison of *elt-7; elt-3* with *elt-3* alone was not significant (p>0.8).

Strain	Genotype	% making gut (n)	# of gut nuclei ± SD (n)
JR1130	wild type	100% (40)	19.7 ± 0.8 (40)
MS2046	*elt-3(gk121)*	100% (40)	20.0 ± 0.5 (40)
JR2719	*end-1(ok558)*	100% (40)	19.9 ± 0.6 (40)
MS2048	*end-1(ok558); elt-3(gk121)*	100% (130)	19.8 ± 0.6 (30)
MS2041	*end-3(ok1448)*	95.5% (201)	17.8 ± 5.3 (192)
MS2042	*end-3(ok1448); elt-3(gk121)*	95.0% (161)	18.1 ± 5.6 (153)
MS2049	*elt-7(tm840); elt-3(gk121)*	100% (104)	20.0 ± 0.6 (30)

Table S3. Nematode strains used in this work

C. elegans strains	
Strain Designation	Description
JR1130	*wIs84 [elt-2::NLS::GFP::lacZ, rol-6D] X*
JR2719	*end-1(ok558) V; wIs84 [elt-2::NLS::GFP::lacZ, rol-6D] X*
MS2041	*end-3(ok1448) V; wIs84 [elt-2::NLS::GFP::lacZ, rol-6D] X*
MS2042	*end-3(ok1448) V; elt-3(gk121) wIs84 [elt-2::NLS::GFP::lacZ, rol-6D] X*
MS2046	*elt-3(gk121) wIs84 [elt-2::NLS::GFP::lacZ, rol-6D] X*
MS2048	*end-1(ok558) V; elt-3(gk121) wIs84 [elt-2::NLS::GFP::lacZ, rol-6D] X*
MS2049	*elt-7(tm840) V; elt-3(gk121) wIs84 [elt-2::NLS::GFP::lacZ, rol-6D] X*
MS2582	*elt-7(tm840) V; elt-2(ca15) X; irEx798 [pGB598(Can-ELT-2::GFP), pMM809(unc-119::CFP), pDP#MM016B(unc-119(+))]*
MS2583	*unc-119(ed4); end-3(ok1448) V; irEx798 [pGB598(Can-elt-2_5kbp_promoter::ELT-2::GFP) + pMM809(unc-119::CFP) + pDP#MM016B(unc-119(+))]*
MS2584	*unc-119(ed4) III; end-1(ok558) end-3(ok1448) V; irEx498 [pMM768(end-3(+)), pMM824(unc-119::mCherry)]; irEx798 [pGB598(Can-elt-2_5kbp_promoter::ELT-2::GFP) + pMM809(unc-119::CFP) + pDP#MM016B(unc-119(+))]*
MS2606	*unc-119(ed4) III; him-8(e1489) IV; elt-7(tm840) end-1(ok558) end-3(ok1448) V; irEx804 [pGB612(end-3::END-3upstream::CanELT-3DBD::CFP) + pDP#MM016B(unc-119(+)) + pMM531(unc-119::YFP)]*
MS2612	*elt-2(ca15) X; elt-7(tm840) III; irEx803 [pGB608(Can-elt-2_3kbp_promoter::ELT-2::GFP), pDP#MM016B(unc-119(+))]; irEx808 [pGB619(hsp16-41::CanELT-3B::CFP) + rol-6D]*
MS2617	*elt-2(ca15) X; elt-7(tm840) III; irEx803 [pGB608(Can-elt-2_3kbp_promoter::ELT-2::GFP), pDP#MM016B(unc-119(+))]; irEx809 [pGB620(hsp16-41::CanELT-3A::CFP) + rol-6D]*
MS2625	*elt-7(tm840) end-1(ok558) end-3(ok1448) V; irEx814 [pGB618(Cel-end-3::Can-ELT-3B::CFP) + pMM824(unc-119::mCherry)]*
MS2629	*elt-7(tm840) end-1(ok558) end-3(ok1448) V; elt-2(ca15) X; irEx813 [pGB608(Can-elt-2_3kbp_promoter::Can-ELT-2::GFP) + pGB618(Cel-end-3::Can-ELT-3B::CFP) + pMM824(unc-119::mCherry)]*
MS2636	*elt-7(tm840) end-1(ok558) end-3(ok1448) V; irEx798 [pGB598(Can-elt-2_5kbp_promoter::ELT-2::GFP) + pMM809(unc-119::CFP) + pMM016B]; irEx814 [pGB618(Cel-end-3::Can-ELT-3B::CFP) + pMM824(unc-119::mCherry)]*
N2	*C. elegans* reference strain
Other species	
Strain Designation	Description
EG4788	*C. portoensis* reference strain
JU1667	*C. monodelphis* reference strain
MS2628	*C. angaria elt-3(ir79)/+ X*
PS1010	*C. angaria* reference strain
RGD1	*C. angaria* wild isolate

References

Koh, K. and Rothman, J. H. (2001). ELT-5 and ELT-6 are required continuously to regulate epidermal seam cell differentiation and cell fusion in C. elegans. *Development* **128**, 2867-2880.

McGhee, J.D., Sleumer, M.C., Bilenky, M., Wong, K., McKay, S.J., Goszczynski, B., Tian, H., Krich, N.D., Khattra, J., Holt, R.A., et al. (2007). The ELT-2 GATA-factor and the global regulation of transcription in the C. elegans intestine. *Dev. Biol.* **302**, 627-645.

Owraghi, M., Broitman-Maduro, G., Luu, T., Roberson, H., Maduro, M.F. (2010). Roles of the Wnt effector POP-1/TCF in the C. elegans endomesoderm specification gene network. *Dev. Biol.* **340**, 209-221.

Page, B. D., Zhang, W., Steward, K., Blumenthal, T. and Priess, J. R. (1997). ELT-1, a GATA-like transcription factor, is required for epidermal cell fates in Caenorhabditis elegans embryos. *Genes Dev.* **11**, 1651-1661.

Supplementary Materials and Methods:

1) gene models - sequences and coding DNA sequences (CDSs)

2) RNAi clone information

3) plasmid maps and cloning information

4) smiFISH probesets

5) CRISPR/Cas9 PCR primers and sequence of *ir79* allele

1) GENE MODELS:

Can-eft-3 (eef1A.1)

join(2248..2391,2447..>3436,3494..3760)

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Can-skn-1

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Partial Can-SKN-1 coding region:

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Only a single SKN-1 orthologue is apparent in C. angaria. A search of the partial Can-SKN-1 protein sequence to C. elegans returns Cel-skn-1 with an expected value of 1E-109.

Cpo-elt-2

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4..11631)

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C. monodelphis elt-2, *elt-3.1* and *elt-3.2* are described in Eurmsirilerd and Maduro, 2020.

2) <u>RNAi clone information</u>

Sequences used for RNAi constructs (feeding and injection, where applicable): DNA was amplified from genomic PS1010 DNA, or synthesized directly, and cloned as HindIII / SacI fragments into the same sites of pPD129.36.

Can-skn-1 (smaller 520bp region from genomic DNA, contains a small intron):

gatacgtcatcgacttgttcaagattgtcatcggaatcaccaagatataatagagaggaaggaacgagtcaaggaattccagctagc
aggtaagaaatagagttatgacgtggcaaattgacactttgctggaattacgaaattagccaacaaatcaattgatttttcagattctatg
gaaaattgattccaaatgatttcactttcggtcgtcaaacttcgccaattcggaaaatctcacgtgtcattccattgactggacaacaaca
aagaaaacgtggaagacaatcaaaagatgaacaacttgcaactgaaatggcacttccagtcactgctcatcaaataagcgaaatg
agcctttccgaattgcaacaagtactcaaaatcgacacattgtccgaaggtcaaaaacaattgatccgcaaaattcgtcgccgcgga
aagaacaaggttgccgcgcggacttgccgtcaaagaagaactgatcgacatggaagagccattgatatgaattatattt

Can-skn-1 (larger fragment, 750bp, synthesized cDNA):

aaaatgttctcgcaatcaaataatcaggaaggatattcggattatcaacaaaatccaaatcaatattcaaatgatcccaaaatgacgc
caggtttctatcgtccaatgggaattcgttggagagatgatcaaagtttgatgcaagttcaaactatgccaattttaactgaacacagatc
agcctcgccatttggaacaagaagatcaaattcaccatcaagaccaaccacttcgaatcgttttgatacatattcatatgattcgccatct
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ggtatttgatcaacaaaatattgtgcagacacaacaatcacagcagccaattttgcaaaatgtttcattgagcgagggaattgtttattcg
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ccaatgatcaacgacactgcgcacgaacaagtttcgaatggaaatgttgattttgatcatagatattcgagaattccaccaaccgcaac
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caacactttcaagtccactttttgatccatatcattcgatgagacaatcgttttctgatgttgatacgtcatcgacttgttcaagattgtcatcgg
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ctcaaaatcgacacattgtccgaaggtcaaaaacaattgatccgcaaaattcgtcgccgcggaaagaacaaggttgccgcgcgga
cttgccgtcaaagaagaactgatcgacatggaagagccattgatatgaattatatttaaagtttttattggttctccttactttttaccctattat
atacctcgaaagggtct

Can-pop-1 (773bp from genomic, contains a small intron):

cctcaatatggattacgagccggactttcaccgaattttgttctcccaatgcaagctggtctttcaccttcatttaatatgtttccaacatctcc
attttatggtgctgcaatggctgcagttgctaaacaacatatggaaaatgcagttccattagcagcaagtatgagaggaacaccattga
atccgttgaatcaaatgagaatgccaccgtatctaaatccaggaaatggacatttacatcaaaaagaaagaggacatggcggaggt
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aagaagaataagaaaaagaggagggataagagtattagtaagttgtttttgttagaaaatgaaaatatatttgtatttttgtagtttcagttt
cggagaatggtgatcagaagaaatgtcgagccagatttggtgtgaataatcaagaaatgtggtgtaaattctgtaagagaaagaaga
agtgcgaatattcgagtgatagaaatacagatacaatgatgatggatttacaagatagagg

Partial Can-POP-1 protein sequence:

Pqyglraglspnfvlpmqaglspsfnmfptspfygaamaavakqhmenavplaasmrgtplnplnqmrmppylnpgnghlhqk
erghggggkikkddhikkplnafmwfmkenrkklldeignnekqsaelnkelgkrwhdlpkeeqqkyfelakkdredhkqkypqw
sarenyavnkknkkkrrdksiisengdqkkcrarfgvnnqemwckfckrkkkceyssdrntdtmmmdlqdr

Only a single POP-1 orthologue is apparent in C. angaria. A search of the partial Can-POP-1 protein sequence to C. elegans returns Cel-pop-1 with an expected value of 3E-64.

Can-elt-2 (no DBD) (522 bp genomic fragment, contains small intron):

aactttcgcaactggaacaaatcaagattatttgtataacacttcttcaatttttaacacatattcaactcaaccgcctccatcacaatatca
tactttatcatcatcgtaagttggccgggaaattttgaattcttgaatgtttctgatttcacaataagaaaatattttcaagacatcaaactcca
attttttaacagattttacgataattcggcatttataccacaacaacctttcgcgccagcaaatccagaatgtgtcaaatgcgcaaatcca
gtcatttccggaagacaagttgatggtggatatatgtgtgatacttgtagtaattctaatgtatatgatttgtcacgaattcaaacaaactatt
cgatgccactttctcttgttcaaccaattgaacctgctccaattgaacaaattccacctccagcaccagcaccatcaaaaccaccaaaa
tcatcaagtaacaaaaaagctggatcagctggaagcacaaatcgccgacaaggattggtt

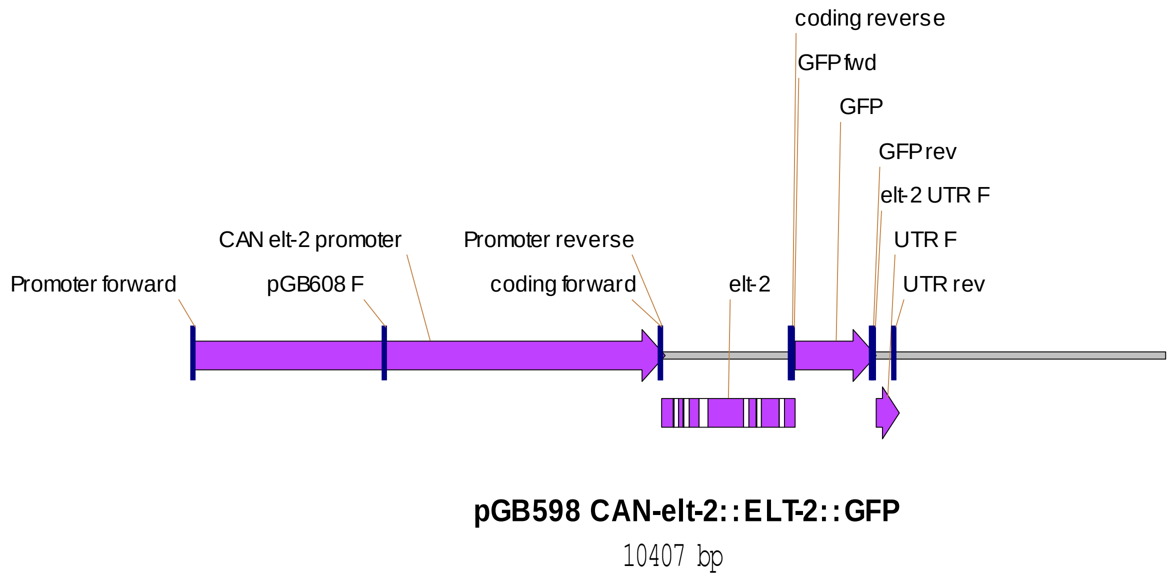
Can-elt-3 (no DBD) (270 bp fragment, common to short and long isoforms):

ccgatgctcaatcaacaagatatgctaaacccgatgaatttctccaccaactacatgggagccacacaattctcccaccaacagact
gtgcaactcaatttgctagatctcagcggcttctccacattcaactcgtgtgatccagctctaccattaatgcaccagtcaccttctcaaca
aaattatgtgatgccacattcaactttacaccacctccacaagatccattggtcgctgaacaaaaaccacttgtcaaaaagcgaatgg
ct

3) <u>Plasmid maps and cloning information</u>

All fragments were amplified by PCR using Q5 polymerase (New England Biolabs). Plasmids were cloned by Gibson Assembly into pBluescript KS- digested with BamHI/HinDIII. Fragment sizes include primer overhangs for Gibson Assembly. Linearized plasmid maps and primer sequences are shown below.

pGB598: *Can-elt-2::ELT-2::GFP*



<u>*Can-elt-2* promoter (5040 bp amplified from PS1010 genomic DNA)</u>

Promoter forward, ctatagggcgaattggagcttcatttattatatgtcattacggaaaaccg
Promoter reverse, gacgcatactatcgatccagggactctcgatcg

<u>*Can-elt-2* coding (1436 bp amplified from PS1010 genomic DNA)</u>

Coding forward, ctggatcgatagtatgcgtccaccaacattgcaact
Coding reverse, ctttactcatttgttcctcttccagatctctcgcag

<u>GFP (870bp amplified from pPD95.67, introns not shown above for simplicity)</u>

GFP_fwd agaggaacaaatgagtaaaggagaagaacttttcactggag
GFP_rev aatagttgaactatttgtatagttcatccatgccatg

<u>*Can-elt-2 3'*UTR (251bp amplified from PS1010 genomic DNA)</u>

UTR_fwd atacaaatagttcaactatttttatcaataatttgctc
UTR_rev cgaggtcgacggtatcgatagaatgctcatcttcgcaaacttcgg

pGB598 Sequence:

aactagtgcattgtcagagagcccgtggttctgataccaatttttacagataaagttgttcccacacgtttttgagtttttttcgaaaaatatt
gattgacttacttactttgaactcaatttcaaactatctttcgaatattccaccttatttgtgagatgtctcttatttccctggaaaaatttgagtag
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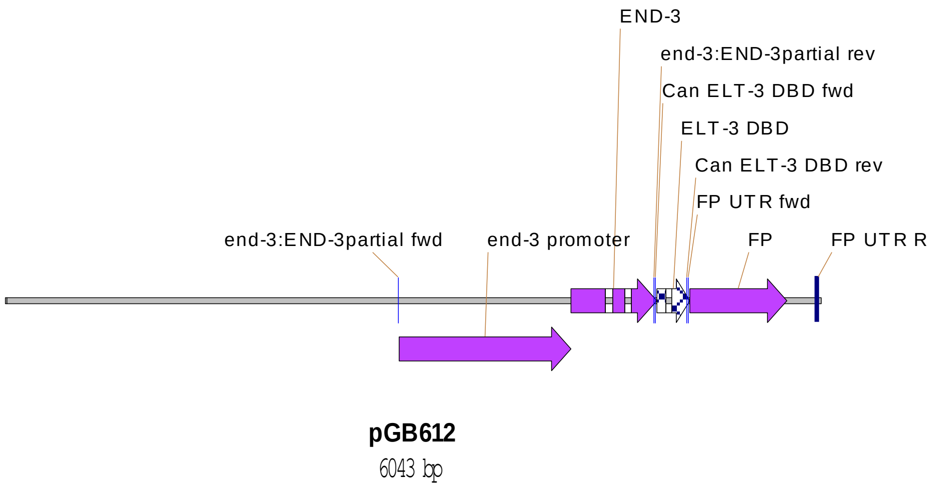
pGB608 *Can-elt-2::ELT-2::GFP*- smaller promoter- (diagrammed in pGB598)

all primers used to generate pGB608 are the same as for pGB598 except the Forward promoter primer :

<u>*Can-elt-2* promoter (2990 bp amplified from PS1010 genomic DNA):</u>

608F, cggccgctctagaactagtggatcaaagtgaatgatgataaatctttcaaag
Promoter reverse, gacgcatactatcgatccagggactctcgatcg

pGB612 *Cel-end-3::END-3upstream::Can-ELT-3_DBD::CFP*



Cel-end-3 promoter and partial coding (1976bp from genomic DNA):

end-3:END-3partial_fwd cggccgctctagaactagtgcatccaatttagtgtatatttatttcc
end-3:END-3partial_rev agtttgagcacgaagagttgatatgcattg

Can-ELT-3 DBD (265 bp from PS1010)

Can ELT-3 DBD_fwd caactcttcgtgctcaaactgcaagacac
Can ELT-3 DBD_rev ctttactcatgcatgtggcttgtcttctg

CFP+UTR (988bp cassette)

CFP was originally amplified from pEB1-SCFP3A and the *Cel-end-3* UTR was amplified from genomic DNA. The CFP and UTR were stitched together by PCR using the following primers:

CFP UTR_fwd agccacatgcatgagtaaaggagaagaac
CFP UTR_rev cgaggtcgacggtatcgataagctctttccactatagtc

pGB608 sequence

tatcgataccgtcgacctcgagggggggcccggtacccagcttttgttccctttagtgagggttaatttcgagcttggcgtaatcatggtca
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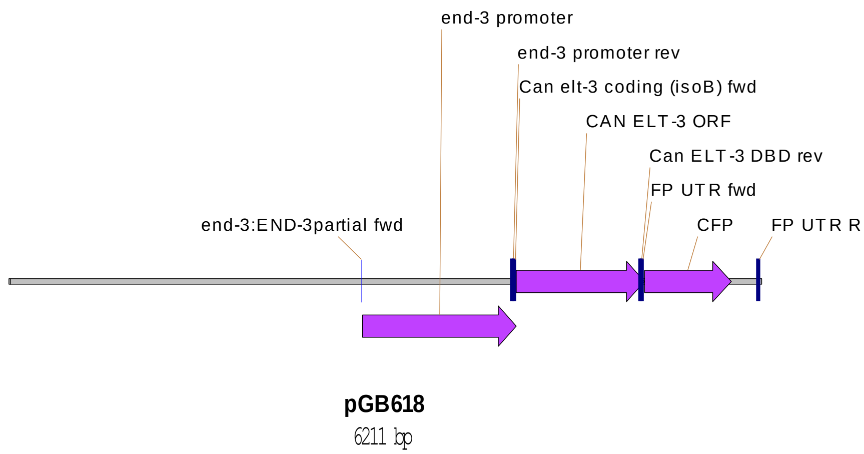
pGB618: *Cel-end-3::Can-ELT-3B::CFP*



<u>*Cel-elt-3* promoter (1290 bp amplified from N2 genomic DNA)</u>

end-3:END-3partial_fwd cggccgctctagaactagtgcatccaatttagtgtatatttatttcc
end-3 promoter_rev gagtattcatgtttatactttgaatgagaatgc

<u>*Can-elt-3* coding (1076 bp amplified from IDT synthesized minigene)</u>

Can elt-3 coding (isoB)_fwd aagtataaacatgaatactcaaagtcctctcg
Can ELT-3 DBD_rev ctttactcatgcatgtggcttgtcttctg

<u>CFP UTR (988bp cassette –as in pGB612 (see above)</u>

pGB618 sequence

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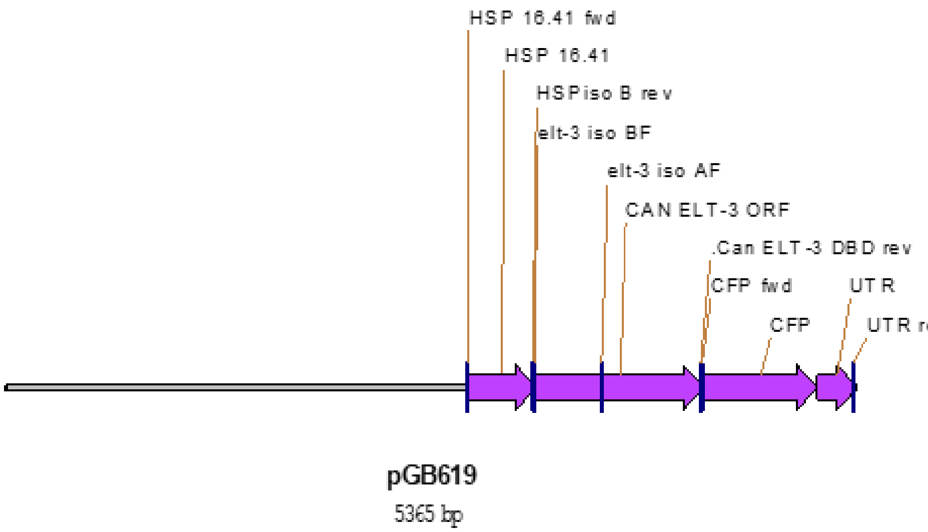
Development • Supplementary information

pGB619 (*hsp16.41promoter::ELT-3B::CFP::end-3_3'UTR*)



<u>Heat-shock promoter (444bp amplified from pPD49.83)</u>

HSP 16.41_fwd cggccgctctagaactagtg<u>gatcaccaaaaacggaac</u>
HSPiso B rev gagtattcatggatcccgatgaggattttc

<u>*Can-elt-3B* coding (1076 bp amplified from IDT synthesized minigene)</u>

elt-3 iso BF, atcgggatccatgaatactcaaagtcctctcgggattcaa
Can ELT-3 DBD_rev ctttactcatgcatgtggcttgtcttctg

<u>CFP UTR (988bp cassette –as in pGB612 (see above)</u>

pGB619 sequence

tatcgataccgtcgacctcgagggggggcccggtacccagcttttgttccctttagtgagggttaatttcgagcttggcgtaatcatggtca
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aattgatttcctgtgtggagtgtgaacaataagcatgactatagtggaaagagct

pGB620 *hsp16-41promoter::ELT-3A::CFP::end-3_3'UTR* - see notation in pGB619

Heat-shock promoter (444bp amplified from pPD49.83)

HSP 16.41_fwd cggccgctctagaactagtggatcaccaaaaacggaac
HSP16.41 isoArev cttccttcatggatcccgatgaggattttc

Can elt-3 coding isoA (665bp amplified from IDT synthesized minigene (noted in pGB619 above)

elt-3 isoAF atcgggatccatgaaggaagttcagatcaaagaggag
Can ELT-3 DBD_rev ctttactcatgcatgtggcttgtcttctg

CFP UTR 988bp cassette –as in pGB612 (see above)

4) smiFISH PROBESETS:

Conjugated FLAP-X oligos (CACTGAGTCCAGCTCGAAACTTAGGAGG) that were 5′ and 3′ end-labelled with Quasar 570 or CalFluor 610 were synthesized by Biosearch Technologies. FLAP-X oligos that were 5′ and 3′ end-labelled with Cy5 or Cy3 were synthesized by IDT.

smiFISH antisense probesets (with complementary FLAP-X sequence added to 5' end of oligo (CAPS)):

Can-skn-1: 19 probes

CCTCCTAAGTTTCGAGCTGGACTCAGTGttcctttcgacaaatcttcata
CCTCCTAAGTTTCGAGCTGGACTCAGTGcaaatcttcatcgcttggaatt
CCTCCTAAGTTTCGAGCTGGACTCAGTGttcttcttttgaaacatcttcc
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgtctgcacaatattttgttga
CCTCCTAAGTTTCGAGCTGGACTCAGTGtcgctcaatgaaacattttgca
CCTCCTAAGTTTCGAGCTGGACTCAGTGatgctcatttcggttaaattgt
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CCTCCTAAGTTTCGAGCTGGACTCAGTGttgttgcattgtattcgaatca
CCTCCTAAGTTTCGAGCTGGACTCAGTGgtgtcgttgatcattggaatat
CCTCCTAAGTTTCGAGCTGGACTCAGTGttggtggaattctcgaatatct

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CCTCCTAAGTTTCGAGCTGGACTCAGTGttgttcatcttttgattgtctt
CCTCCTAAGTTTCGAGCTGGACTCAGTGctcatttcgcttatttgatgag
CCTCCTAAGTTTCGAGCTGGACTCAGTGatgtgtcgattttgagtacttg

Cmo-elt-3.2: 24 probes

CCTCCTAAGTTTCGAGCTGGACTCAGTGaaaagcggcgggaactcgaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGcgtgtcgcatcactgttagc
CCTCCTAAGTTTCGAGCTGGACTCAGTGttgtccaagtcctcgaactc
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CCTCCTAAGTTTCGAGCTGGACTCAGTGctgaatctccggcaacggag
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CCTCCTAAGTTTCGAGCTGGACTCAGTGaacacacgttcttggcgttg
CCTCCTAAGTTTCGAGCTGGACTCAGTG tcaggtgtgccccaagagct
CCTCCTAAGTTTCGAGCTGGACTCAGTG gtggcgtactgctgctcgcc
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgcttcatcatccgatgcat

Can-elt-3 (probeset1, detects both isoforms): 24 probes

CCTCCTAAGTTTCGAGCTGGACTCAGTGggattgggagctgtaaataac
CCTCCTAAGTTTCGAGCTGGACTCAGTGaacgcgatagtcgttcattga
CCTCCTAAGTTTCGAGCTGGACTCAGTGccaacggattggcaatctttt
CCTCCTAAGTTTCGAGCTGGACTCAGTGtcggtataaattggttgctca
CCTCCTAAGTTTCGAGCTGGACTCAGTGgattgagcatcggagcatttg
CCTCCTAAGTTTCGAGCTGGACTCAGTGcatcgggtttagcatatcttg
CCTCCTAAGTTTCGAGCTGGACTCAGTGccatgtagttggtggagaaat
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgttggtgggagaattgtgtg

CCTCCTAAGTTTCGAGCTGGACTCAGTGtctagcaaattgagttgcaca
CCTCCTAAGTTTCGAGCTGGACTCAGTGacgagttgaatgtggagaagc
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CCTCCTAAGTTTCGAGCTGGACTCAGTGttgttgagaaggtgactggtg
CCTCCTAAGTTTCGAGCTGGACTCAGTGgttgaatgtggcatcacataa
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CCTCCTAAGTTTCGAGCTGGACTCAGTGacgatttcgtttcattattcc
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Can-elt-3B-specific (probeset2): 16 probes

CCTCCTAAGTTTCGAGCTGGACTCAGTGccgagaggactttgagtatt
CCTCCTAAGTTTCGAGCTGGACTCAGTGagtttgggtagtaggtttga
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgtatcagtgcttgaatgcg
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CCTCCTAAGTTTCGAGCTGGACTCAGTGctggaaggggagcagcagag
CCTCCTAAGTTTCGAGCTGGACTCAGTGcttccttcattaaatgagcg

Can-elt-2: 24 probes

CCTCCTAAGTTTCGAGCTGGACTCAGTGggattgggagctgtaaataac
CCTCCTAAGTTTCGAGCTGGACTCAGTGaacgcgatagtcgttcattga
CCTCCTAAGTTTCGAGCTGGACTCAGTGccaacggattggcaatctttt
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CCTCCTAAGTTTCGAGCTGGACTCAGTGacgatttcgtttcattattcc
CCTCCTAAGTTTCGAGCTGGACTCAGTGcatttggtgattcgtttcttg

Can-elt-5: 24 probes

CCTCCTAAGTTTCGAGCTGGACTCAGTGtgatgatgactcgatttgtgtc
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgttaattcgggagtttcagcg
CCTCCTAAGTTTCGAGCTGGACTCAGTGttgatttcgtggtgcaattgta
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CCTCCTAAGTTTCGAGCTGGACTCAGTGtccttctctttatcactgataa
CCTCCTAAGTTTCGAGCTGGACTCAGTGcacgattaatcgattctttggt
CCTCCTAAGTTTCGAGCTGGACTCAGTGagaaattttccgaattctgcgc
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgtcgtttctcatctgataatc
CCTCCTAAGTTTCGAGCTGGACTCAGTGgaaatggtgaactagccgagac
CCTCCTAAGTTTCGAGCTGGACTCAGTGgatcgaacataaatccgtttgc
CCTCCTAAGTTTCGAGCTGGACTCAGTGtattcatcatttgttgttggtg
CCTCCTAAGTTTCGAGCTGGACTCAGTGggtttcttggagtttttgtttg
CCTCCTAAGTTTCGAGCTGGACTCAGTGcgatggtgatgttttgatgttg
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgaattgttggtggatggtgtg
CCTCCTAAGTTTCGAGCTGGACTCAGTGatcttcatcattattcgaagcg
CCTCCTAAGTTTCGAGCTGGACTCAGTGtttggtcgtcatacaattcgag
CCTCCTAAGTTTCGAGCTGGACTCAGTGgatgagtccgatgtaatcgata
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgttgaatgaaatctttgcgca
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgcgttttgatttgatacatca
CCTCCTAAGTTTCGAGCTGGACTCAGTGcatggctagaagttgactgaac
CCTCCTAAGTTTCGAGCTGGACTCAGTGatgagaaggcgtttggtgtatt
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgagttgattgaattgctccaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGtattcaattgctcctgaacttg

Can-elt-1: 24 probes

CCTCCTAAGTTTCGAGCTGGACTCAGTGcgcaatttacacactctcgg

CCTCCTAAGTTTCGAGCTGGACTCAGTGccaaagtggcgtgttttgaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGgcaaagataatgtccgccgc
CCTCCTAAGTTTCGAGCTGGACTCAGTGgtgatataatccacaggcat
CCTCCTAAGTTTCGAGCTGGACTCAGTGccgattttgtccgttcattt
CCTCCTAAGTTTCGAGCTGGACTCAGTGctttttcggcttcaccaacg
CCTCCTAAGTTTCGAGCTGGACTCAGTGtcgtttttgagtgctctgtc
CCTCCTAAGTTTCGAGCTGGACTCAGTGatttacacatgatacgccgg
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgttgttgtgttcgttttgc
CCTCCTAAGTTTCGAGCTGGACTCAGTGggaaccatttcgtcgccaaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgcattacaaactggctctc
CCTCCTAAGTTTCGAGCTGGACTCAGTGtcaacctgatgcagcttata
CCTCCTAAGTTTCGAGCTGGACTCAGTGtttttcatcgtcaatggtcg
CCTCCTAAGTTTCGAGCTGGACTCAGTGcgatttctcgtctgaattcc
CCTCCTAAGTTTCGAGCTGGACTCAGTGcctccttttgttgtcaattt
CCTCCTAAGTTTCGAGCTGGACTCAGTGcaaatccatttttcccatcg
CCTCCTAAGTTTCGAGCTGGACTCAGTGagcagctgcatttacagatg
CCTCCTAAGTTTCGAGCTGGACTCAGTGcttcattccccaaacagtgg
CCTCCTAAGTTTCGAGCTGGACTCAGTGcattggttgtgttgttgaag
CCTCCTAAGTTTCGAGCTGGACTCAGTGtcctcctgatgatgtcatta
CCTCCTAAGTTTCGAGCTGGACTCAGTGattatttgtggcgaatggat
CCTCCTAAGTTTCGAGCTGGACTCAGTGtcatcgatttgtggggtgaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGtccatgattggaattgatga
CCTCCTAAGTTTCGAGCTGGACTCAGTGcaggtcgtgcaataattgct

Cel-skn-1: 19 probes

CCTCCTAAGTTTCGAGCTGGACTCAGTGaatgtaggcgtagttggatgtt
CCTCCTAAGTTTCGAGCTGGACTCAGTGttgaagactgtcgttttgcatt
CCTCCTAAGTTTCGAGCTGGACTCAGTGaatctgtccatttgatacaact
CCTCCTAAGTTTCGAGCTGGACTCAGTGgtggattgataggaatgatcat
CCTCCTAAGTTTCGAGCTGGACTCAGTGagaggagacagtggagtctgac
CCTCCTAAGTTTCGAGCTGGACTCAGTGttgacgtcctgaagatccaatg
CCTCCTAAGTTTCGAGCTGGACTCAGTGaacgattgagtagctgttgcag
CCTCCTAAGTTTCGAGCTGGACTCAGTGctctgtgagtgatatggatcga
CCTCCTAAGTTTCGAGCTGGACTCAGTGgtagtgcaatcactaaacgagt
CCTCCTAAGTTTCGAGCTGGACTCAGTGcgagagcacgttgatgacgaat
CCTCCTAAGTTTCGAGCTGGACTCAGTGtatcgtggagattccgaagaga
CCTCCTAAGTTTCGAGCTGGACTCAGTGaactttccgtagaaacgagact
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgatgaacgtggagacgatgat
CCTCCTAAGTTTCGAGCTGGACTCAGTGaacaactctcgcgatcttaatc
CCTCCTAAGTTTCGAGCTGGACTCAGTGaatgacatctccgaaatctgga
CCTCCTAAGTTTCGAGCTGGACTCAGTGttcttcaacacttgttgcaact
CCTCCTAAGTTTCGAGCTGGACTCAGTGtctttgatactcgctgagactc
CCTCCTAAGTTTCGAGCTGGACTCAGTGtcgacgaatcttgcgaatcaac
CCTCCTAAGTTTCGAGCTGGACTCAGTGatgatgtaatgggacatcttgt

Cel-elt-3: 18 probes

CCTCCTAAGTTTCGAGCTGGACTCAGTGtttttccactcgataatcgttc
CCTCCTAAGTTTCGAGCTGGACTCAGTGtatggatctagtagtgtgttgg
CCTCCTAAGTTTCGAGCTGGACTCAGTGtaggtcggttggtcaagttgaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGcagtacctgtgcattagtgaag
CCTCCTAAGTTTCGAGCTGGACTCAGTGaagttcatctggagcatttctt
CCTCCTAAGTTTCGAGCTGGACTCAGTGtaacttgtggataatggtgttg
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgtgtgacttgtgctgtattca
CCTCCTAAGTTTCGAGCTGGACTCAGTGatattgaacggcatttgatgtg
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgttgcgaagtttgaaaggttc
CCTCCTAAGTTTCGAGCTGGACTCAGTGgtgagctgttgagaagtggaag
CCTCCTAAGTTTCGAGCTGGACTCAGTGtttgacattgttgtgtatggat
CCTCCTAAGTTTCGAGCTGGACTCAGTGcattcttttttcatcggcttc
CCTCCTAAGTTTCGAGCTGGACTCAGTGattttgatgacattgtacagct
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgttttgcaatttgagcagatg
CCTCCTAAGTTTCGAGCTGGACTCAGTGgtttttccggaagtagagattg
CCTCCTAAGTTTCGAGCTGGACTCAGTGtaagagacagtggacgttttcg
CCTCCTAAGTTTCGAGCTGGACTCAGTGttctcttcataattccatcttt
CCTCCTAAGTTTCGAGCTGGACTCAGTGgagtttggagactcatttctag

Cel-end-3: 20 probes

CCTCCTAAGTTTCGAGCTGGACTCAGTGgaagagaatgagttcgagtaca
CCTCCTAAGTTTCGAGCTGGACTCAGTGaagacattggtgagttggaaga
CCTCCTAAGTTTCGAGCTGGACTCAGTGgggaaactgagggaatccaaaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGttcttcattgaattggacttgt
CCTCCTAAGTTTCGAGCTGGACTCAGTGatattctggaggacatctggag
CCTCCTAAGTTTCGAGCTGGACTCAGTGttgtgccggataatgatgataa
CCTCCTAAGTTTCGAGCTGGACTCAGTGcattgtatgaatttgtcgtcat
CCTCCTAAGTTTCGAGCTGGACTCAGTGctgttgcattgaattgtcatag
CCTCCTAAGTTTCGAGCTGGACTCAGTGaccattatctggttgcataaaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGgatagttttcattgtagtatcc
CCTCCTAAGTTTCGAGCTGGACTCAGTGattaataggcgtcgtgaattca
CCTCCTAAGTTTCGAGCTGGACTCAGTGttgatggaagtttgagattgct
CCTCCTAAGTTTCGAGCTGGACTCAGTGcatggaattttcattgttgtcg
CCTCCTAAGTTTCGAGCTGGACTCAGTGcttgaacttcttgagctggaaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGtatttttcgaagcttttctagc
CCTCCTAAGTTTCGAGCTGGACTCAGTGatgcattgtggaaatcttcttg
CCTCCTAAGTTTCGAGCTGGACTCAGTGacaattggagcacgaagagttg
CCTCCTAAGTTTCGAGCTGGACTCAGTGtcgttcgtaaagattacacgga
CCTCCTAAGTTTCGAGCTGGACTCAGTGaagatgttgtggtcttttatgt
CCTCCTAAGTTTCGAGCTGGACTCAGTGtagagtcaacaagtggagcaac

Cel-end-1: 20 probes

CCTCCTAAGTTTCGAGCTGGACTCAGTGtggggagatggagatgaagaag

CCTCCTAAGTTTCGAGCTGGACTCAGTGatgactccataggtatatggaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGtttcatgggaatgaacatttcc
CCTCCTAAGTTTCGAGCTGGACTCAGTGtttccatgattatccggaaaat
CCTCCTAAGTTTCGAGCTGGACTCAGTGtacatttgtgtagcatcgagtg
CCTCCTAAGTTTCGAGCTGGACTCAGTGtcaaaatattgatctccaccac
CCTCCTAAGTTTCGAGCTGGACTCAGTGgattgaacaaagtattgcccaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGtatcgagacttccaaacattgg
CCTCCTAAGTTTCGAGCTGGACTCAGTGgatactgttgtgagtagcaatt
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgttgatagtcttgtccaattt
CCTCCTAAGTTTCGAGCTGGACTCAGTGgactggtggaatattttcgatt
CCTCCTAAGTTTCGAGCTGGACTCAGTGgttcacaattttgcgatttgta
CCTCCTAAGTTTCGAGCTGGACTCAGTGagagttcgtgtggaacgttgag
CCTCCTAAGTTTCGAGCTGGACTCAGTGtcacgagttctacaatttggat
CCTCCTAAGTTTCGAGCTGGACTCAGTGgaatcagttcttctccatagag
CCTCCTAAGTTTCGAGCTGGACTCAGTGattttttcggaagtacagggag
CCTCCTAAGTTTCGAGCTGGACTCAGTGttttcattattgttttacggca
CCTCCTAAGTTTCGAGCTGGACTCAGTGtggtgttggagattgtacttcg
CCTCCTAAGTTTCGAGCTGGACTCAGTGattatgacatagtttcgagtct
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgtgatggaatgttctgtagtg

Cel-elt-2: 24 probes

CCTCCTAAGTTTCGAGCTGGACTCAGTGcagccgttgacattatcatt
CCTCCTAAGTTTCGAGCTGGACTCAGTGcattattttgctctggtgga
CCTCCTAAGTTTCGAGCTGGACTCAGTGtttgccgttcaattggagat
CCTCCTAAGTTTCGAGCTGGACTCAGTGtagttatttgtgccacttgt
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgagtagtctatgccagtat
CCTCCTAAGTTTCGAGCTGGACTCAGTGgtttggtccaacattccaaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGagtgtttacgggaattccac
CCTCCTAAGTTTCGAGCTGGACTCAGTGtggagtttgtatatcccgaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGgaatactggggtcgtacaga
CCTCCTAAGTTTCGAGCTGGACTCAGTGgccacagttggataagtaga
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgtgagcatttgacgcattc
CCTCCTAAGTTTCGAGCTGGACTCAGTGgtgttgaacagttgacgcac
CCTCCTAAGTTTCGAGCTGGACTCAGTGactggaggagaatacgtggt
CCTCCTAAGTTTCGAGCTGGACTCAGTGttttagaggattgcttggca
CCTCCTAAGTTTCGAGCTGGACTCAGTGcaattggagcacacaagtcc
CCTCCTAAGTTTCGAGCTGGACTCAGTGcatttcttctccagagagtt
CCTCCTAAGTTTCGAGCTGGACTCAGTGggagtttgaagtaaagcccg
CCTCCTAAGTTTCGAGCTGGACTCAGTGctgtaaagcaccttctttct
CCTCCTAAGTTTCGAGCTGGACTCAGTGcgttgatggtgtggaagagt
CCTCCTAAGTTTCGAGCTGGACTCAGTGtctctcaaacttcctttctc
CCTCCTAAGTTTCGAGCTGGACTCAGTGcggcatgtgacacatatgtg
CCTCCTAAGTTTCGAGCTGGACTCAGTGtactgtacgtttgatctggc
CCTCCTAAGTTTCGAGCTGGACTCAGTGttgttgctgcatacacgtag
CCTCCTAAGTTTCGAGCTGGACTCAGTGctggcatcacatgaacttgg

Cpo-elt-3: 22 probes

CCTCCTAAGTTTCGAGCTGGACTCAGTGtgtcgagtagaagctgttca
CCTCCTAAGTTTCGAGCTGGACTCAGTGcggattctgtagattgtggg
CCTCCTAAGTTTCGAGCTGGACTCAGTGattccagtcattgtagttgg
CCTCCTAAGTTTCGAGCTGGACTCAGTGcttgctgatccgaatttgtt
CCTCCTAAGTTTCGAGCTGGACTCAGTGgcaaattcggagctgtttga
CCTCCTAAGTTTCGAGCTGGACTCAGTGagttatcagtgttgttctgc
CCTCCTAAGTTTCGAGCTGGACTCAGTGggaaatcgctggcaatggag
CCTCCTAAGTTTCGAGCTGGACTCAGTGacttggagttccttcatgaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGtccagctgtataatagtcgt
CCTCCTAAGTTTCGAGCTGGACTCAGTGtttttcaacgcgatagtcgc
CCTCCTAAGTTTCGAGCTGGACTCAGTGggttgttccaattgagtgta
CCTCCTAAGTTTCGAGCTGGACTCAGTGtggtgtgttggtaaattccg
CCTCCTAAGTTTCGAGCTGGACTCAGTGcatgtggttcatcatgtctt
CCTCCTAAGTTTCGAGCTGGACTCAGTGgttactggcattactgtcat
CCTCCTAAGTTTCGAGCTGGACTCAGTGagggaatgctgagaggttga
CCTCCTAAGTTTCGAGCTGGACTCAGTGgctcgcaaactgatgtgttg
CCTCCTAAGTTTCGAGCTGGACTCAGTGaaggcgattgatgcatcagt
CCTCCTAAGTTTCGAGCTGGACTCAGTGttgaatgtgcagccattacg
CCTCCTAAGTTTCGAGCTGGACTCAGTGctaatggatcttgtggtggt
CCTCCTAAGTTTCGAGCTGGACTCAGTGttttgatggcattggacagc
CCTCCTAAGTTTCGAGCTGGACTCAGTGgcaatgagagtggtcgtttg
CCTCCTAAGTTTCGAGCTGGACTCAGTGatttctcttcatgatgccat

Cpo-elt-2: 22 probes

CCTCCTAAGTTTCGAGCTGGACTCAGTGaatttgtcctccattcacat
CCTCCTAAGTTTCGAGCTGGACTCAGTGatgtgatgtaggttgttcgt
CCTCCTAAGTTTCGAGCTGGACTCAGTGgtgtttcaatgattggcgga
CCTCCTAAGTTTCGAGCTGGACTCAGTGcttattaccagacttggtgg
CCTCCTAAGTTTCGAGCTGGACTCAGTGttgatgaattacccgatccg
CCTCCTAAGTTTCGAGCTGGACTCAGTGgaacacaccaatccttgacg
CCTCCTAAGTTTCGAGCTGGACTCAGTGctccacagagttgtatttgt
CCTCCTAAGTTTCGAGCTGGACTCAGTGgcagcttgtagtaaagacca
CCTCCTAAGTTTCGAGCTGGACTCAGTGgaccttctttcttcatcgaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGtttccttttctagtttgca
CCTCCTAAGTTTCGAGCTGGACTCAGTGgagttgagtgtgtttcactc
CCTCCTAAGTTTCGAGCTGGACTCAGTGttcttttgtgcgggaacttg
CCTCCTAAGTTTCGAGCTGGACTCAGTGgcagcagtagatccatcaaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGatgaagcatttctagcagcg
CCTCCTAAGTTTCGAGCTGGACTCAGTGgtatggtgtttcttgaagcc
CCTCCTAAGTTTCGAGCTGGACTCAGTGcacgatgcttgttggtaagt
CCTCCTAAGTTTCGAGCTGGACTCAGTGaatatgagcgtcttgcgacg
CCTCCTAAGTTTCGAGCTGGACTCAGTGctaggcccaaatgtggtaag
CCTCCTAAGTTTCGAGCTGGACTCAGTGagacagttgtttgggaagct
CCTCCTAAGTTTCGAGCTGGACTCAGTGccggtatgacttgaagatct

CCTCCTAAGTTTCGAGCTGGACTCAGTGgcttttgtttcatcgtcttg
CCTCCTAAGTTTCGAGCTGGACTCAGTGgtagtgtcttccaaatctct

Can-myo-2: 22 probes

CCTCCTAAGTTTCGAGCTGGACTCAGTGaaccttttgctcattcatgg
CCTCCTAAGTTTCGAGCTGGACTCAGTGaagagcgcggatttgatgtt
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgcaagtcttcgctcaattt
CCTCCTAAGTTTCGAGCTGGACTCAGTGcttcaagcgattgcatgagt
CCTCCTAAGTTTCGAGCTGGACTCAGTGttgagcaatcttgagctctc
CCTCCTAAGTTTCGAGCTGGACTCAGTGttgttcgtcttccaatttca
CCTCCTAAGTTTCGAGCTGGACTCAGTGctcatcagccaattgatctt
CCTCCTAAGTTTCGAGCTGGACTCAGTGatctatcagcacgttgacga
CCTCCTAAGTTTCGAGCTGGACTCAGTGagctcatcgaattcagcttg
CCTCCTAAGTTTCGAGCTGGACTCAGTGttgatcttcgagttgctcat
CCTCCTAAGTTTCGAGCTGGACTCAGTGcttcttccaaatcacgacgg
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgagttgctctccgaattta
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgcaattgttcgacttggtc
CCTCCTAAGTTTCGAGCTGGACTCAGTGtttttcgatgcggttctttt
CCTCCTAAGTTTCGAGCTGGACTCAGTGagaagtctgacttcatgtcc
CCTCCTAAGTTTCGAGCTGGACTCAGTGttcgtcagccttaagacgaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGtcaaacgtcccttggatgaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGtagcgtgatccaattcgttg
CCTCCTAAGTTTCGAGCTGGACTCAGTGcttggacaagttcgagaggt
CCTCCTAAGTTTCGAGCTGGACTCAGTGccttgagttgttcaagttca
CCTCCTAAGTTTCGAGCTGGACTCAGTGttggagttgacgagtagctt
CCTCCTAAGTTTCGAGCTGGACTCAGTGattcatcggctccaataagt

Can-eft-3 (eef1A.1): 16 probes

CCTCCTAAGTTTCGAGCTGGACTCAGTGgcgtacttgaacgaaccttt
CCTCCTAAGTTTCGAGCTGGACTCAGTGcaatggtgataccacgttca
CCTCCTAAGTTTCGAGCTGGACTCAGTGcctggagcatcaataatggt
CCTCCTAAGTTTCGAGCTGGACTCAGTGcttgacaccaagagtttggg
CCTCCTAAGTTTCGAGCTGGACTCAGTGttgttgcaggcaacgatcaa
CCTCCTAAGTTTCGAGCTGGACTCAGTGgaaacgggcctcagagaatg
CCTCCTAAGTTTCGAGCTGGACTCAGTGccgatcttcttgatgaatcc
CCTCCTAAGTTTCGAGCTGGACTCAGTGacgaatggaacagcctttgg
CCTCCTAAGTTTCGAGCTGGACTCAGTGaacttcgagcatgttgtctc
CCTCCTAAGTTTCGAGCTGGACTCAGTGgacgtcttggagtggaagac
CCTCCTAAGTTTCGAGCTGGACTCAGTGacttcagtggtgacgttttg
CCTCCTAAGTTTCGAGCTGGACTCAGTGtgacgttgaatccgacgttg
CCTCCTAAGTTTCGAGCTGGACTCAGTGatgtccttgacggaaacgtt
CCTCCTAAGTTTCGAGCTGGACTCAGTGtttcgagtcagagcagactg
CCTCCTAAGTTTCGAGCTGGACTCAGTGagaactggagtgtaaccagc
CCTCCTAAGTTTCGAGCTGGACTCAGTGccttgagctcgttgaatttg

5) CRISPR/Cas9 PCR primers and sequence of *ir79* allele

Can-elt-3 crRNA targeting site 1 :gtgcttgaatgcggtgagtttgg
Can-elt-3 crRNA targeting site 2: gaatttctccaccaactacatgg

Primers used to identify deletion mutant:

Elt-3CC1F: 5'-ctgactgttttgaggtctgctcaaaaacaac-3'
Elt-3CC1B: 5'-caaattgaattttgatgacattgcacggc-3'

Sequence of *ir79* deletion:

tctggtagctcacttgctcctgggaaggtagcagtactatatatagtgcatgcgcaattcattacctccctctttcattaaaaataatcttaaa
tattttttttgttgatgatagataattttttttgtagtatttgctttttactgactgttttgaggtctgctcaaaaacaacaaaactgtttttttctccattttt
ggtaccttttcgaaatcgatcgtttattatcagttgtttttcagacactaacaacaaaatgaatactcaaagtcctctcgggattcaaaccta
ctaccca

(DELETION, 2916 base pairs)

tgggagccacacaattctcccaccaacagactgtgcaactcaatttgctagatctcagcggcttctccacattcaactcgtgtgatccag
ctctaccattaatgcaccagtcaccttctcaacaaaattatgtgatgccacattcaacttttacaccacctccacaagatccattggtcgct
gaacaaaaaccacttgtcaaaaagcgaatggctgtaagttttgtttttttttttttgaaaacgggttttattgattttccttgttgacagaaaaca
gtgatttcagaaaaacaaagag