

Fig. S1. Egg production decreases in response to chronic exposure of adult

Oregon-R-C and Canton-S females to suboptimal temperatures. (A,B)

Representative graph showing daily temperature and humidity measurements for three separate 20-day experiments performed at different times of the year. (C,D) Average number of eggs laid per female per day over time upon chronic exposure of adult Oregon-R-C (C) or Canton-S (D) females to 18° C (cold) or 29° C (warm) compared to 25° C controls. Data shown as mean \pm s.e.m. from six replicates. **p < 0.01; ****p < 0.0001, F-test of third order polynomial fitted curves using 25° C as control.

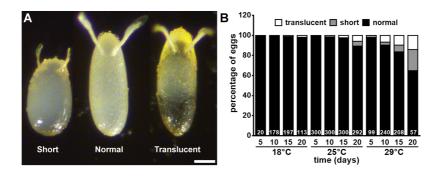


Fig. S2. Chronic exposure of females to 29°C has a small effect on eggshell morphology. (A) Examples of eggshell phenotypes observed in eggs laid by *y w* females maintained at 29°C for 20 days. Scale bar, 100 μm. (B) Frequency of eggs showing normal, short, or translucent eggshell phenotypes as in (A) laid by *y w* females maintained at 18°C, 25°C, or 29°C for five, 10, 15, or 20 days. Numbers of eggs analyzed are shown inside bars. Data represent one experiment.

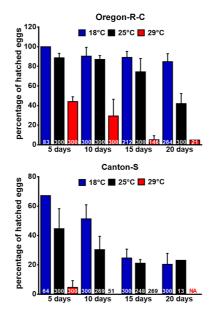


Fig. S3. Hatching rates of eggs laid by Oregon-R-C and Canton-S females maintained at suboptimal temperatures are affected. Percentage of hatched eggs laid by Oregon-R-C (top), or Canton-S (bottom) females maintained at 18°C, 25°C, or 29°C for five, 10, 15, or 20 days. No eggs were obtained from Canton-S females incubated at 29°C for 20 days (NA). Data shown as mean ± s.d. for one experiment with three replicates.

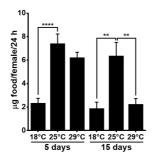


Fig. S4. Food consumption by adult females is influenced by temperature.

Graph showing total food consumption per female per day for y w females incubated at 18°C, 25°C, or 29°C for five or 15 days, based on the Consumption-Excretion assay. (See text for details.) Data shown as mean \pm s.e.m. from three independent experiments. **p < 0.01; ****p < 0.0001, Unpaired two-tailed t-test using 25°C as control.

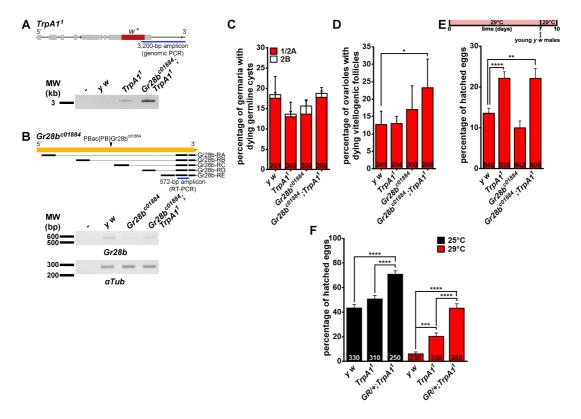


Fig. S5. Canonical warm temperature sensors do not play a major role in mediating the effects of 29°C exposure on oogenesis. (A) Verification of TrpA1¹ allele (originally generated by ends-out homologous recombination (Kwon et al., 2008)) by genomic PCR. The 3,200-bp diagnostic PCR product in indicated in blue. (See Methods for details.) (B) Verification of *Gr28b*^{c01884} allele by RT-PCR from female heads (Thorne and Amrein, 2008). The 572-bp RT-PCR product, which reflects expression of all Gr28b isoforms (black lines and bars), is indicated in blue. (See Methods for details.) (C) Percentage of germaria containing Apoptag-positive dying cysts in Region 1/2A or Region 2B from y w control, TrpA1¹ homozygous, Gr28b^{c01884} homozygous, or Gr28b^{c10084}; TrpA1¹ double homozygous females incubated with y w males for 10 days at 29°C. Numbers of germaria analyzed are shown inside bars. Data shown as mean ± s.e.m. from three independent experiments. No statistically significant differences, Chisquare test. (D) Frequencies of ovarioles containing dying vitellogenic follicles in same females as in (C). Numbers of ovarioles analyzed are shown inside bars. Data shown as mean \pm s.e.m. from three independent experiments. *p < 0.05, Chi-Square test using y w as control. (E) Quantification of effect of 29°C on oocyte quality in females of same

genotypes as in (C) and (D). Control and experimental females (with y w males) were incubated at 29°C for seven days, then two-day-old y w males replaced original males followed by incubation for three additional days at 29°C prior to collection of eggs laid within the last 24 hours and hatching rate quantification. Numbers of eggs analyzed shown inside bars. **p < 0.01, ****p < 0.0001, Unpaired two-tailed t-test using y w as control.(F) Quantification of oocyte quality in y w or homozygous $TrpA1^{1}$ females with or without a TrpA1 genomic rescue (GR) incubated with y w males for 10 days at 29°C as in (E) or maintained at the 25°C control temperature. These results show that the $TrpA1^{1}$ mutation is not responsible for the observed phenotype. It is conceivable that remaining genetic background differences cause the observed differences in hatching rates (despite the fact that the $TrpA1^{1}$ mutant and rescue transgenes were backcrossed into the isogenized y w). Numbers of eggs analyzed shown inside bars. ***p < 0.001, *****p < 0.0001, Unpaired two-tailed t-test.