

Table S1. Primer sequences used in this study.

Loci	Forward	Reverse	Purpose
<i>HBB</i>	CTCCTGGGCAACGTGATAGT	GGTTCAGAGGAAAAAGGGCTCCTCCT	Copy number
<i>HTT</i>	ATGAAGGCCTTCGAGTCCCTCAAGTCCTTC	CGGCGGCGGTGGCGGTTGCTGTTGCTGCTG	CAG sizing
<i>MT-RNR1</i>	TCGCAACTGCCTAAAACCTCA	GAATTGGCAAGGGTTGGTAA	mtDNA damage/copy number/mutation frequency
<i>NDUFA9</i>	GTTGTGAATGGTGCTAACTGCT	ACCAGAGACAATAAAGCAGAGGAG	nDNA damage

The table shows sequences of primers used for various experimental methods as indicated in right column.

Table S2. Information about antibodies used in supplementary western blotting (Fig.S3)

Target	Name	Manufacturer	Catalogue no.	Species raised in	Dilution
PDH E1 α , PDH E1 β , PDH E2, PDH E2/E3bp, OSCP	PDH antibody cocktail	Mitosciences	MSP02	Mouse	1:2000
	Complex III subunit Core 2 monoclonal antibody	Mitosciences	MS304	Mouse	1:10000
NDUFA9	Anti-NDUFA9 antibody [20C11B11B11]	Abcam	ab14713	Mouse	1:3000
SDH70	Anti-SDHA antibody [2E3GC12FB2AE2]	Abcam	ab14715	Mouse	1:5000
SDH30	Complex II subunit 30 kDa	Mitosciences	MS203		1:2000
CORE1	Anti-Ubiquinol-Cytochrome C Reductase Core Protein I antibody [16D10AD9AH5]	Abcam	ab110252	Mouse	1:5000
COX1	Anti-MTCO1 antibody [1D6E1A8]	Abcam	ab14705	Mouse	1:2000
COX5A	Anti-COX5A antibody [6E9B12D5]	Abcam	ab110262	Mouse	1:2000
ATPA	Anti-ATP5A antibody [7H10BD4F9]	Abcam	ab110273	Mouse	1:1000
Mitofilin	Anti-Mitofilin antibody [2E4AD5] - Mitochondrial Marker	Abcam	ab110329	Mouse	1:1000
Aconitase	Anti-Aconitase 2 antibody [6F12BD9]	Abcam	ab110321	Mouse	1:2000
OPA1	Purified Mouse Anti-OPA1 Clone 18/OPA1	BD Biosciences	612606	Mouse	1:2000
Porin	Anti-VDAC1/ Porin antibody [20B12AF2]		ab14734	Mouse	1:2000
ATP β	Anti-ATPB antibody [3D5] - Mitochondrial Marker	Abcam	ab14730	Mouse	1:1000
COX2	Anti-MTCO2 antibody [12C4F12]	Abcam	ab110258	Mouse	1:10000
CS	Anti-Citrate synthetase antibody [2H8BB6]	Abcam	ab128564	Mouse	1:2500
GAPDH	Anti-GAPDH antibody [6C5]	Abcam	ab8245	Mouse	1:3333
MRPS31	Anti-MRPS31 antibody [EPR10707]	Abcam	ab167406	Rabbit	1:3333
Anti-rabbit IgG	Anti-Rabbit IgG (whole molecule)-Peroxidase antibody produced in goat	Sigma-Aldrich	A0545	Goat	1:2500
Anti-mouse IgG	Anti-Mouse IgG (whole molecule)-Peroxidase antibody produced in goat	Sigma-Aldrich	A8924	Goat	1:2500

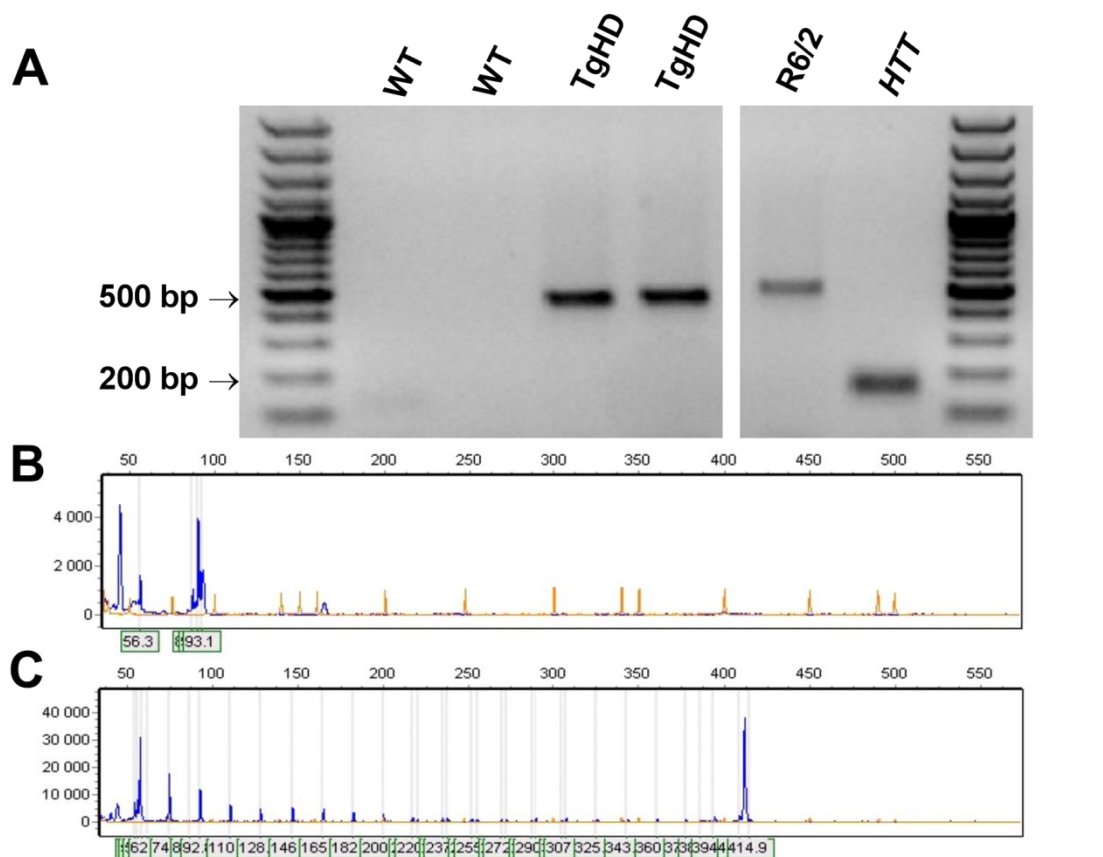


Fig. S1. Validation of TgHD minipig model. (A) PCR-mediated genotyping of the TgHD model transgene. Electrophoretic separation of PCR products made using *HTT* specific primers confirmed the presence of the expanded CAG repeat in TgHD minipigs. (B) Size determination of CAG repeat tracts. Capillary electrophoresis and fragment analysis using size standards (yellow) and GeneMapper® software confirmed the absence of polyQ *mHTT* in WT minipigs (C) and showed presence of *mHTT* with ~121 CAG repeats in TgHD minipigs. Representative samples shown.

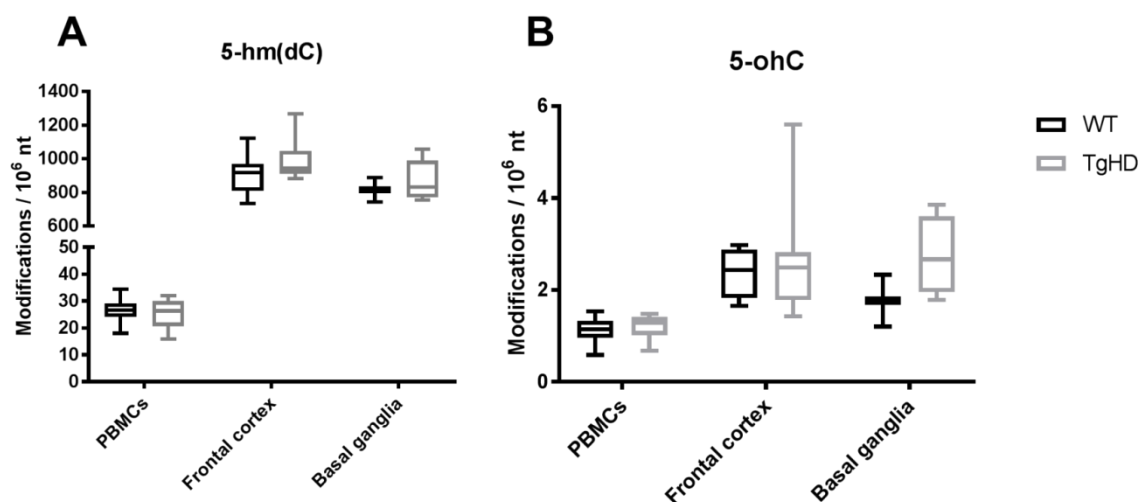


Fig. S2. Base modifications in the TgHD minipig model. LC-/MS/MS analysis of DNA showed normal levels of 5-hm(dC) and 5-ohC in all tissues (A, B). Student's t-test. Box plot whiskers indicate minimum to maximum values, with hinges representing the 25th and 75th percentile and median indicated by the center line. Sample sizes: PBMCs: WT n=10, TgHD n=6; frontal cortex: WT n=10, TgHD n=7; basal ganglia: WT n=2, TgHD n=5.

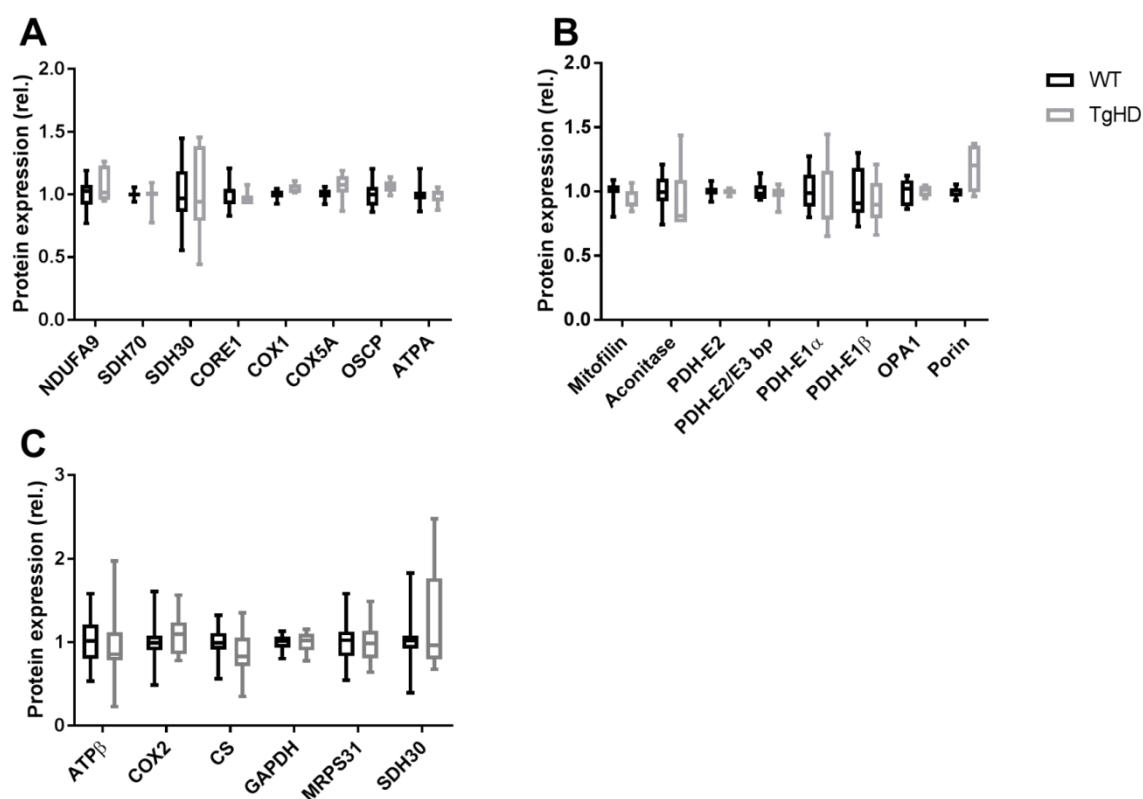


Fig. S3. Mitochondria-related protein expression in the TgHD minipig model.

Western analysis showed normal levels of mitochondria associated proteins in frontal cortex (A,B) and PBMCs (C). Box plot whiskers indicate minimum to maximum values, with hinges representing the 25th and 75th percentile and median indicated by the center line. Student's t-test, adjusted for multiple comparisons using the Holm-Sidak method. Sample sizes: Frontal cortex: WT n=9, TgHD n=7, except OPA1 and Porin WT n=6, TgHD n=4; PBMCs: WT n=13, TgHD n=13, except CS WT n=13, TgHD n=12, MRPS31 WT n=11, TgHD n=12, SDH30 WT n=13, TgHD n=12.

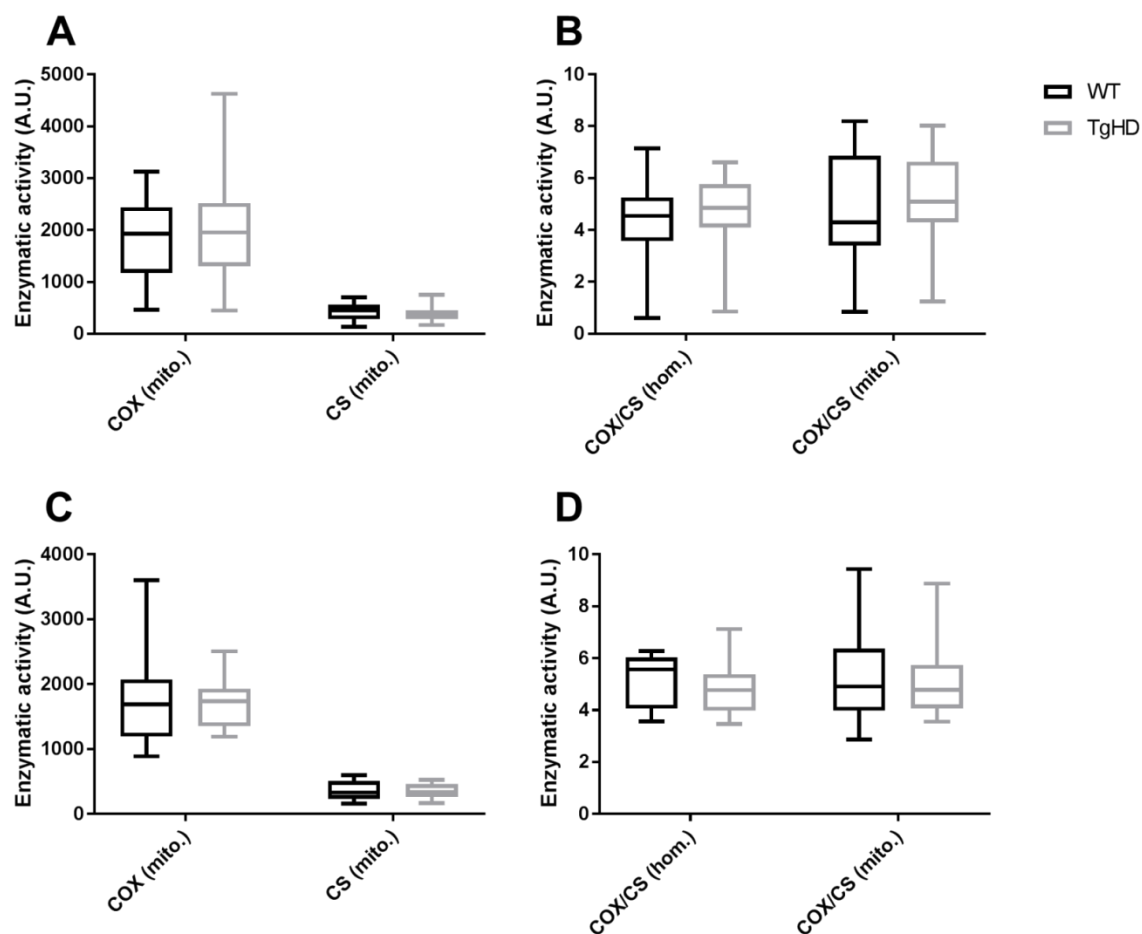


Fig. S4. Cytochrome c oxidase (COX) and citrate synthase (CS) activity in homogenates and isolated mitochondria from TgHD minipig brain. COX and CS activity unchanged in mitochondrial isolates in frontal cortex (A) and after normalizing to CS (B). COX and CS activity unchanged in mitochondrial isolates in basal ganglia (C) and after normalizing to CS (D). Arbitrary units (A.U.) represent enzymatic activity as nmol/min/mg. Student's t-test. Box plot whiskers indicate minimum to maximum values, with hinges representing the 25th and 75th percentile and median indicated by the center line. Sample sizes: Frontal cortex: COX (mito.) WT n=19, TgHD n=21, CS (mito.) WT n=19, TgHD n=21, COX/CS (hom.) WT n=19, TgHD n=21, COX/CS (mito.) WT n=19, TgHD n=21; basal ganglia: COX (mito.) WT n=10, TgHD n=9, CS (mito.) WT n=10, TgHD n=9, COX/CS (hom.) WT n=10, TgHD n=9, COX/CS (mito.) WT n=10, TgHD n=9. «mito»: isolated mitochondria; «hom»: homogenate.

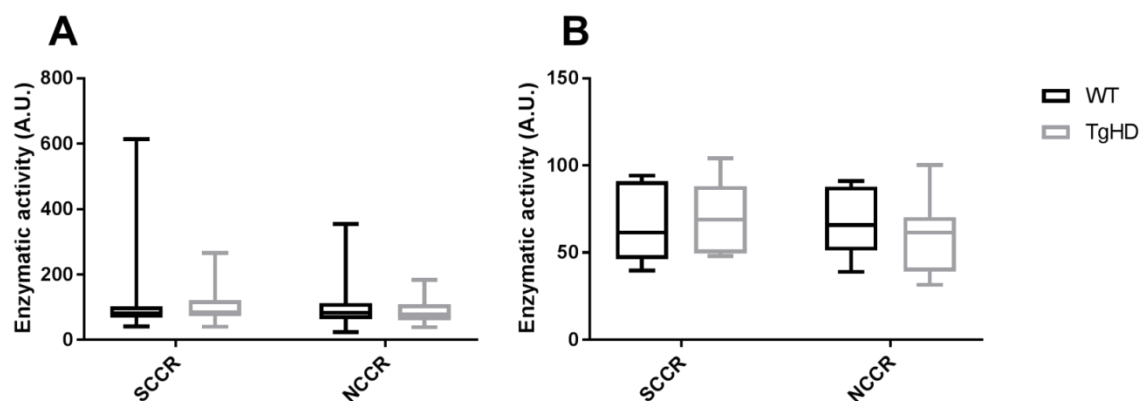


Fig. S5. Coupled activity of mitochondrial complex II+III and I+III in TgHD minipig model brain. SCCR activity (II+III) and NCCR activity (I+III) in frontal cortex (A) and basal ganglia (B) showed normal levels compared to WT animals. Arbitrary units (A.U.) represent enzymatic activity as nmol/min/mg. Student's t-test. Box plot whiskers indicate minimum to maximum values, with hinges representing the 25th and 75th percentile and median indicated by the center line. Sample sizes: Frontal cortex: SCCR WT n=19, TgHD n=21, NCCR WT n=19, TgHD n=21; basal ganglia: SCCR WT n=10, TgHD n=9, NCCR WT n=10, TgHD n=9. SCCR- succinate cytochrome c oxidase, NCCR- rotenone sensitive NADH cytochrome c oxidase.