

Fig. S1. Toxicity of $A\beta$ alone. (A) Ten-fold dilutions of exponentially growing cultures of BY4742 cells transformed with plasmids carrying the different chimeric constructions under the GAL10 promoter were spotted onto SD (-) or SG (+) agar supplemented with 20 mg/l histidine, 20 mg/l lysine and 60 mg/l leucine. The cells were incubated at 30°C for 3 days. (B) BY4742 cells expressing the different chimeric proteins (6 hours of expression) were collected for total-protein extracts. Equal quantities of proteins were separated by SDS-PAGE on a 12% polyacrylamide gel, transferred onto a nitrocellulose membrane and exposed to anti- $A\beta$ (Tebu) antibodies.

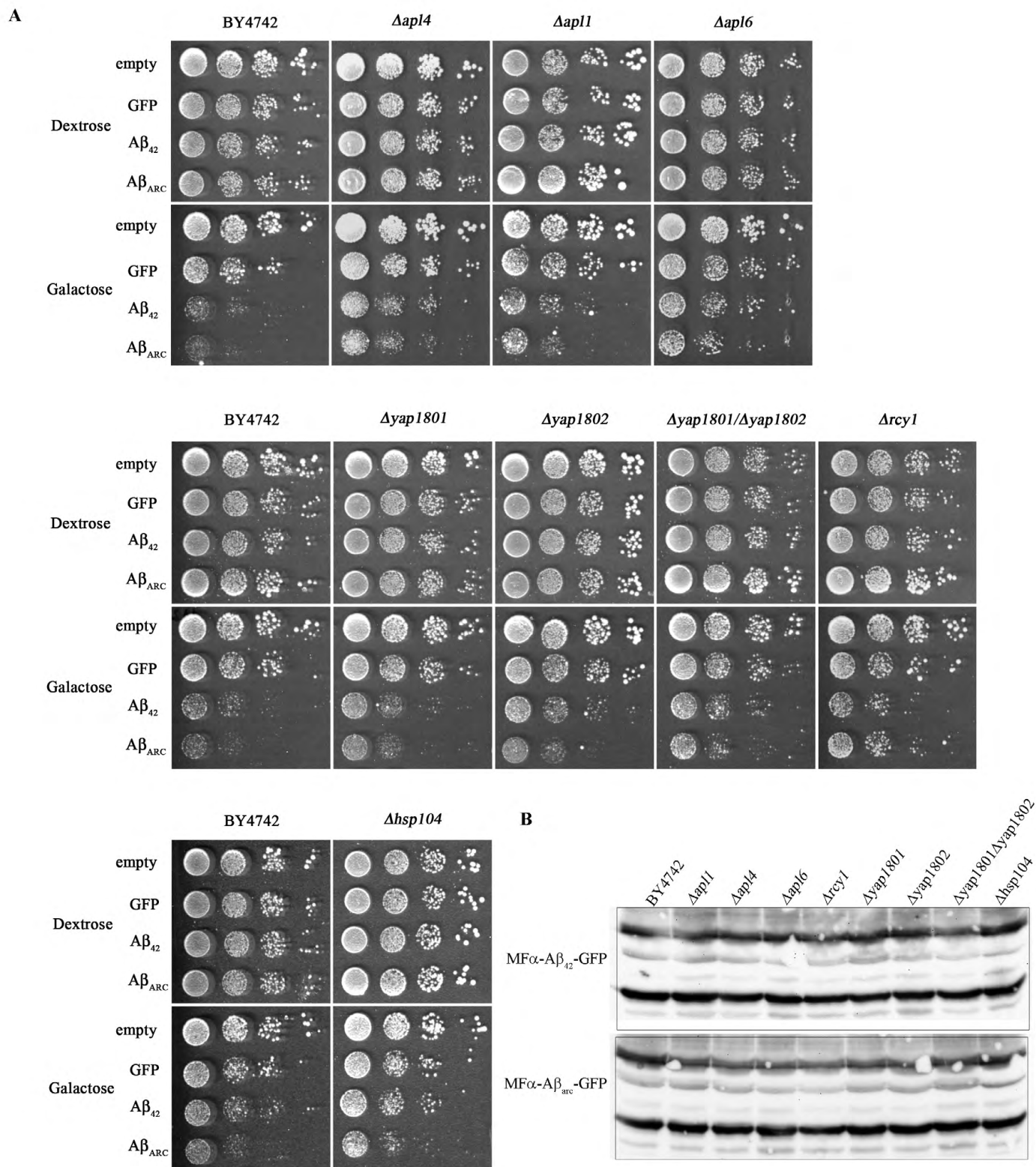


Fig. S2. Yeast mutants modulate A β toxicity without changing its protein level. (A) Ten-fold dilutions of exponentially growing cultures of deleted strains transformed with plasmids carrying the different chimeric constructions under the GAL10 promoter were spotted on the same plate onto SD (-) or SG (+) agar supplemented with 20 mg/l histidine, 20 mg/l lysine and 60 mg/l leucine. (B) Cells expressing the different chimeric proteins (6 hours of expression) were collected for total-protein extracts. Equal quantities of proteins were separated by SDS-PAGE on a 12% polyacrylamide gel, transferred onto a nitrocellulose membrane, and then exposed to monoclonal anti-GFP antibodies (Sigma).

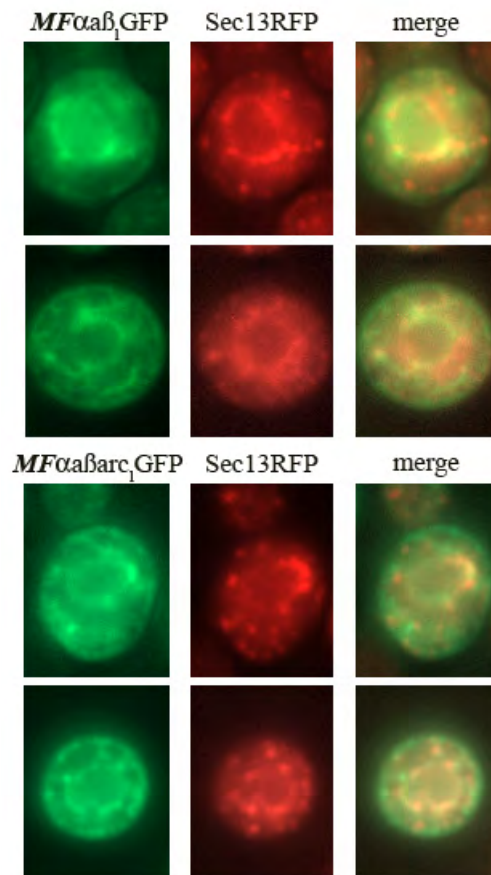


Fig. S3. Aβ-GFP species colocalize with ER. Wild-type or RFP-tagged strains were grown for 6 hours in SG liquid medium supplemented with 0.67% casaminoacids to induce the expression of the chimeric proteins and were then examined by epifluorescence microscopy.

Table 1. Yeast strains used in this study

Name of strain	Genotype	Source
BY4742	<i>MATa, his3Δ1, leu2Δ0, ura3Δ0</i>	Eurocarf yeast deletion library
<i>Δapl4</i>	<i>MATa, his3Δ1, leu2Δ0, ura3Δ0, YPR029C::KanMX4</i>	Eurocarf yeast deletion library
<i>Δapl1</i>	<i>MATa, his3Δ1, leu2Δ0, ura3Δ0, YJR005W::KanMX4</i>	Eurocarf yeast deletion library
<i>Δapl2</i>	<i>MATa, his3Δ1, leu2Δ0, ura3Δ0, YKL135C::KanMX4</i>	Eurocarf yeast deletion library
<i>Δapl3</i>	<i>MATa, his3Δ1, leu2Δ0, ura3Δ0, YBL037W::KanMX4</i>	Eurocarf yeast deletion library
<i>Δapm1</i>	<i>MATa, his3Δ1, leu2Δ0, ura3Δ0, YPL259C::KanMX4</i>	Eurocarf yeast deletion library
<i>Δapm4</i>	<i>MATa, his3Δ1, leu2Δ0, ura3Δ0, YOL062C::KanMX4</i>	Eurocarf yeast deletion library
<i>Δgga2</i>	<i>MATa, his3Δ1, leu2Δ0, ura3Δ0, YHR108W::KanMX4</i>	Eurocarf yeast deletion library
<i>Δent5</i>	<i>MATa, his3Δ1, leu2Δ0, ura3Δ0, YDR153C::KanMX4</i>	Eurocarf yeast deletion library
<i>Δclc1</i>	<i>MATa, his3Δ1, leu2Δ0, ura3Δ0, YGR167W::KanMX4</i>	Eurocarf yeast deletion library
<i>Δrcy1</i>	<i>MATa, his3Δ1, leu2Δ0, ura3Δ0, YJL204C::KanMX4</i>	Eurocarf yeast deletion library
<i>Δyap1801</i>	<i>MATa, his3Δ1, leu2Δ0, ura3Δ0, YHR161C::KanMX4</i>	Eurocarf yeast deletion library
<i>Δyap1802</i>	<i>MATa, his3Δ1, leu2Δ0, ura3Δ0, YGR241C::KanMX4</i>	Eurocarf yeast deletion library
<i>Δhsp104</i>	<i>MATa, his3Δ1, leu2Δ0, lys2Δ0, ura3Δ0, YLL026W::KanMX4</i>	Eurocarf yeast deletion library
<i>SEC13-RFP</i>	<i>MATa, his3Δ1, leu2Δ0, lys2Δ0, ura3Δ0, YLR208W-RFP-KanMX6</i>	Peter Arvidson
<i>Δyap1801/Δyap1802</i>	<i>MATa, his3Δ1, leu2Δ0, ura3Δ0, YHR161C::KanMX4, YGR241C::KanMX4</i>	This study

Table S1. Yeast strains used in this study.**Table S2. Oligonucleotides used in this study**

Number	Sequence
792	AAATACACACACTAAATTACCGGATCCTATGGATGCAGAATTCCGACATG
794	ACCAGTGAATAATTCTTCACCTTTAGACATCGCTATGACAACACCGCCCACC
705	GGATGGCCAGGCAACTTTAG
856	GAATAATTCTTCACCTTTAGACATAGCTTCAGCCTCTCTTTTATC
858	GAATAATTCTTCACCTTTAGACATGGATCCGGTAATTTAGTGTGT
859	GTCATGTCGGAATTCTGCATCCATGGATCCGGTAATTTAGTGTGT
706	TTTACACTTTATGCTTCCGG
857	ATGTCTAAAGGTGAAGAATTATTC
860	ATGGATGCAGAATTCCGACATG

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