

PKC at a glance

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The protein kinase C (PKC) family is represented in all eukaryotes and in *Homo sapiens* comprises the related PKC α through PKC ι gene products (Dempsey et al., 2000). The kinase

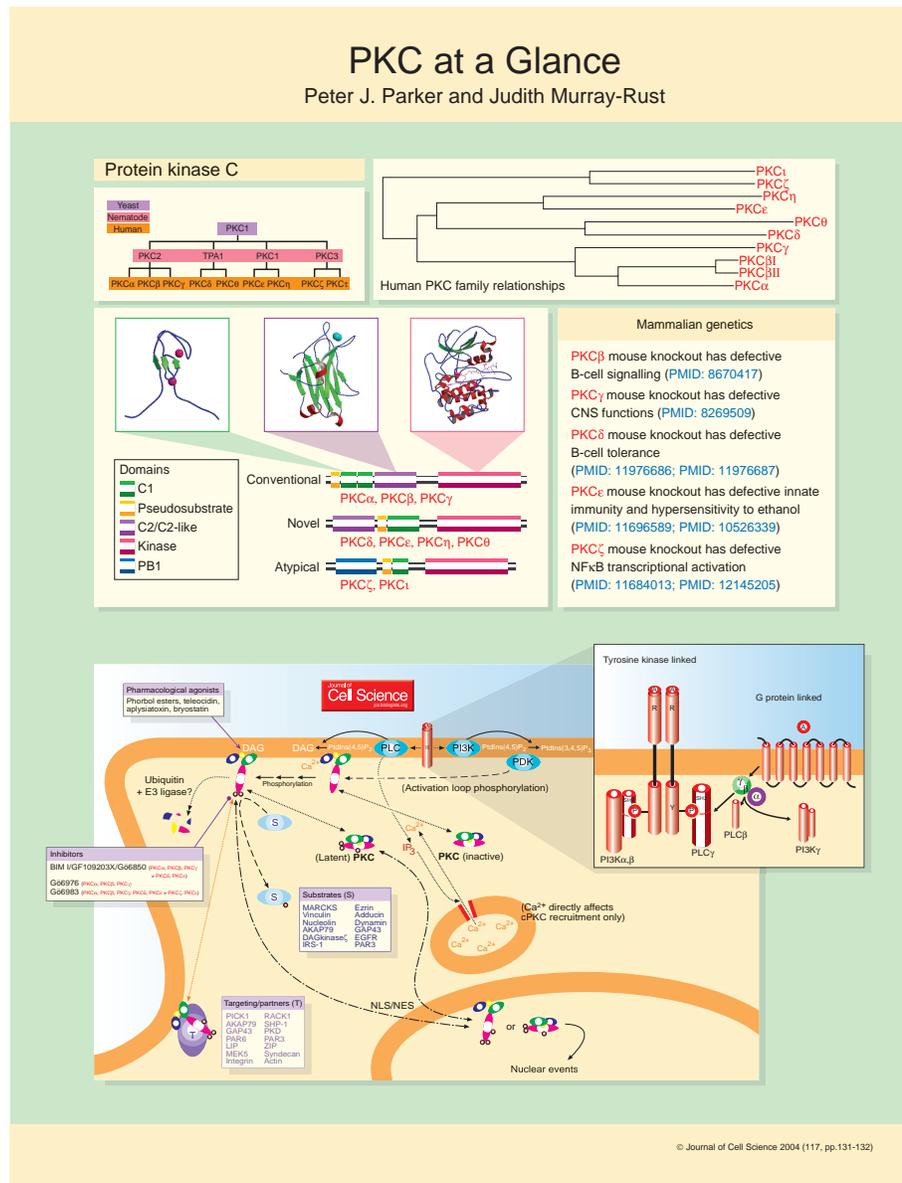
domains are closely related, as illustrated in the dendrogram, and form part of the AGC kinase superfamily. For clarity, the PKC-related kinases (PRK/PKN) are not included here; similarly excluded is the PKD/PKC μ family, members of which have distinct kinase domains but some related regulatory properties. The three PKC subgroups shown are structurally and functionally distinguished. The conventional, cPKC, isoforms (PKC α , PKC β and PKC γ) are diacylglycerol (DAG) sensitive and Ca²⁺ responsive (responding through an archetypal C2 domain). The novel, nPKC, isoforms (PKC δ , PKC ϵ , PKC η and PKC θ) are DAG sensitive but Ca²⁺ insensitive; their C2-related domains do not retain Ca²⁺-

coordinating residues. The atypical, aPKC, isoforms (PKC ζ and PKC ι/λ) have altered C1 domains and are not DAG sensitive; regulation occurs in part through the N-terminal PB1 domain. Structures for C1 domains and C2 domains have been determined (examples are illustrated for PKC δ), and kinase domain models based on the highly related PKA have been built (that for PKC α is shown here).

Genetic studies in the mouse indicate that particular PKC isoforms are essential in a number of specific contexts, some of which are highlighted here (Abeliovich et al., 1993; Leitges et al., 1996; Hodge et al., 1999; Castrillo et al., 2001; Leitges et al., 2001; Martin et al., 2002; Mecklenbrauker et al., 2002; Miyamoto et al., 2002). However, this is a very much narrower phenotypic profile than is reflected in the rather broad pattern of expression for most of these proteins. Thus it is likely that some functional redundancy exists.

The activation of cPKC and nPKC isoforms typically involves recruitment to membranes and interaction with or allosteric activation by DAG. Agonist-induced production of DAG is effected by multiple mechanisms. For receptor tyrosine kinases and receptors linked to non-receptor tyrosine kinases, this involves the recruitment of phosphatidylinositol (4,5)-bisphosphate [PtdIns(4,5)P₂]-specific phospholipases C γ _{1/2} (PtdIns-PLC γ _{1/2}) through their SH2 domains. For serpentine receptors, coupling is effected by PtdIns-PLC β family members through G α_q -GTP and G $\beta\gamma$ family member allosteric interactions. Additionally, PtdIns-PLC ϵ is a target for the small GTPase Ras (not illustrated). For cPKC isoforms, the initial recruitment event is the Ca²⁺-sensitive step, which is driven by C2 domain interactions with Ca²⁺ and anionic-phospholipids. For nPKC isoforms, no equivalent mechanism promoting interaction with DAG has been elucidated. For aPKC isoforms activation can be driven in part by interaction with the Cdc42-GTP-Par6 complex, which binds the PB1 domain of aPKC. In each case, the allosteric effects of these lipids/proteins on PKC isoforms lead to a loss of the

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inhibition exerted by the inhibitory pseudosubstrate sequence that otherwise occupies the active site.

For optimum catalytic output all the PKC family members appear to require phosphorylation in their activation loops at a TFCGT motif conserved in many members of the AGC kinase superfamily (Parekh et al., 2000). This is catalysed by phosphoinositide-dependent kinase 1 (PDK1), which is itself recruited to membranes by PtdIns(3,4,5)P₃. The upstream enzyme PI 3-kinase is, similarly to PtdIns-PLC, coupled to receptors through tyrosine kinases, heterotrimeric G proteins and/or Ras proteins. The PDK1-dependent activation loop phosphorylation occurs in conjunction with C-terminal phosphorylations to lock the kinase domains in their active conformations; there remains some debate as to the order and requirements for these phosphorylations.

In the membrane-bound, open, active conformations, substrate phosphorylation is efficient. Some examples of PKC substrates are shown and include a number of cytoskeleton-associated proteins that contribute to localised cytoskeletal reorganisation (Jaken and Parker, 2000). For many substrates – the so-called STICKs (substrates that interact with C-kinases) – direct kinase-substrate interaction contributes to specificity/efficiency of action. Some of the complexes involve scaffolding proteins termed RICKs (receptors for inactive C-kinases), which can operate prior to PKC activation, restricting membrane and substrate access; other complexes form post-

activation, such as RACKs (receptors for activated C-kinases), determining substrate access (Mochly-Rosen and Gordon, 1998).

Pharmacological probes for the action of PKC family proteins include membrane-permeant allosteric activators (e.g. phorbol esters) and catalytic inhibitors. In general these do not distinguish isoforms and, furthermore, they can modulate other C1-domain-containing proteins (e.g. phorbol esters) or protein kinases (catalytic site inhibitors). However, their combined use is a helpful guide to PKC involvement in cellular processes.

PKC action can be localised to multiple compartments, including the plasma membrane, (recycling) endosomes, the Golgi and the nucleus. Location is determined in part by the scaffolds but also by targeting information intrinsic to individual isoforms – for example, nuclear localisation/export sequences (NLS/NES). The requirements for the individual isoforms are in part revealed by those non-redundant properties manifest in the mouse knockout phenotypes. It remains to be determined how specific actions relate to these patterns of behaviour at a molecular level.

References

Abeliovich, A., Chen, C., Goda, Y., Silva, A. J., Stevens, C. F. and Tonegawa, S. (1993). Modified hippocampal long-term potentiation in PKC gamma-mutant mice. *Cell* **75**, 1253-1262.
 Castrillo, A., Pennington, D. J., Otto, F., Parker, P. J., Owen, M. J. and Bosca, L. (2001). Protein kinase Cepsilon is required for macrophage activation and defense against bacterial infection. *J. Exp. Med.* **194**, 1231-1242.

Dempsey, E. C., Newton, A. C., Mochly-Rosen, D., Fields, A. P., Reyland, M. E., Insel, P. A. and Messing, R. O. (2000). Protein kinase C isozymes and the regulation of diverse cell responses. *Am. J. Physiol. Lung Cell Mol. Physiol.* **279**, L429-L438.
 Hodge, C. W., Mehmert, K. K., Kelley, S. P., McMahon, T., Haywood, A., Olive, M. F., Wang, D., Sanchez-Perez, A. M. and Messing, R. O. (1999). Supersensitivity to allosteric GABA(A) receptor modulators and alcohol in mice lacking PKCepsilon. *Nat. Neurosci.* **2**, 997-1002.
 Jaken, S. and Parker, P. J. (2000). Protein kinase C binding partners. *Bioessays* **22**, 245-254.
 Leitges, M., Schmedt, C., Guinamard, R., Davoust, J., Schaaf, S., Stabel, S. and Tarakhovskiy, A. (1996). Immunodeficiency in protein kinase Cbeta-deficient mice. *Science* **273**, 788-791.
 Leitges, M., Sanz, L., Martin, P., Duran, A., Braun, U., Garcia, J. F., Camacho, F., Diaz-Meco, M. T., Rennert, P. D. and Moscat, J. (2001). Targeted disruption of the zetaPKC gene results in the impairment of the NF-kappaB pathway. *Mol. Cell* **8**, 771-780.
 Martin, P., Duran, A., Minguet, S., Gaspar, M. L., Diaz-Meco, M. T., Rennert, P., Leitges, M. and Moscat, J. (2002). Role of zeta PKC in B-cell signaling and function. *EMBO J.* **21**, 4049-4057.
 Mecklenbrauker, I., Saijo, K., Zheng, N. Y., Leitges, M. and Tarakhovskiy, A. (2002). Protein kinase Cdelta controls self-antigen-induced B-cell tolerance. *Nature* **416**, 860-865.
 Miyamoto, A., Nakayama, K., Imaki, H., Hirose, S., Jiang, Y., Abe, M., Tsukiyama, T., Nagahama, H., Ohno, S., Hatakeyama, S. and Nakayama, K. I. (2002). Increased proliferation of B cells and auto-immunity in mice lacking protein kinase Cdelta. *Nature* **416**, 865-869.
 Mochly-Rosen, D. and Gordon, A. S. (1998). Anchoring proteins for protein kinase C: a means for isozyme selectivity. *FASEB J.* **12**, 35-42.
 Parekh, D. B., Ziegler, W. and Parker, P. J. (2000). Multiple pathways control protein kinase C phosphorylation. *EMBO J.* **19**, 496-503.

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