

Table S3. Primers for PCR identification of deletion mutants

Target gene	Primer designation	Sequence
<i>dgaI</i> ⁺	Dga1kc5 ^a	5'-CGATAGTAGTCAATACCAAG-3'
	KanI5 ^b	5'-AATCAAATGTTAGCGTGAT-3'
	Dga1I3 ^c	5'-GCAATTGAAATCGCGATAC-3'
	Pca1kc3 ^a	5'-GTGGTTAGGCAATAAAGC-3'
<i>pcaI</i> ⁺	KanI3 ^b	5'-ACCTCAGTGGCAAATCCTA-3'
	PomYCA1-5 ^d	5'-GGAATTCCATATGAGCTACAACCTCCAATCC-3'
	PomYCA1-3 ^e	5'-GCGGATCCCTATAAAACCATGGCAAGA-3'
	Aif1kc5 ^a	5'-AGAGTAGACATTGTGTATG-3'
<i>aifI</i> ⁺	KanI3 ^b	5'-ACCTCAGTGGCAAATCCTA-3'
	Aif1I3 ^c	5'-GTGTTTCGAGCACGAACTAG-3'
	Rad9kc5 ^a	5'-GGTGCATGATTATTTTAC-3'
<i>rad9</i> ⁺	Ura4I5 ^b	5' GCTCCTACAACATTACCAAC-3'
	Rad9I3 ^c	5'-ACAGGACGGAATAAGGATG-3'
	Apg6kc5 ^a	5'-GGAAATGTTACCAAGTTG-3'
<i>apg6</i> ⁺	Ura4I5 ^b	5' GCTCCTACAACATTACCAAC-3'
	Apg6I3 ^c	5'-GAATGACACCTCTGACACA-3'
	Pck1kc5 ^a	5'-GTACTATGAGGAGATTGCA-3'
<i>pck1</i> ⁺	Ura4I5 ^b	5' GCTCCTACAACATTACCAAC-3'
	Pck1I3 ^c	5'-GACTCTCCCGTTCTACTT-3'
	Pck2kc5 ^a	5'-CTAATAGTTAGCAAGCA-3'
<i>pck2</i> ⁺	Ura4I5 ^b	5' GCTCCTACAACATTACCAAC-3'
	Pck2I3 ^c	5'-AGGGCGCCATGAATAACTG-3'
	Bzz1kc3 ^a	5'-TTCGGGCTTGAGCTCCAAC-3'
<i>bzz1</i> ⁺	Ura4I3 ^b	5'-GAGAAAGAATGCTGAGTAG-3'
	Bzz1I5 ^c	5'-CTTCAAGGCCATTGCGCGG-3'

^a Forward primers for positive and negative PCR, located ~550bp upstream of the coding regions.

^b Reverse primers for positive PCR, located within the respective markers.

^c Reverse primers for negative PCR, located within the respective coding regions.

^{d-e} Primers for negative PCR checking of *pcaI* disruption mutants.