

Fig. S2. Phylogenetic analysis of *ETU1*. (A) Sequences were identified and aligned for the 5 conserved tubulin family members. For α , β , γ , and δ -tubulin, 5 representative sequences were used. In the case of ϵ -tubulin we used as many known or predicted sequences as could be identified. The sequences were aligned using MUSCLE (Edgar, 2004) and the phylogeny was determined with PhyML (Guindon et al., 2010) using Phylogeny.fr (Dereeper et al., 2008). *Tetrahymena* ϵ -tubulin clustered with other ϵ -tubulin proteins from other organisms indicating that it is indeed a unique branch of the tubulin superfamily. *Tt*=*T. thermophila*, *Cr*=*C. reinhardtii*, *Xl*=*X. laevis*, *Hs*=*H. sapiens*, *Dr*=*D. rerio*, *Mm*=*M. musculus*, *Gl*=*G. lamblia*, *Pf*=*P. falciparum*, *Tg*=*t. gondii*, *Lm*=*Leishmania major*, *Tb*=*T. brucei*, *Ng*=*N. gruberi*, *Pt*=*P. tetra*, *Ci*=*C. intestinalis*, *Ta*=*T. adhaerens*, *Sp*=*S. purpuratus*, *Nv*=*N. vectensis*, *Gg*=*G. gallus*. (B) The *Tetrahymena* α , β , γ , δ , and ϵ -tubulin sequences were aligned. Although the family members share a high degree of identity and similarity, unique features are visible. ϵ -tubulin contains two unique and conserved insertions at amino acids 60 and 221 of 5 and 36 amino acids, respectively.

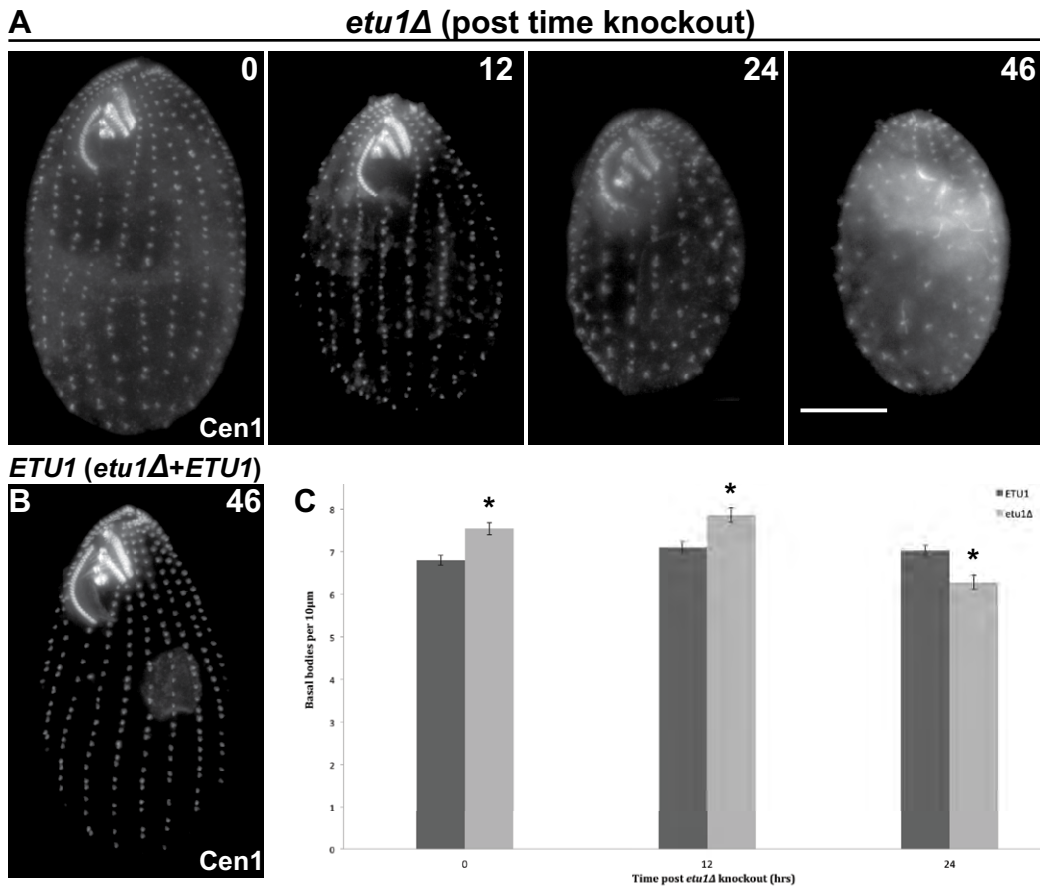


Fig. S3. *ETU1* is essential for viability and maintenance of basal body number. (A) *etu1Δ* cells were fixed and stained for centrin (Cen1) at the times indicated post knockout. Scale=10 μm. (B) An *etu1Δ* strain was mated to a wild-type strain to create an *ETU1* strain. Scale=10 μm. (C) Basal body density was measured within 10 μm of a cortical row. At 0 and 12 hrs, the *etu1Δ* strain had a significantly higher basal body density than *ETU1*. 24 hours after knockout the *etu1Δ* strain showed a significant decrease in basal body numbers. $n=100$ cortical rows from 20-22 cells. Error bars=s.e.m. * $P<0.01$.

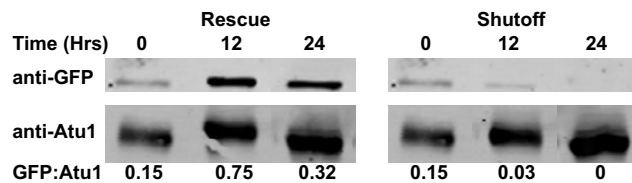


Fig. S4. Western blot analysis of *GFP-ETU1* expression. Cells were grown in the presence (rescue) or absence (shutoff) of CdCl_2 and then samples were taken at the indicated times after depletion of GFP-Etu1p and blotted for GFP and Atu1. The ratio of GFP to Atu1 signal was calculated and is indicated below the corresponding time.

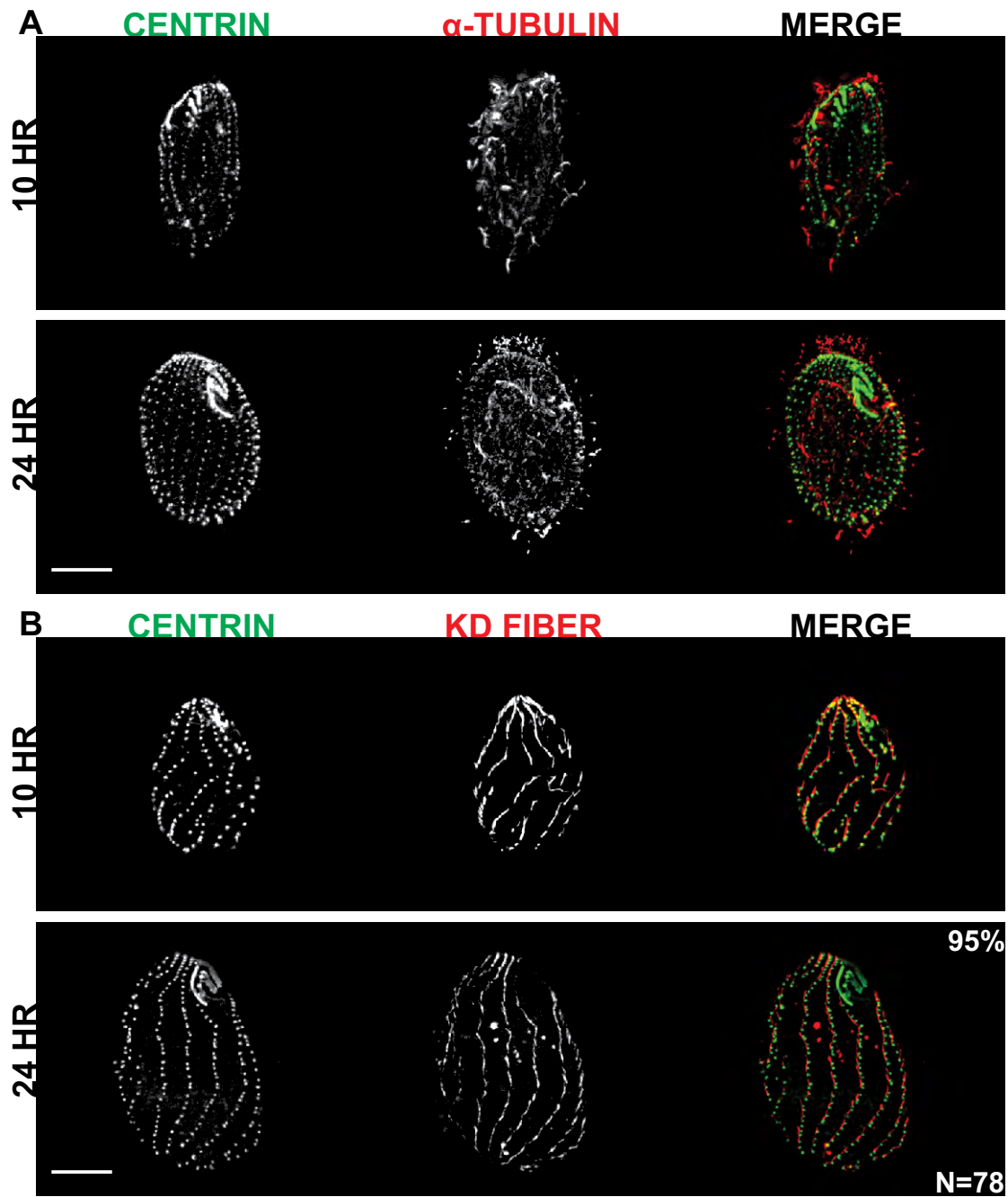


Fig. S5. Depletion of GFP-Etu1p is reversible. After 24 hrs of depletion of GFP-Etu1p, CdCl₂ was re-introduced to induce expression of *GFP-ETU1*. Cells were then fixed after 10 hrs and 24 hrs of induction. (A) Cells were stained for Cen1 (centrin) and Atu1 (α -tubulin). Basal body organization is nearly fully recovered after 24 hrs and the cells have cilia. (B) Staining of the KD fiber reveals that basal bodies have also regained proper orientation after 24 hrs (95%). $n=78$ cells. Scale=10 μ m.

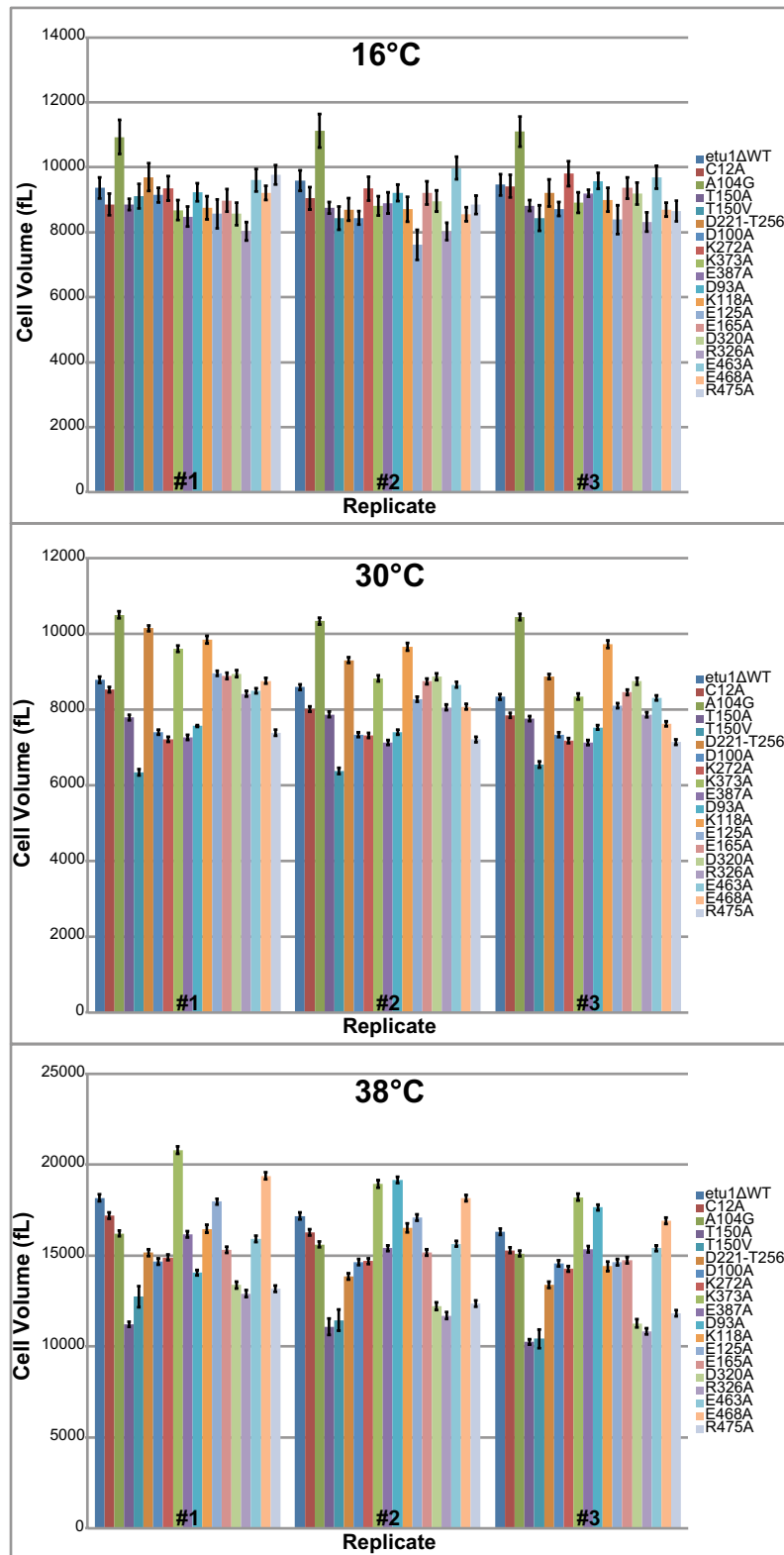


Fig. S6. Size analysis of *ETU1* mutants. *ETU1* mutant cells were grown at 16°C (A), 30°C (B), and 38°C (C) for 24 hrs then assayed for cell size using a Z2 Coulter counter. Cell counts ranged from 315-14,454 cells, depending on the strain and condition. Each mutant was assayed three times for each condition. Error bars=CI of 95%.

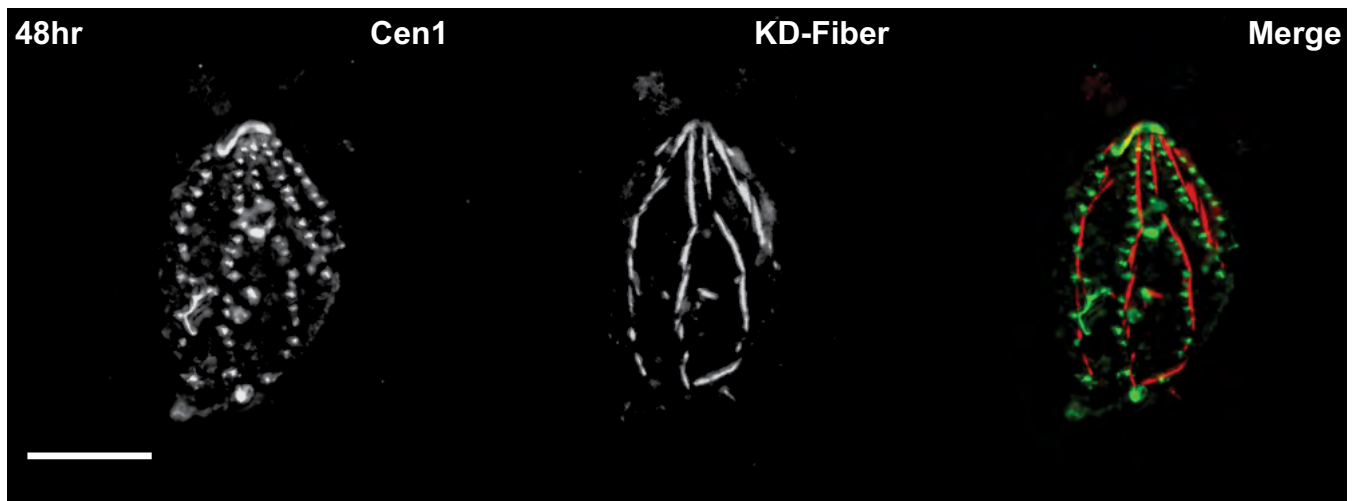


Fig. S7. Starved Etu1-T150V cells maintain basal body organization. Etu1-T150V cells lose basal body density but maintain organization within cortical rows as indicated by the KD fiber staining. This indicates that the misoriented basal bodies observed in cycling cells are likely newly assembled basal bodies that are misoriented during assembly. Scale=10 μm .