

SUPPLEMENTAL TABLES

**Table S1. Plasmids used in this study**

<b>Plasmid</b>	<b>Description</b>	<b>Ref</b>
Yep13-GFP-Bem3	GFP-Bem3, 2 $\mu$ <i>LEU2</i>	(Caviston et al., 2003)
pRS316-HA-mRFP-cSNC1	HA-mRFP-cSnc1, CEN <i>URA3</i>	(Robinson et al., 2006)
PHO5pr-RFP-Vps21	RFP-Vps21 expressed from <i>PHO5</i> promoter, CEN <i>TRP1</i>	(Markgraf et al., 2009)
pRS416-Cherry-Vps4 <sup>E233Q</sup>	Cherry-Vps4 <sup>E233Q</sup> expressed from Vps21 promoter, CEN <i>URA3</i>	(Davies et al., 2010)
pRS315-GFP-Sec4	GFP-Sec4, CEN <i>LEU2</i>	(Calero et al., 2003)
pRS315-GFP-Rga1	GFP-Rga1, CEN <i>LEU2</i>	(Caviston et al., 2003)
pRS315-Cdc24-GFP	GFP-Cdc24, CEN <i>LEU2</i>	Erfei Bi Lab
Gic1-GFPx3	<i>GIC1</i> in pRS316-P*(a derivative of pRS316) with triple GFP tag inserted in frame with construct. CEN <i>URA3</i> variant	(Takahashi and Pryciak, 2007)
pGAL1.416-Bem3-GFP	GFP-Bem3 expressed from a GAL1 promoter, CEN <i>URA3</i>	(Knaus et al., 2007)
pYES2.1/V5-HIS-TOPO-Sec15	Sec15 expressed from the GAL1 promoter, 2 $\mu$ <i>URA3</i>	This study
pYES2.1/V5-HIS-TOPO-Sro7	Sro7 expressed from the GAL1 promoter, 2 $\mu$ <i>URA3</i>	This study
pRS315-GFP-Sec4 <sup>Q79L</sup>	GFP- Sec4 <sup>Q79L</sup> , CEN <i>LEU2</i>	This study
pRS315-GFP-Sec4 <sup>S29N</sup>	GFP- Sec4 <sup>S29N</sup> , CEN <i>LEU2</i>	This study
Yep13-GFP-Bem3 <sup>PHm</sup>	GFP-Bem3 <sup>R644S, R645S, K647D</sup> 2 $\mu$ <i>LEU2</i>	This study
Yep13-GFP-Bem3 <sup>K1003A</sup>	GFP-Bem3 <sup>K1003A</sup> 2 $\mu$ <i>LEU2</i>	This study
Yep13-GFP-Bem3PX mutant	GFP-Bem3 <sup>Y524W, R578S, L580W, F581M</sup> 2 $\mu$ <i>LEU2</i>	This study
PX-PH domain-His6	pET28a HIS <sub>6</sub> -BEM3 <sup>491-774</sup>	This study
Yep13-HA-Bem3	HA-Bem3, 2 $\mu$ <i>LEU2</i>	This study
pGAL1.426-Bem3-HA-His6	Bem3 expressed from GAL1 promoter, 2 $\mu$ <i>URA3</i>	This study

pAD54-RFP-SEC4	RFP-Sec4 expressed from the ADH promoter, 2μ <i>LEU2</i>	(Aronov and Gerst, 2004)
pYES2.1/V5-HIS-TOPO-CaBem3	CaBem3 expressed from the GAL1 promoter, 2μ <i>URA3</i>	This study
pGAL1.426-Bem3 <sup>K1003A</sup> -HA-His6	Bem3 <sup>K1003A</sup> expressed from GAL1 promoter, 2μ <i>URA3</i>	This study

**Table S2. Strains used in this study**

<b>Name</b>	<b>Genotype</b>	<b>Source</b>
W303	<i>Mata ade2-1 his3-1 leu2-3112 trp1-1 ura3-1 can1-100</i>	Laboratory Strain
SEY6210	<i>MATa leu2-3,112 ura3-52 his3-200 trp1-901 lys2-801 suc2-9</i>	Laboratory Strain
BY4741	<i>MATa his3Δ1 leu2Δ0 met15Δ0 ura3Δ0</i>	Invitrogen
RH4344 ( <i>rcy1Δ</i> )	<i>Mata yjl204c::kanMX his4 leu2 ura3 lys2 bar1</i>	(Wiederkehr et al., 2000)
RLY 3090	<i>Mata BEM3-GFP::HIS5 his3Δ1;leu2Δ0;met15Δ0;ura3Δ0</i>	(Huh et al., 2003)
<i>vps29Δ</i>	<i>vps29::kanMX</i> in W303	This study
Bem3-GFP (Diploid)	<i>MATa/α his3Δ1/his3Δ1 leu2Δ0/leu2Δ0 lys2Δ0/lys2Δ0 met15Δ0/ met15Δ0 ura3Δ0/ura3Δ0</i> transformed with Yep13-GFP-Bem3	This study
<i>sla2Δ</i> [see Note1]	<i>MATa sla2:: HisMX leu2-3,112 ura3-52 his3-200 trp1-901 lys2-801 suc2-9</i>	(Stefan et al., 2005)
<i>BWY2595</i> ( <i>ent1Δ, ent2Δ, yap1801Δ, yap1802Δ</i> ) [see Note2]	<i>Mata leu2-3 ura3-52 his3-Δ200 trp1-Δ901 lys2-801 suc2- Δ9 ent1::LEU2 ent2::HIS3 yap1801::HIS3 yap1802::LEU2+ pBW0778 (ent1ENTH domain, CEN)</i>	Wendland lab
<i>bem3Δ</i>	<i>MATa bem3::kanMX his3Δ1 leu2Δ0 met15Δ0 ura3Δ0</i>	Hazbun Lab
DAO2C	<i>Mata ura3, leu2, met1,cdc3-6</i>	Haarer Lab
<i>sec4-8</i>	<i>Mata sec4-8 ura 3-52</i>	Novick Lab
<p><b>Note1:</b> <i>sla2Δ</i>: Sla2 knockout strain defective for endocytosis and proper actin cytoskeleton organization. Formation of actin comet tails associated with endocytic sites in these cells has been reported. (Kaksonen et. al., <i>Cell</i> <b>115</b>, 475–48 (2003).</p> <p><b>Note2:</b> Quadruple mutant strain with deletions of ENT1, ENT2, YAP1801, and YAP1802. The ENTH domain of Ent1 is expressed from a plasmid to maintain viability. These cells are defective in endocytosis and are inviable at 37°C.</p>		

<b>Table S3. Antibodies used in this study</b>				
<b>Host</b>	<b>Antigen</b>	<b>Clone</b>	<b>Source</b>	<b>Dilution</b>
Mouse	HIS <sub>6</sub>	6XHIS	Clontech, CA	1:5000
Mouse	HA	HA.11	Covance, NJ	1:2000
Rabbit	Bgl2	9937F	Schekman Lab	1:10,000
Mouse	Pma1	40B7	Encor Biotechnology Inc., FL	1:5000

## SUPPLEMENTAL REFERENCES

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## SUPPLEMENTARY FIGURE LEGENDS

**Figure S1.** (A) *sla2Δ* cells expressing GFP-Bem3 and RFP-tagged Ede1/Abp1, (B) GFP-Bem3-expressing *ent1Δ/ent2Δ /yap1801Δ/yap1802Δ* cells or (C) WT cells expressing GFP-Rga1, GFP-Cdc24 and GFP $\alpha$ 3-Gic1 from their respective endogenous promoters, were imaged at 100X using a FITC filter. Scale bars: 5 $\mu$ m

**Figure S2.** GFP-Bem3 was expressed from the indicated promoters and copy numbers in W303 WT cells (except RL 3090=endogenous promoter, single copy) and grown overnight at 30°C in the appropriate selective media containing 2% glucose and imaged at 100X using a FITC filter. High levels of Bem3 expression from the *GALI* promoter were achieved by growing cells in 2% galactose containing media for 4 hours prior to imaging. Arrows and arrowheads point to Bem3 at polarized cap and intracellular Bem3-containing compartments, respectively. Scale bar: 5 $\mu$ m

**Figure S3.** Intracellular compartments marked by Yep13-GFP-Bem3 were visualized in W303 WT cells expressing empty vector (EV) control or overexpressing Ypt31<sup>N126I</sup> from a *GALI* promoter. Cells were grown overnight in galactose containing selective media at 30°C and imaged at 100X using a FITC filter. The total area of Bem3-containing compartments is significantly larger in cells overexpressing Ypt31<sup>N126I</sup> compared to EV control. The total area of the Bem3-containing compartment present within a cell was measured using ImageJ (see material and methods) and plotted as a function of bud/mother area ratio. Scale bar: 5 $\mu$ m

**Figure S4.** (A) Wild-type W303 cells expressing GFP-Bem3 were grown at 24°C (permissive temperature) overnight or shifted to 37°C for 6h (restrictive temperature) before imaging at 100X using a FITC filter. Quantification of the Bem3-compartment area was performed using ImageJ. No significant

difference in Bem3-compartment area was observed when wild-type cells were grown at the restrictive temperature.

**(B)** Cells overexpressing GFP-Bem3<sup>K1003A</sup> (mutant unable to bind Cdc42) were imaged at 100X using a FITC filter and DIC.

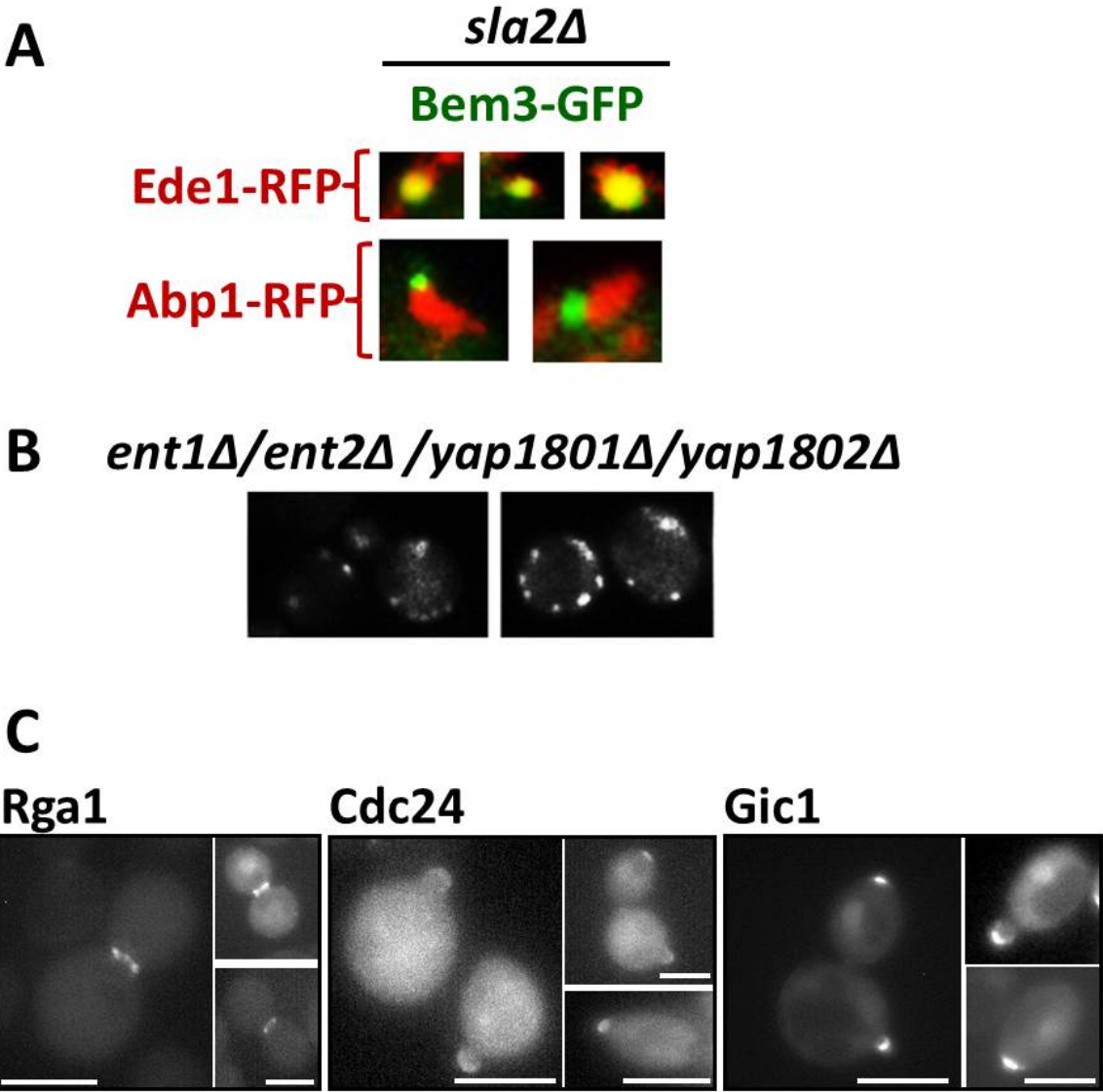
**(C)** Cells expressing Sec4-GFP and transformed with either Bem3 full-length, Bem3 PX-PH fragment or empty vector were imaged at 100X using a FITC filter.

**Figure S5.** (A) Wild-type W303 cells expressing GFP-Sec4 and *Candida albicans* CaBem3 from a *GALI* promoter were grown overnight in media containing 2% glucose at 30°C with shaking at 250 RPM, transferred to 2% galactose containing selective media for 4h and imaged at 100X using a FITC filter. Arrows point to clustered GFP-Sec4. Scale bar: 5µm. (B) *sec4-10* cells expressing GFP-Bem3 were imaged at 100X using a FITC filter.

**Supplementary Movie 1. Dynamics of the Bem3-containing compartment.**

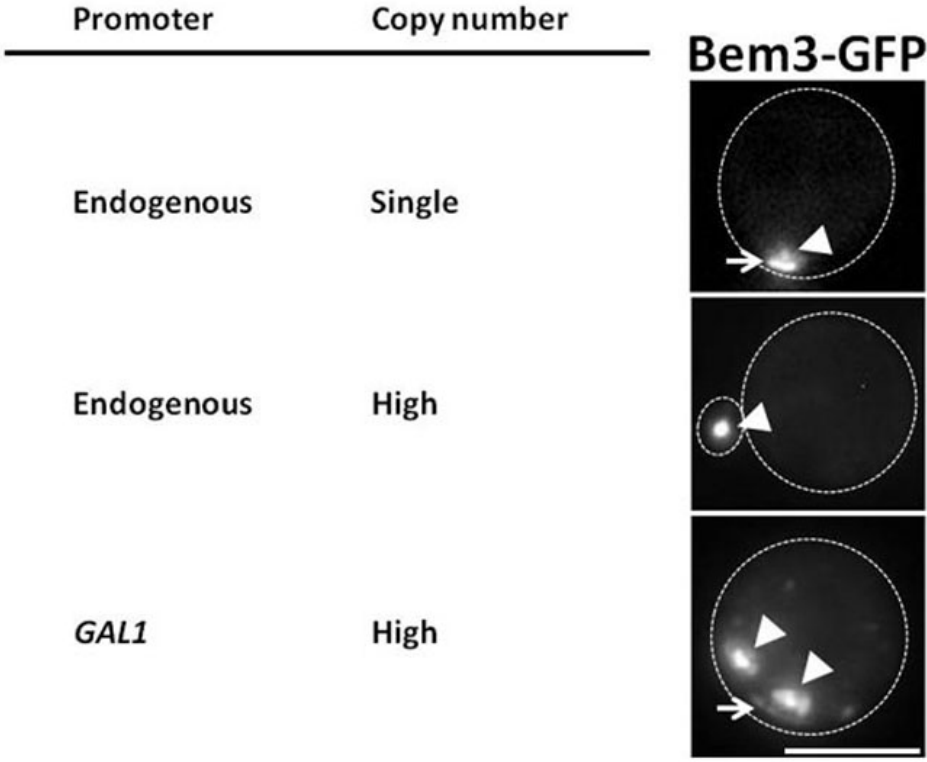
W303 yeast cells transformed with Yep13-GFP-Bem3 (2µ, *LEU2*) were grown overnight in selective media, spotted on media-embedded agarose beds the next morning and imaged with a FITC filter using a 100X objective at 10 second intervals. The cell outline is marked. Movie playback rate: 7 frames/second. Scale bar: 5µm.

Supplementary Figure 1





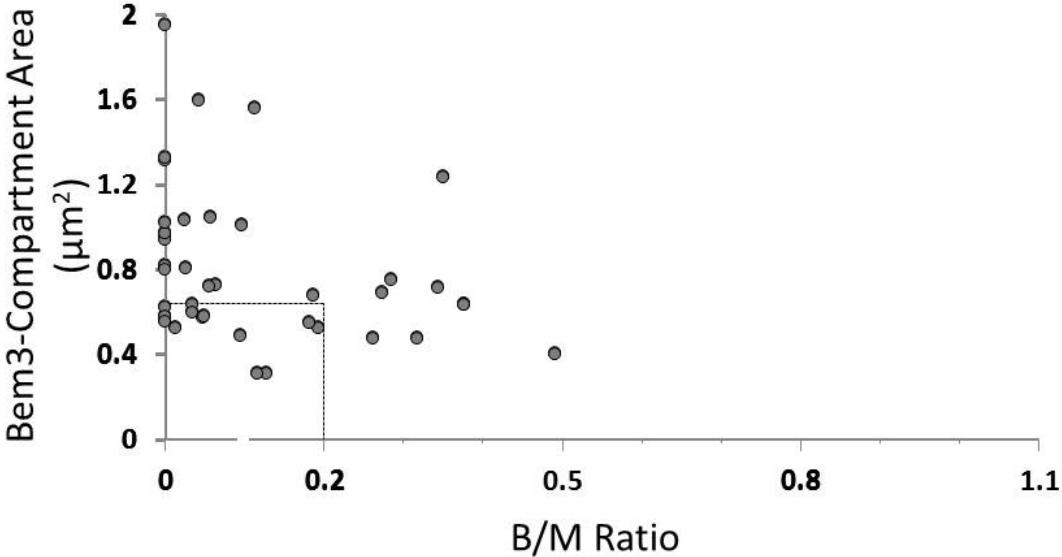
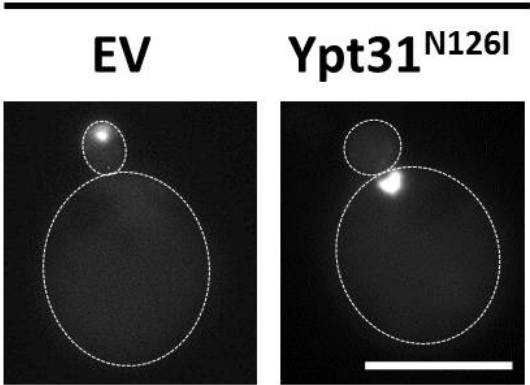
Mukherjee et al., **Supplementary Figure 2**



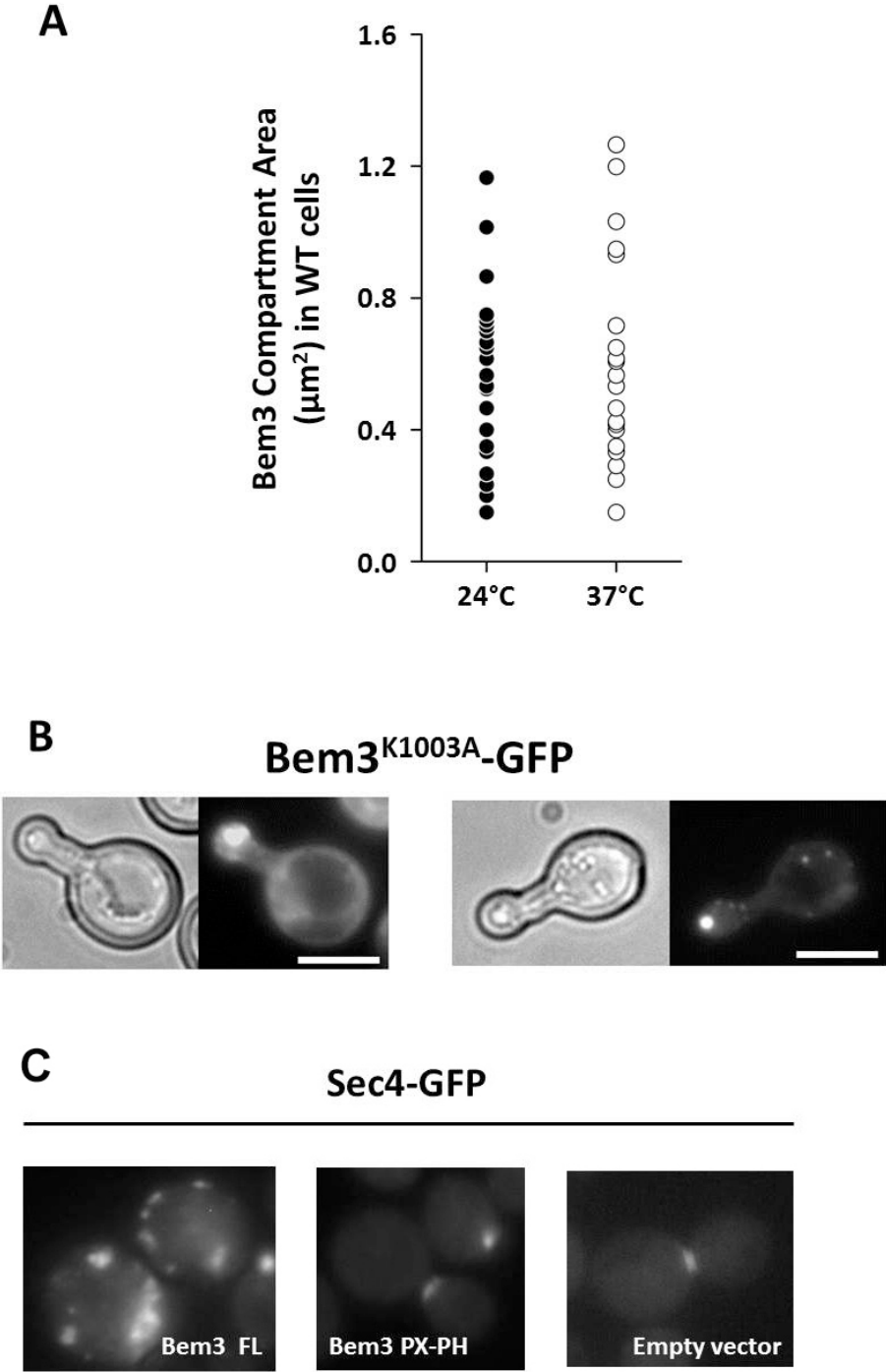
Mukherjee et al.,

### Supplementary Fig. 3

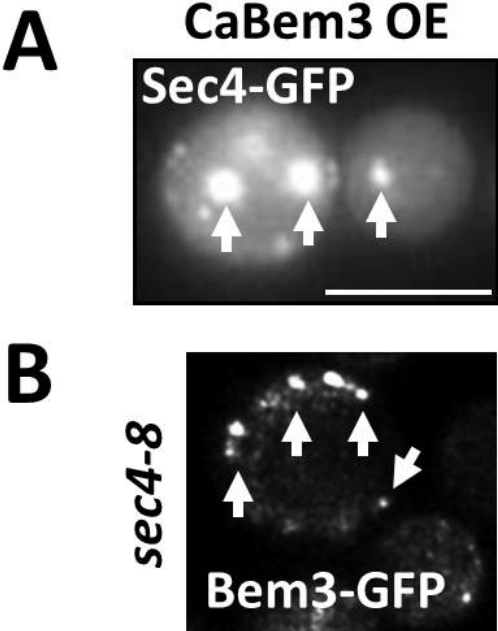
### Bem3-GFP



Supplementary Figure 4



Supplementary Figure 5





**Movie 1. Dynamics of the Bem3-containing compartment.** W303 yeast cells transformed with Yep13-GFP-Bem3 ( $2\mu$ , *LEU2*) were grown overnight in selective media, spotted on media-embedded agarose beds the next morning and imaged with a FITC filter using a  $100\times$  objective at 10 second intervals. The cell outline is marked. Movie playback rate: 7 frames/second. Scale bar: 5  $\mu\text{m}$ .