

**Fig. S1. Control experiments showing the specificity of FZ-3 and NBD-TPEA for Zn and the quenching of NBD-TPEA by Cu. A.-C.** Cells were incubated with the different chelators or CuSO<sub>4</sub> at the indicated concentrations for 3 hrs before the staining with FZ-3 and NBD-TPEA was performed. Afterwards cells were fed with 3 μm and images were taken by live-microscopy. Importantly, the chelators and CuSO<sub>4</sub> were present throughout the experiment.

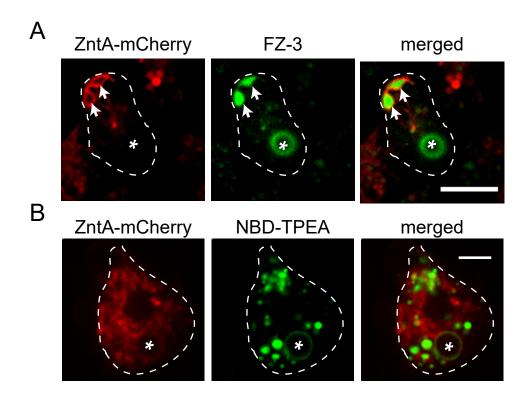


Fig. S2. ZntA-mCherry is not present on BCPs. A. and B. Cells expressing ZntA-mCherry were stained with FZ-3 and NBD-TPEA and fed with 3  $\mu$ m latex beads. Images were taken live. Asterisks label BCPs, arrows point to FZ-3-positive CV-bladders. Scale bars 10  $\mu$ m (A), 5  $\mu$ m (B).

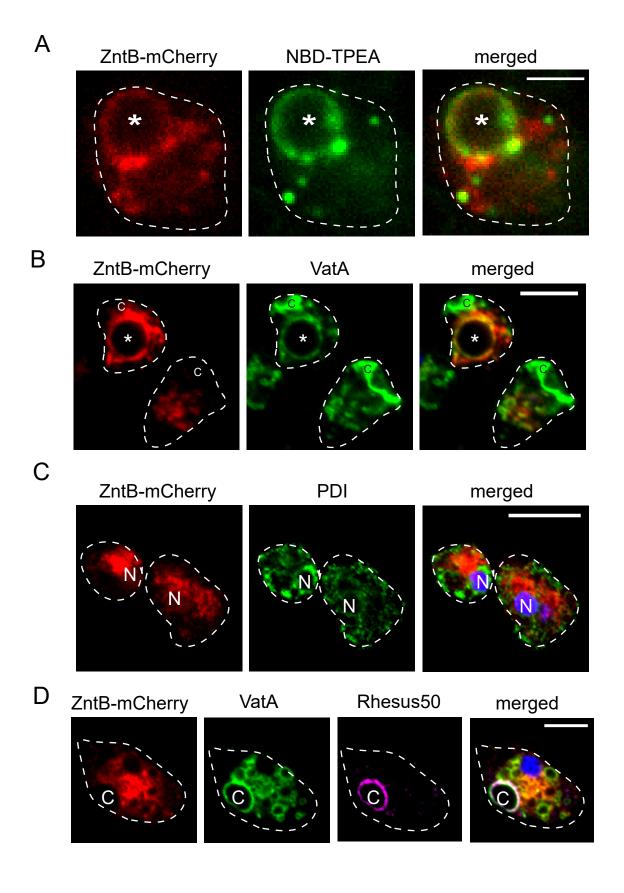
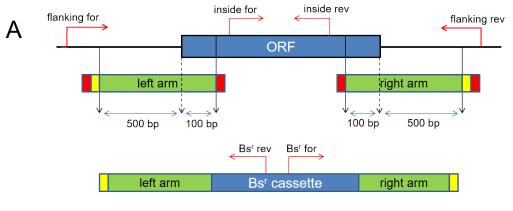


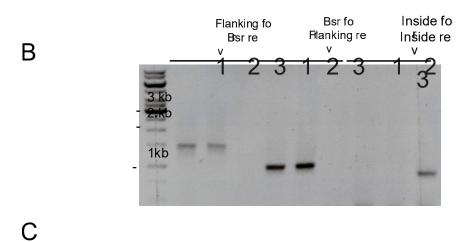
Fig. S3. ZntB-mCherry decorates BCPs and does not co-localize with ER- and CV-markers. A. ZntB is present at BCPs at the lysosomal maturation stage. Cells expressing ZntB-mCherry were stained with NBD-TPEA and fed with 3  $\mu$ m latex beads. Images were taken live. Scale bar, 5  $\mu$ m. B. ZntB-mCherry co-localizes with the vATPase at BCPs. Scale bar, 10  $\mu$ m. C. and D. ZntB-mCherry does not localize at the ER and the at the CV. ZntB-mCherry expressing cells were fixed and stained with antibodies against PDI (C) and Rhesus50 (D). Nuclei were stained with DAPI. Scale bars, 10  $\mu$ m (C) and 5  $\mu$ m (D). Asterisks label a BCPs, C: tubular network of the CV, N: nucleus.

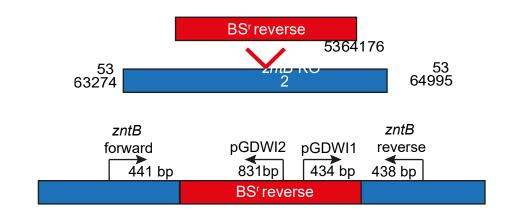


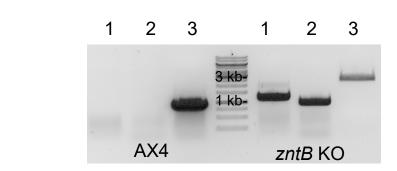
Recombination site and StarCombinase™ recognition area

Unique restriction site

D



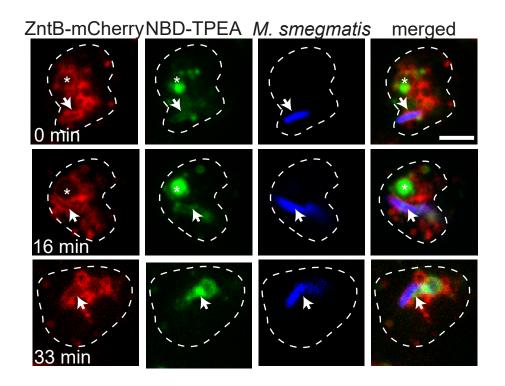




1: zntB forward / pGWDl2 2: pGWDl1 / zntB reverse

3: zntB forward & reverse

Fig. S4. Generation of a *zntA* KO by homologous recombination and localization of the insertion in the *zntB* KO. A. Schematic drawing of the *zntA*-encoding gene locus (ORF, blue) flanked by noncoding segments. For gene disruption, the resistance cassette (BSr, green) was integrated removing a segment in the middle of the gene (between the inside forward/inside reverse primers) using the StarCombinase and the StarGate cloning kit. The red arrows indicate primers that were used to monitor correct integration. B. PCR-analysis of two *zntA* mutants (#1 and #2) and wild type (#3). Using the flanking forward/BSR reverse or the flanking reverse/BSR forward primer combinations small products were obtained in both mutants, but not in the wild type. The inside forward/inside reverse primer combination yielded a small product in the wild type, but not in the mutants. Experiments were performed using mutant #1. C. The restriction-mediated insertion of the *zntB* KO interrupts the gene approximately in the middle at chromosomal position 5364176 (Chromosome 3). D. A diagnostic PCR was performed to confirm the insertion into *zntB* using the primers indicated in C.



**Fig. S5. ZntB-mCherry is localized at the** *M. smegmatis***-containing phagosome.** Cells expressing ZntB-mCherry were stained with NBD-TPEA and fed with *M. smegmatis*. Images were recorded live. Scale bar, 5 μm. Arrows label phagosomes, asterisks point to zincosomes.

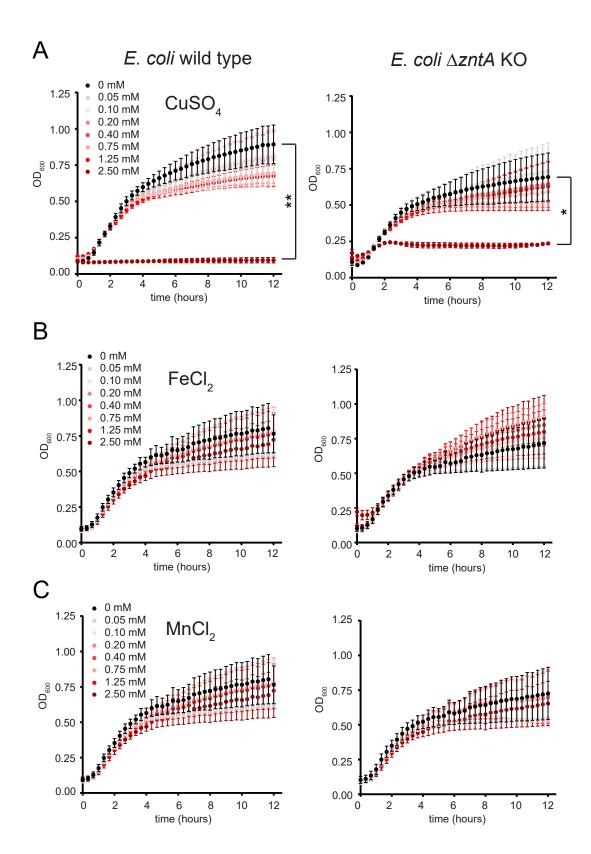
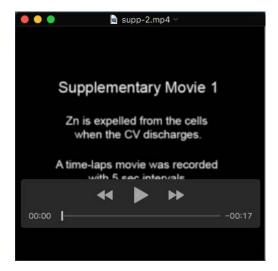


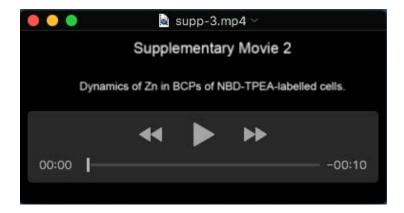
Fig. S6. The *zntA E. coli* KO is not susceptible to increasing concentrations of CuSO<sub>4</sub>, FeCl<sub>2</sub>, MnCl<sub>2</sub>. *E. coli* strains were incubated in LB. Metals were added as indicated. The OD<sub>600</sub> was measured with the help of a 96-well plate reader (SpectraMax i3, Molecular Devices). Statistical differences were calculated with a Bonferroni post hoc test after two-way ANOVA. Significantly different values were indicated by an asterisk (\* P < 0.5, \*\* P < 0.01).

Table S1. Dictyostelium material used for this study.

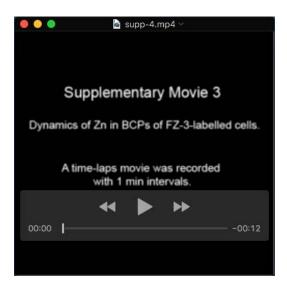
| Dictyostelium strains  | Plasmids used for transformation        | Reference                      |
|------------------------|---|--------------------------------|
| Ax2(Ka) ZntA-mCherry   | ZntA-mCherry                            | This study                     |
|                        | (see below)                             |                                |
| Ax2(Ka) ZntB-mCherry   | ZntB-mCherry                            | This study                     |
|                        | (see below)                             |                                |
| Ax2(Ka) ZntC-mCherry   | ZntC-mCherry                            | This study                     |
|                        | (see below)                             |                                |
| Ax2(Ka) ZntD-mCherry   | ZntD-mCherry                            | This study                     |
|                        | (see below)                             | ·                              |
| Ax2(Ka) zntA KO        | See below                               | This study                     |
| Ax2(Ka) zntA KO        | mCherry-ZntA                            | This study                     |
| mCherry-ZntA           | (see below)                             | ·                              |
| AX4 zntB KO            | ·                                       | REMI library Prof. Christopher |
|                        |   | Thompson                       |
| AX4 zntB KO            | ZntB-mCherry                            | This study                     |
| ZntB-mCherry           | (see below)                             | ·                              |
| Ax2(Ka) AmtA-mCherry   | ,                                       | (Barisch et al., 2015)         |
| Ax2(Ka) VatB-RFP       |   | (Carnell et al., 2011)         |
| Ax2(Ka) RFP-VacA       |   | This study                     |
| AX2 wshA KO            |   | (Carnell et al., 2011)         |
| AX2 pikfyve KO         |   | (Buckley et al., 2018)         |
| Plasmids generated for | Insert                                  | Plasmid &Reference             |
| this work              |   |                                |
| ZntA-mCherry           | zntA gDNA(DDB_G0283629)                 | pDM1044 (Veltman et al., 2009) |
| ZntB-mCherry           | zntB gDNA(DDB_G0282067)                 | pDM1044 (Veltman et al., 2009) |
| ZntC-mCherry           | <i>zntC</i> cDNA( <i>DDB_G0269332</i> ) | pDM1044 (Veltman et al., 2009) |
| ZntD-mCherry           | zntD cDNA(DDB_G0291141)                 | pDM1044 (Veltman et al., 2009) |
| mCherry-ZntA           | zntA amplified from ZntA-mCherry        | pDM1042 (Veltman et al., 2009) |
| ZntA KO plasmid        | pKOSG-IBA-ZntA                          | StarGate® Acceptor Vector      |
|                        | (please see Materials and Methods)      | pKOS-IBA-Dicty1                |
|                        | -                                       | (Wiegand et al., 2011)         |
|                        |   | pDM 324 (Veltman et al., 2009) |



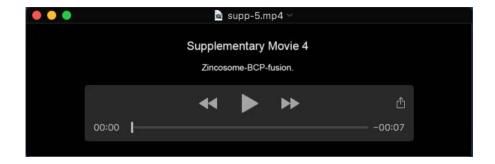
Movie 1. Zn is expelled from the cells when the CV discharges. For more information, see Fig. 1C.



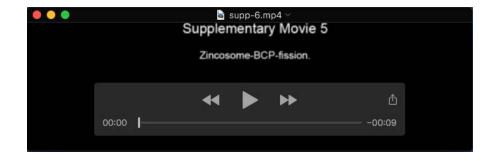
Movie 2. Dynamics of Zn in BCPs of NBD-TPEA-labelled cells. For more information, see Fig. 2A.



Movie 3. Dynamics of Zn in BCPs of FZ-3-labelled cells. For more information, see Fig. 2C.



Movie 4. Zincosome-BCP-fusion. For more information, see Fig. 3A.



Movie 5. Zincosome-BCP-fission. For more information, see Fig. 3C.



Movie 6. Dynamics of ZntB-mCherry at the BCP. For more information, see Fig. 4D.