

Supplementary information

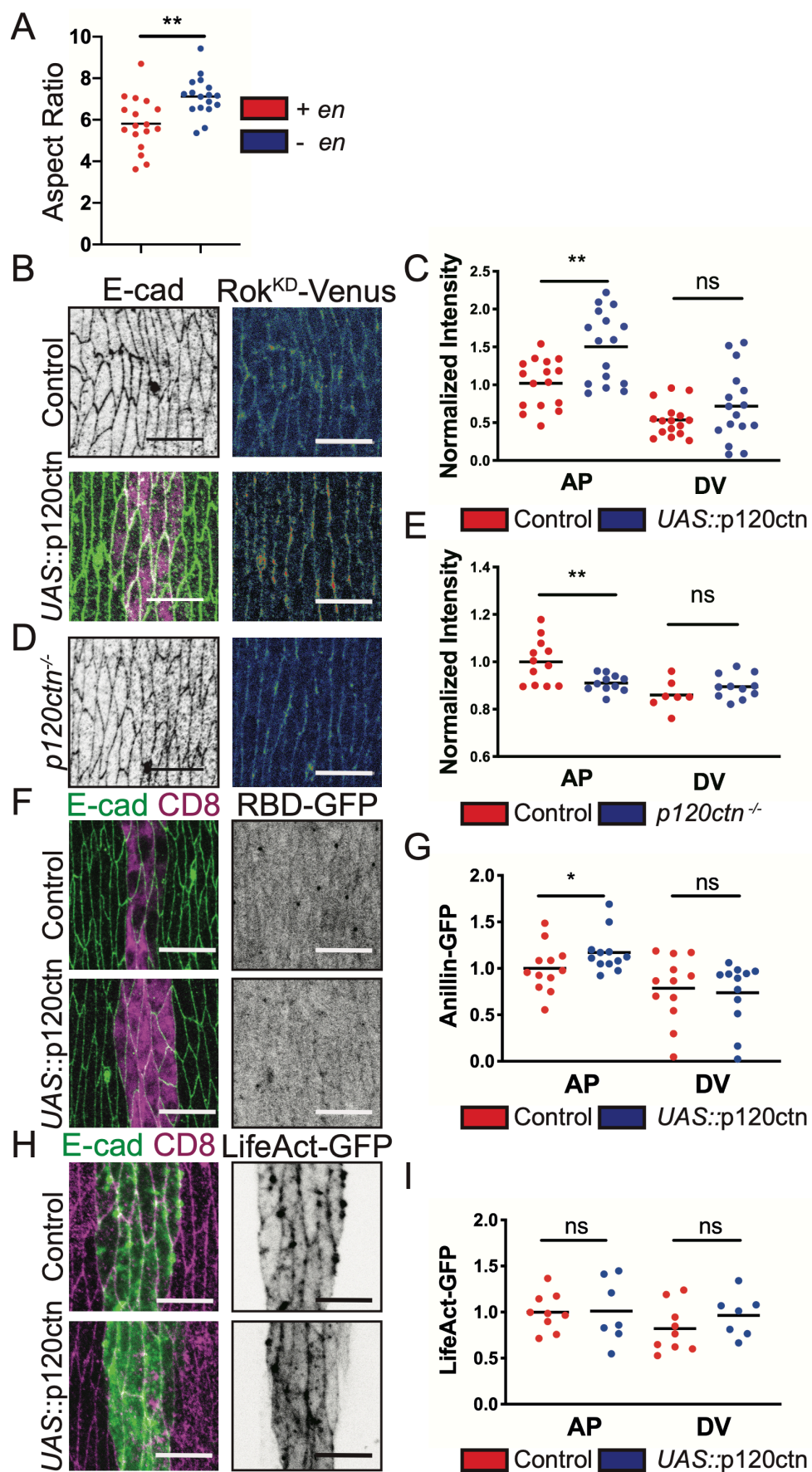


Figure S1. Morphological differences between compartments in embryonic epidermis and effects of p120ctn levels on Rok^{KD}-Venus, RDB-GFP and LifeAct-GFP.

(A) Aspect ratio of cells in the *engrailed*-positive (+*en*) and -negative (-*en*) compartments in embryonic epidermis. (B-E) Levels of Rok^{KD}-Venus following p120ctn level changes. (B) Representative examples (B, D) and levels (C, E) of Rok^{KD}-Venus in cells overexpressing *UAS::p120ctn*-GFP (B-C) or *p120ctn*^{-/-} mutants (D-E). Cell outlines visualized with E-cad antibody (grey or green, left), Rok^{KD}-Venus visualized using rainbow intensity spectrum (right). (F-G) Representative images (F) and levels (G) of RDB-GFP in the epidermal cells of control and *UAS::p120ctn*-GFP expressing embryos. Cell borders were visualized with antibody against E-cad (green in F). (H-I) Representative images (H) and levels (I) of cortical actin visualized using *UAS::LifeAct*-GFP (green, left; grey, right). Cell outlines are visualized with E-cad antibody (magenta, left) Scale bars – 10 μm. Statistical analysis between cell borders measured by two-way ANOVA. *, *p* < 0.05, **, *p* < 0.01. Each dot represents an individual embryo, n number was 10-20 embryos per genotype with a minimum of 9 cells imaged per embryo.

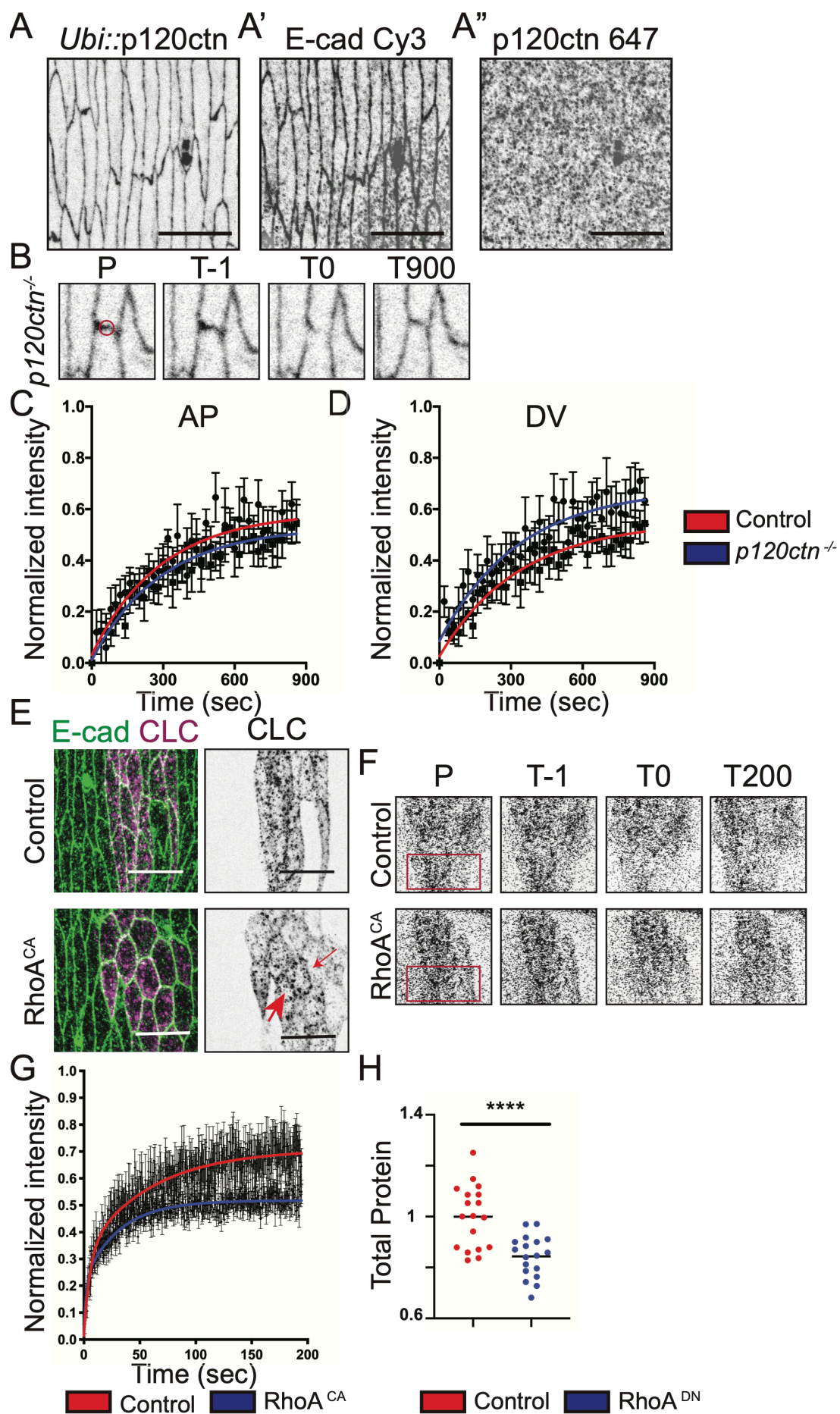


Figure S2. p120ctn antibody staining fails to replicate localisation of GFP tagged variant, while p120ctn loss leads to increase of immobile fraction of endogenously tagged E-cad-GFP and Rho regulatory construct effects the dynamics of CLC.

(A) Apical view of epidermal cells of *ubi::p120ctn-GFP* expressing embryos. The localisation of the p120ctn-GFP (left) compared to the localisation of E-cad stained with an antibody (middle), and the staining pattern of the anti-p120ctn antibody in the same cells (right). (B-D) Representative examples (B) and quantification (C-D) of E-cad-GFP FRAP in *p120ctn^{-/-}* mutant cells. Panels in B show the DV cell border region bleached (Position P, red circle) at the prebleach (Time T-1), bleach (Time T0), and the end (Time T900) time points. Time is in seconds. Average recovery curves (mean \pm s.e.m.) and the best-fit curves (solid lines) are shown. All best-fit and membrane intensity data are in Table S1. (E-G). Localisation (E) and FRAP (F-G) of clathrin (*UAS::CLC-GFP*, grey, right; magenta, left) in cells co-expressing RhoA^{CA} (bottom) and control co-expressing *UAS::CD8-Cherry* (top). Cell borders are visualized by anti-E-cad antibody (green). Panels in F show the region bleached (Position P, red box) at the prebleach (Time T-1), bleach (Time T0), and the end (Time T400) time points. Time is in seconds. Average recovery curves (mean \pm s.e.m.) and the best-fit curves (solid lines) are shown in G. (H) The total levels of E-cad-GFP on the plasma membrane of control and RhoA^{DN} expressing cells. Scale bars – 10 μ m. Statistical analysis was done using a two-tailed students t-test with Welch's correction. ****, $p < 0.0001$. Each dot represents an individual embryo, n number was 10-20 embryos per genotype with a minimum of 26 cells imaged per embryo. For FRAP 8-10 embryos were used.

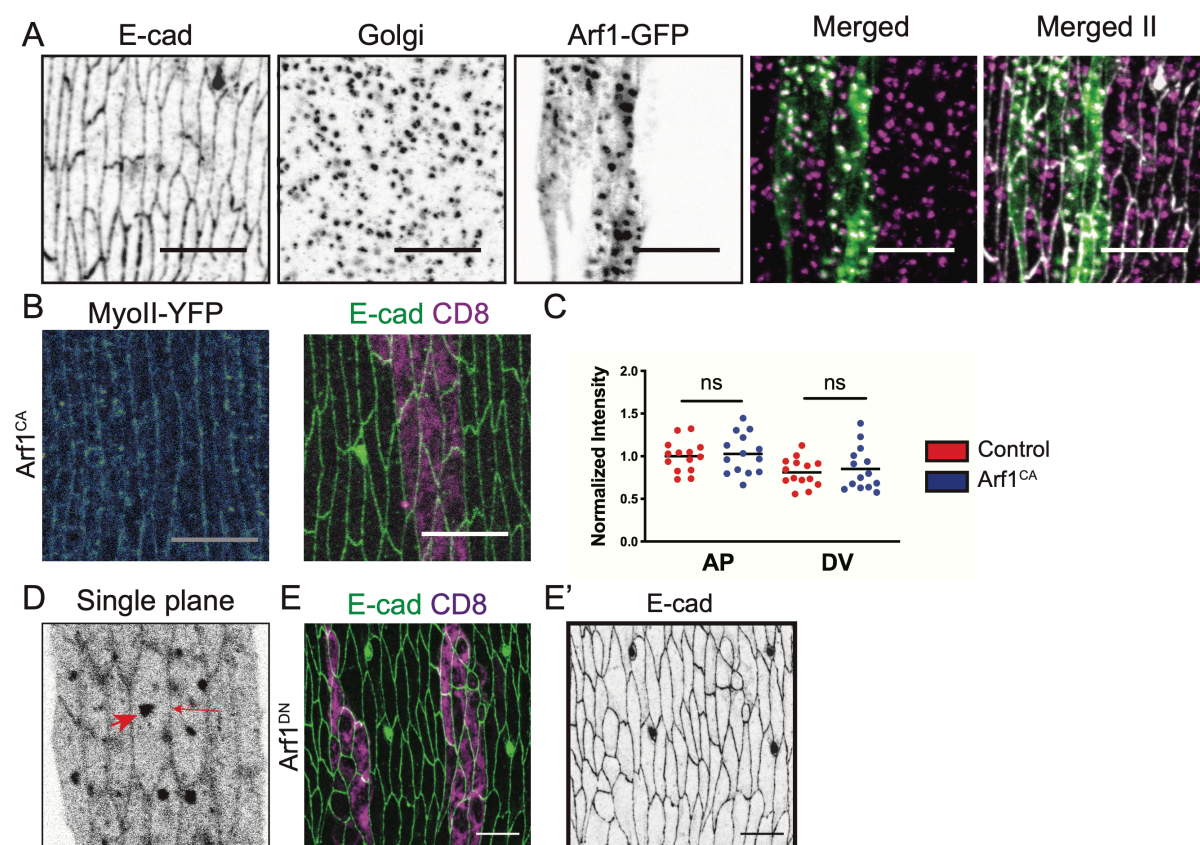
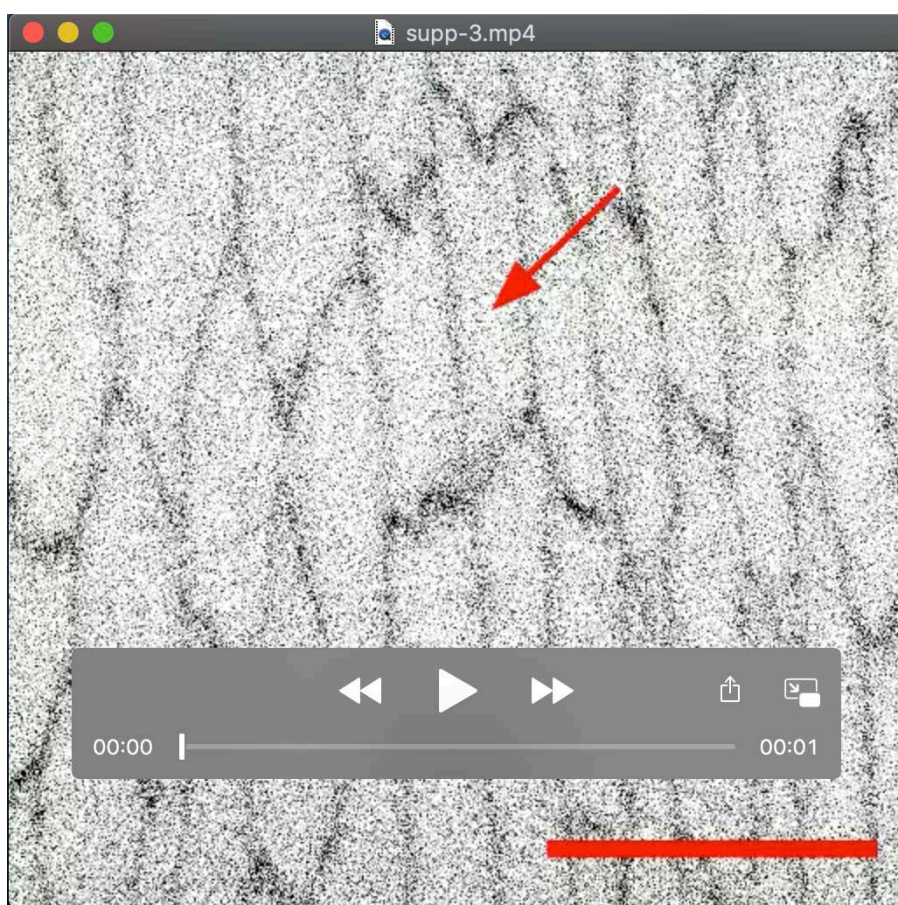


Figure S3. Arf1-GFP localization to the Golgi apparatus and plasma membrane, and effects of Arf^{CA} on MyoII-YFP, and Arf1^{DN} on cell shape.

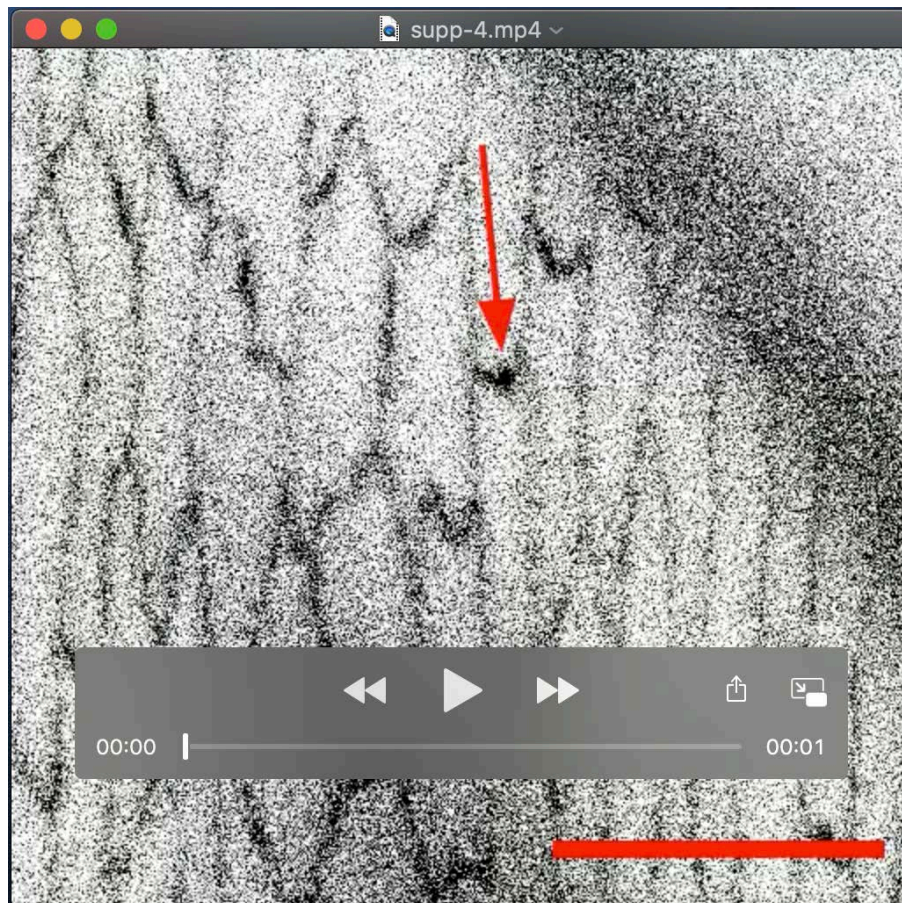
A. Apical view of the dorsolateral epidermis of *UAS::Arf1-GFP* expressing control embryo with cells borders are marked by antibody staining for E-cad (black, left; white, right), Golgi marked by immunostaining of Trans-Golgi (black, second left; magenta, two right-most), and Arf1-GFP localisation in the engrailed expressing cells (black, middle; green, two right-most). The same region is shown in main text (see Fig. 5). **(B-C)** Representative images **(B)** and levels **(C)** of MyoII-YFP (rainbow, left) in the cells expressing a constitutively active Arf1 (Arf1^{CA}). Cell borders are visualized with anti-E-cad antibody (green, right). **(D)** Single plane image of Arf1-GFP in the middle of the AJ. Large arrow indicates Golgi population and small arrow indicates plasma membrane resident population. **(I)** Low magnification image of epidermis expressing Arf1^{DN}. Scale bar is 10 μ m. Statistical analysis between cell borders measured by two-way ANOVA. Each dot represents an individual embryo, n number was 10-20 embryos per genotype with a minimum of 17 cells imaged per embryo.

Table S1. Numerical values for each experiment presented in paper.

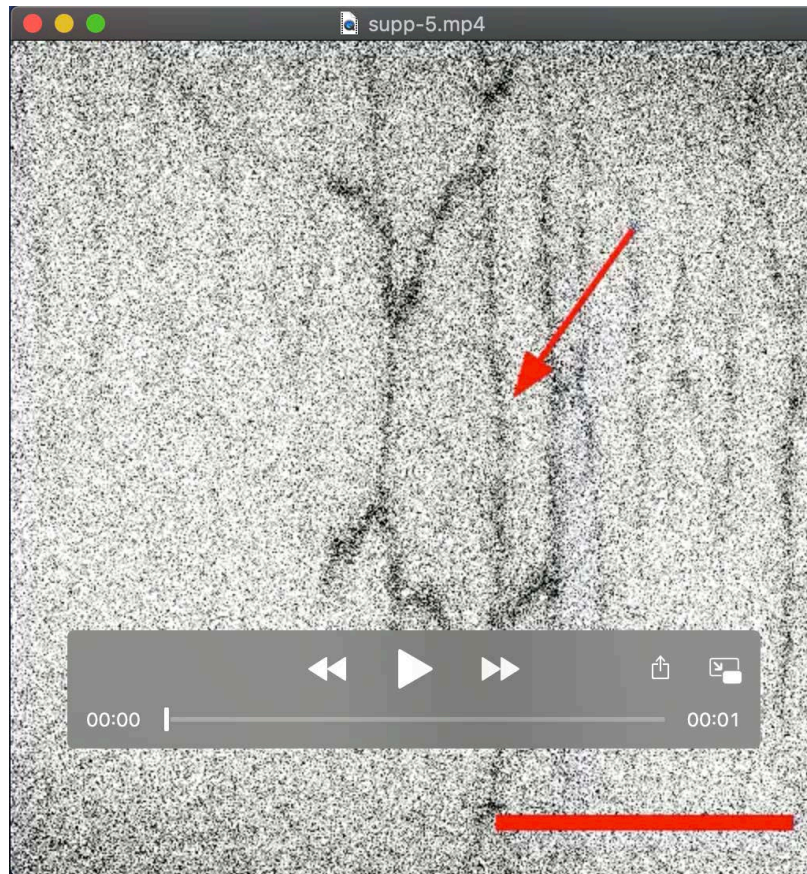
[Click here to Download Table S1](#)



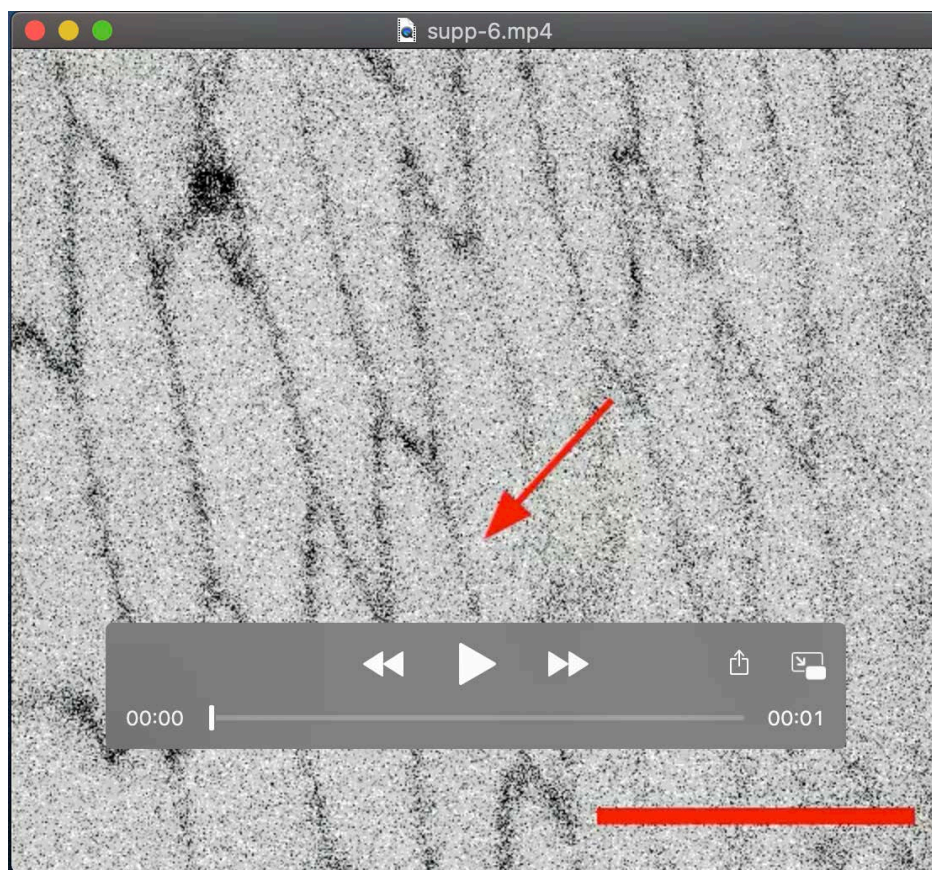
Movie 1. Laser ablation of AP cell border in control embryos. Arrow indicates the ablated junction. Scale bar – 10 μm .



Movie 2. Laser ablation of DV cell border in control embryos. . Arrow indicates the ablated junction. Scale bar – 10 μm .



Movie 3. Laser ablation of AP cell border in *UAS::p120ctn* expressing embryos. . Arrow indicates the ablated junction. Scale bar – 10 μm .



Movie 4. Laser ablation of AP cell border in *p120ctn*^{-/-} mutant embryos. . Arrow indicates the ablated junction. Scale bar – 10 μ m.