Table S1 – Actin-profilin binding free energies computed from molecular dynamics simulations for the wild-type and mutant systems.

		Binding Free		Binding Free Energy
		Energy		
Pb Pfn	GB	-46±13	Pb Pfn	-46±13 (n. d.)
	PB	-48±19		-48±19 (n. d.)
Pf Pfn	GB	-35±10	Pf Pfn	-35±10 (-62±9)
	PB	-35±13		-35±13 (-64±11)
Pb Pfn <sup>Pfloop</sup>	GB	-15±11	Pf Pfn AAA	-17±7 (-34±11)
	PB	44±35		-26±10 (-46±8)
$Pf Pfn^{Pbloop}$	GB	-61±10	Pf Pfn QNQ	-54±14 (-44±13)
	PB	-61±14		-69±14 (-51±11)
$Pf Pfn^{Tgloop}$	GB	-56±14		
	PB	-51±22		

Average binding free energies (kcal/mol) and their standard deviations were calculated from the last 200 ns of the 500 ns long simulation using the MM-GBSA (GB) and MM-PBSA (PB) methods. Note that the (energetically unfavorable) translational, rotational and vibrational entropic contributions to binding were not computed. The values for the wild-type profilins are given followed by values for (left) acidic loop mutants and (right) arm motif mutants. The values obtained from the last 50 ns of the 150 ns long simulations previously reported in (Moreau et al., 2017) are given in brackets for comparison. Blue numbers denote values smaller in magnitude than those for the *P. berghei* wild type profiling, indicating weaker binding. Red numbers denote the weakest binding affinities.

Table S2

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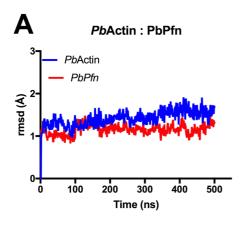
Table S3

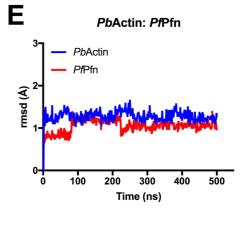
Click here to Download Table S3

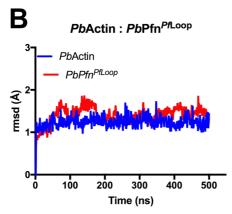
Table S4: Primers used in this study

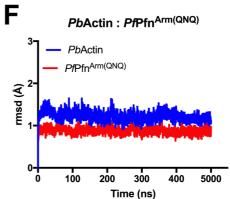
Primer	Sequence	Restriction site	Binding site
			Pb Pfn 5' UTR, upstream
1 fw	GGTGCACACTCATTTGAATGTG	-	of homologous recombination site
1a rv	GCtctagaTTATGCGGCACCTGTATCAG	XbaI	Pb Pfn end of CDS
1b rv	GCtctagaTTACTGTGAGCTTTCTGCCAG	XbaI	Pf Pfn end of CDS
2 fw	CTAGACAGCCATCTCCATCTGG	-	Tg DHFR end of CDS
			Pb Pfn 3'UTR, downstream
2 rv	CAAGTTCTTTCCTCATGTGTTCATG	-	of homologous recombination site
3 fw	ATTTgcggccgcATGGAAGAATATTCATGGG	NotI	Pb Pfn start of CDS
3 rv	GCtctagaTTATGCGGCACCTGTATCAG	XbaI	Pb Pfn end of CDS
4 fw	ATTTgcggccgcAAAATGGCAGAGGAGTATTCTTGG	NotI	Pf Pfn start of CDS
4 rv	GCtctagaTTACTGTGAGCTTTCTGCCAG	XbaI	Pf Pfn end of CDS
5 fw	TCCccgcggGAGATATTACACATTGCTAC	SacII	Pb Pfn 5' UTR
5 rv	TAAAgcggccgcCTTTATTATCTTAAAAAATTATTTATATAATATGATG	NotI	Pb Pfn 5' UTR
6 fw	CCatcgatAATAAAGAAAATATTATAAAAAATGTG	ClaI	Pb Pfn 3' UTR
6 rv	GGggtaccCACACATTGGCATTATATAGAAATTGAG	KpnI	Pb Pfn 3' UTR
	CAAAATTTGGGTCACTCTTCACCCTG	•	
7a	TGCTACGCATTCATATAC	-	Pb Pfn with Pf loop A rv
	CACAGGGTGAAGAGAGTGACCCA		
7b	AATTTTGATAAATGGTCTCTTTTTTATAAAGAAGA	-	<i>Pb</i> Pfn with <i>Pf</i> loop B fw
8a	TCAAAGTCTGGATTATTTTCATCGGGGGTAGCTACACAAGCATAGA	-	Pf Pfn with Pb loop A rv
	AGCTACCCCGATGAAAATAATCCAGAC		
8b	TTTGATAAATGGTCACTTTTTTATAAAGAAGATT	-	<i>Pf</i> Pfn with <i>Pb</i> loop B fw
	CAGCTTGGACCATCCGTCATCATCATCAGCCGCCGCGGCGAA		
9a	GACAACTCCATCTTCTTCCGAAGCTAATC	-	Pf Pfn with $Tg$ loop A rv
	TTCGCCGCGGCGGCTGATGATGATGACGGA		
9b	TGGTCCAAGCTGTATAAAGAAGATTATGATAAGTTGAAGATGAAAATGGTAC	_	Pf Pfn with $Tg$ loop B fw
70	1661CC/MGCTGTAT/AAAAAAATTATTAAAATTGAAAAATGGTAC		1 J I III willi 1 g 100p D I w

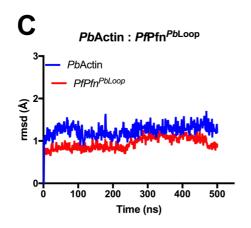
restriction sites in lower case, start codons indicated in **green**, stop codons indicated in **red**, *Pf* Pfn loop in **blue**, *Pb* Pfn loop in **orange** and *Tg* Pfn loop in **purple** 

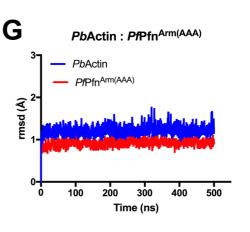












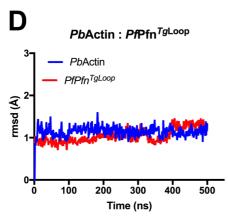


Figure S1: RMSDs of the backbone atoms of the different systems, showing that the individual protein structures are stable during the MD simulations. Plots of RMSD for each protein in the complex of *Pb* Actin with (A) *Pb* Pfn, (B) *Pb* Pfn <sup>PfLoop</sup>, (C) *Pf* Pfn <sup>PbLoop</sup>, (D) *Pf* Pfn <sup>TgLoop</sup>, (E) *Pf* Pfn, (F) *Pf*Pfn<sup>Arm(QNQ)</sup> and (G) *Pf*Pfn<sup>Arm(AAA)</sup> for 500 ns of simulation. Each trajectory is aligned to its respective first frame. The flexible regions, *i.e.* the D2 loop region in actin and the terminal and arm regions in profilin (M1-L10, E42-F84, T169-A174), were excluded while calculating RMSD values.

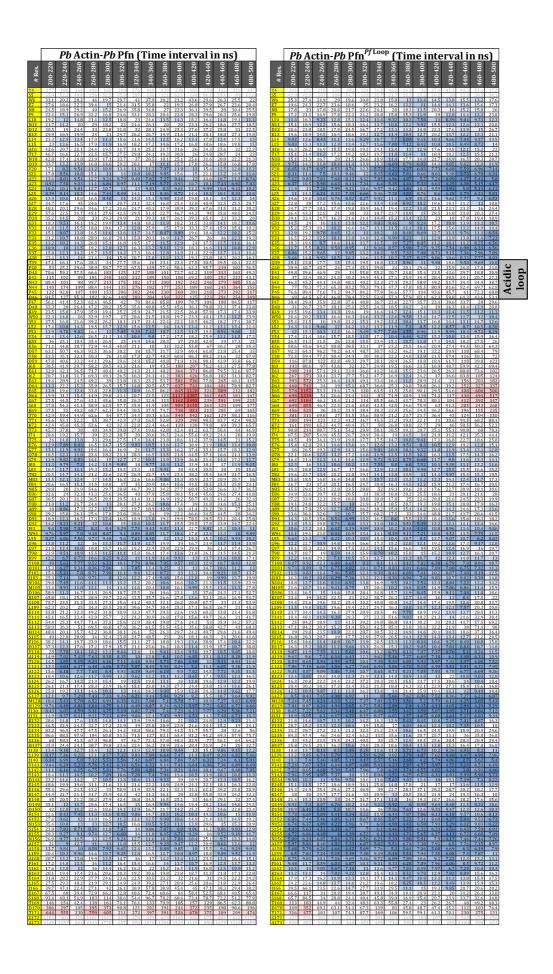


Figure S2: Atomic fluctuations (B-factor) of Pb Pfn (left) or Pb Pfn PfLoop (right) during MD simulations (each complexed with PbActin). Residue numbers are highlighted in yellow at the left corner of each panel. Each column represents B-factor values (units are  $Å^2x(8/3)\pi^2$ ) for the  $C\alpha$  atoms for the interval of 20 ns during 200-500 ns of simulations. The acidic loop residues are bordered with bold lines and are highlighted with grey text (right corner). Color code: blue, white, red: low, intermediate and high B-factor values, respectively. Please note that this data is also available as separate Excel file.

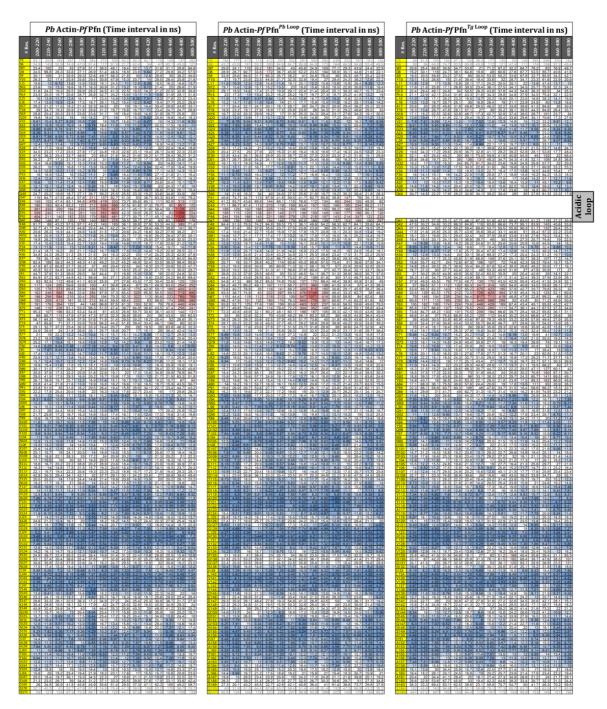
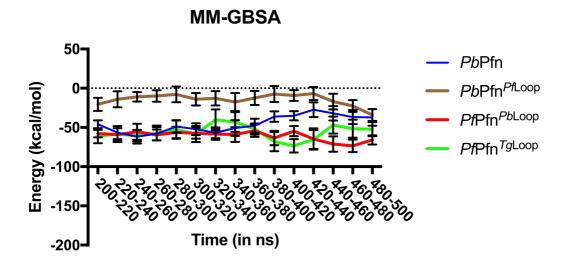


Figure S3: Atomic fluctuations (B-factor) of Pf Pfn (left), Pf Pfn  $_{PbLoop}$  (middle) and Pf Pfn  $_{TgLoop}$  (right) during MD simulations (each complexed with PbActin).

Residue numbers are highlighted in yellow at the left corner of each panel. Each column represents B-factor values (units are  $Å^2x(8/3)\pi^2$ ) for the  $C\alpha$  atoms for the interval of 20 ns during 200-500 ns of simulations. The acidic loop residues are bordered with bold lines and are highlighted with grey text (right corner). Blue, white, red: low, intermediate and high B-factor values, respectively. Please note that this data is also available as separate Excel file.



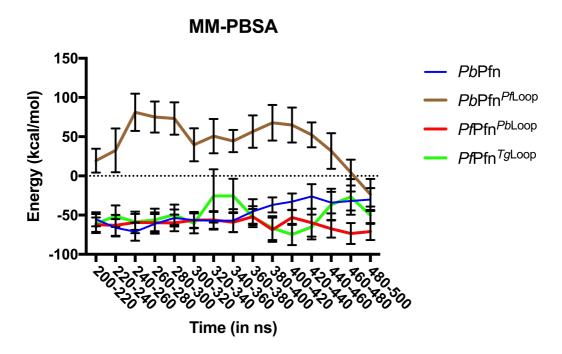
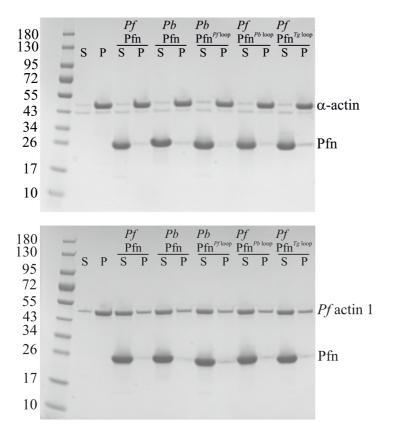
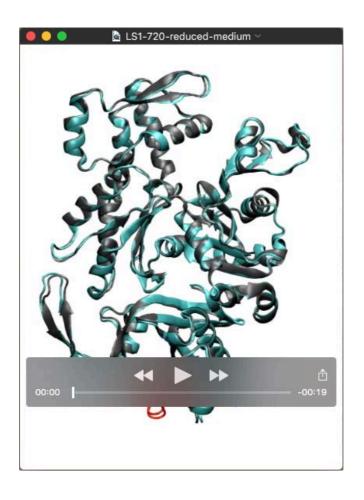


Figure S4: MM-PBSA and MM-GBSA estimated binding free energy of PbActin complexed with different profilin mutants. Average  $\pm$  standard deviation binding free energies (kcal/mol) were calculated for intervals of 20 ns during 200-500 ns of simulations using the MM-GBSA and MM-PBSA methods. Note that the (energetically unfavorable) translational, rotational and vibrational entropic contributions to binding were not computed.



**Figure S5: Co-sedimentation gels.** Sedimentation of 4 μM *S. scrofa* (domestic pig) α-actin (top gel) or *P. falciparum* actin 1 (bottom gel) alone and in the presence of 16 μM *P. falciparum* and *P. berghei* profiling as well as profiling chimeras (*Pb* Pfn<sup>PfLoop</sup>, Pf Pfn $^{Pb}$ Loop and Pf Pfn $^{Tg}$ Loop). Samples were analyzed on 4- 20% SDS-PAGE gels and protein bands were visualized with PageBlue stain (Thermo Scientific). S denotes supernatant and P pellet. Quantification from duplicate gels is presented in Fig. 3B.

## **Supplementary movies**



## Movie 1

500 ns simulation of the chimeric mutant *Pb* Pfn<sup>Pf</sup> Loop in grey compared to *Plasmodium berghei* profilin-WT (cyan) using all atom molecular dynamics simulation. The profilin mutant (or WT) is bound to *Plasmodium berghei* actin. For comparison, see movies in Moreau et al., 2017 (ref 5), for 150 ns simulations for mutants of the profilin arm.



## Movie 2

500 ns simulation of the chimeric mutant Pf Pfn<sup>Pb</sup> Loop in grey compared to  $Plasmodium\ berghei$  profilin-WT (cyan) using all atom molecular dynamics simulation. The profilin mutant (or WT) is bound to  $Plasmodium\ berghei$  actin.



## Movie 3

500 ns simulation of the chimeric mutant Pf  $Pfn^{Tg}$  Loop in grey compared to  $Plasmodium\ berghei$  profilin-WT (cyan) using all atom molecular dynamics simulation. The profilin mutant (or WT) is bound to  $Plasmodium\ berghei$  actin.