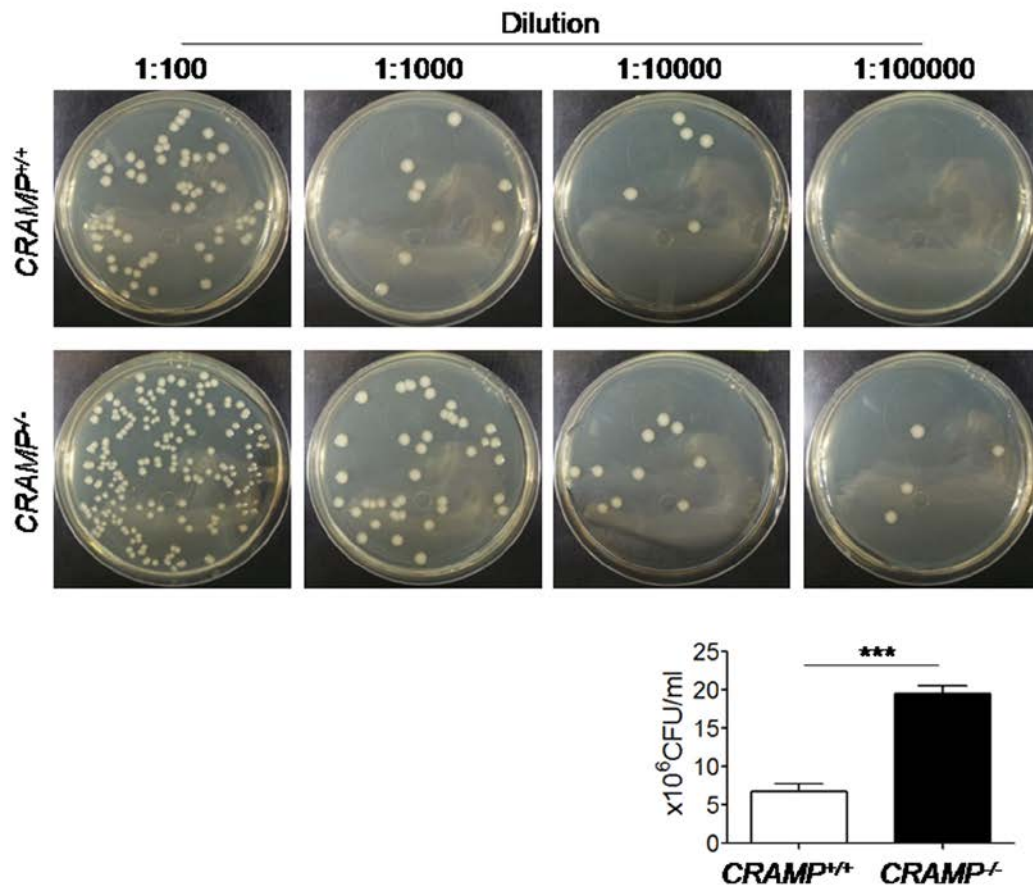
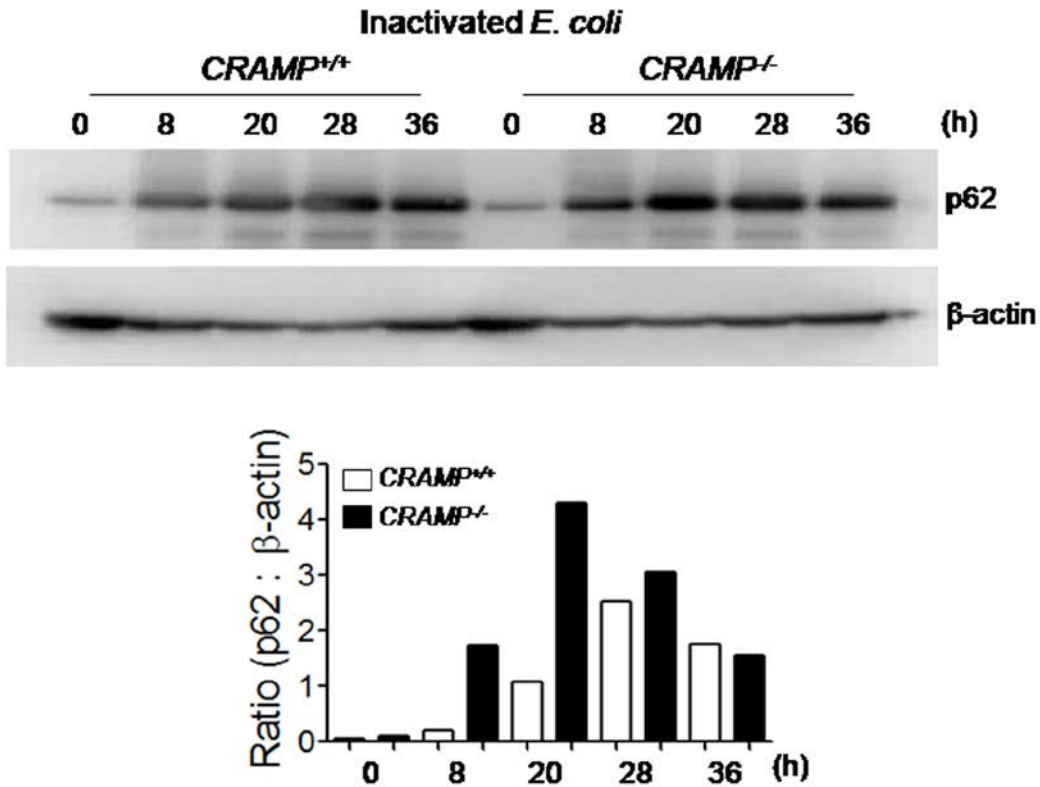


**Fig. S1. Reduced expression of CRAMP in macrophages derived from *Myeloid CRAMP*<sup>-/-</sup> mice.**

*CRAMP*<sup>+/+</sup> and *CRAMP*<sup>-/-</sup> macrophages were seeded in 35 mm dishes at  $1 \times 10^6$ /dish with 14 mm coverslips. The cells were fixed with 4% neutrally buffered formalin for 5 min and stained with an anti-mouse CRAMP antibody followed by a biotinylated anti-Ig secondary antibody (BD Biosciences) and streptavidin-PE. DAPI was used to stain nuclei. **Red:** CRAMP, **blue:** DAPI. Scale bar = 10  $\mu$ m. **Lower panel:** Quantitation of CRAMP positive staining spots per macrophage. Shown is the immunofluorescence intensity per macrophage, n = 20 macrophages/group. \*\*\* $P < 0.001$ .



**Fig. S2. Attenuated killing of intracellular *E. coli* in *CRAMP*<sup>-/-</sup> macrophages.** *CRAMP*<sup>+/+</sup> or *CRAMP*<sup>-/-</sup> macrophages were co-cultured with *E. coli* at MOI= 100 in the snap-cap polypropylene tubes in a shaker at 80 RPM, 37 °C for 1 h. The cells were then treated with gentamicin (50 µg/ml) for 1 h, washed and resuspended in DMEM with 10% FCS in the presence of 5% gentamicin and 50 ng/ml M-CSF followed by further incubation at 37°C, 5% CO<sub>2</sub> for 20 h. Five 1/10 serial dilutions were made in sterile water, which were incubated in triplicates on LB agar at 37°C for 24 to 48 h. **Upper panel:** Representative photos showing the colonies of *E. coli* O22H8 at different dilutions. **Lower panel:** Quantitation of the colonies formed by *E. coli* O22H8 at 1:100 dilution on LB agar. \*\*\**P* < 0.001.



**Fig. S3. Delayed degradation of p62 in *CRAMP*<sup>-/-</sup> macrophages.**

BM-derived *CRAMP*<sup>+/+</sup> control and *CRAMP*<sup>-/-</sup> macrophages grown in 60-mm dishes to sub-confluency were cultured for 3 h in FCS-free media. The cells were incubated with inactivated *E. coli* (MOI= 10) and harvested at indicated time points followed by lysis. The lysates with titrated proteins were analyzed by Western blotting. For detection of p62, the membranes were incubated with anti-mouse p62 Ab.  $\beta$ -actin was used as a loading control. **Lower panels:** Ratio of p62:  $\beta$ -actin.

**Table S1. Antibodies and reagents.**

| REAGENT or RESOURCE                  |                    |                   |
|--------------------------------------|--------------------|-------------------|
| <b>Antibodies</b>                    | <b>Source</b>      | <b>Identifier</b> |
| Ms mAb to CRAMP                      | Santa Cruz, TX     | Cat# SC-166055    |
| Rb mAb to p62                        | Abcam, MA          | Cat# ab240635     |
| Rb pAb to LC3B                       | Abcam, MA          | Cat# ab48394      |
| Rb mAb to ATG5                       | Abcam, MA          | Cat# ab108327     |
| Anti P-p38 Ab                        | Cell signaling, MA | Cat# 9211         |
| Anti p38 Ab                          | Cell signaling, MA | Cat# 9212         |
| Anti P-ERK1/2 Ab                     | Cell signaling, MA | Cat# 9101         |
| Anti ERK1/2 Ab                       | Cell signaling, MA | Cat# 9102         |
| Anti P-I $\kappa$ B- $\alpha$ Ab     | Cell signaling, MA | Cat# 9241         |
| Anti I $\kappa$ B- $\alpha$ Ab       | Cell signaling, MA | Cat# 9242         |
| <b>Other reagents</b>                |                    |                   |
| Mouse CRAMP                          | Hycult Biotech. PA | Cat# HC1106       |
| Elastatinal                          | Abcam, MA          | Cat# 144541       |
| E64d                                 | Abcam, MA          | Cat# 144048       |
| Pepstatin A                          | Abcam, MA          | Cat# 141416       |
| BAY11-7082                           | Abcam, MA          | Cat# 141228       |
| Live/Dead Viability/Cytotoxicity Kit | Them Fisher, MA    | Cat# L3224        |