

Fig. S1. Smad7-myc, TAZ-flag, β-catenin alone or in combination were ectopically expressed along with the FOP flash firefly luciferase reporter gene. *Renilla* luciferase served as the transfection control. Cells were harvested for dual Luciferase determination at 16 h post transfection. Normalized luciferase activity was compared to the control to determine fold changes. Lower panels indicate western blot analysis of the protein levels derived from the transfected plasmids. Each condition was compared to the control for the three individually transfected samples to determine fold changes. Each bar represents the mean of three technical replicates. The error bars represent standard error of the mean (SEM).

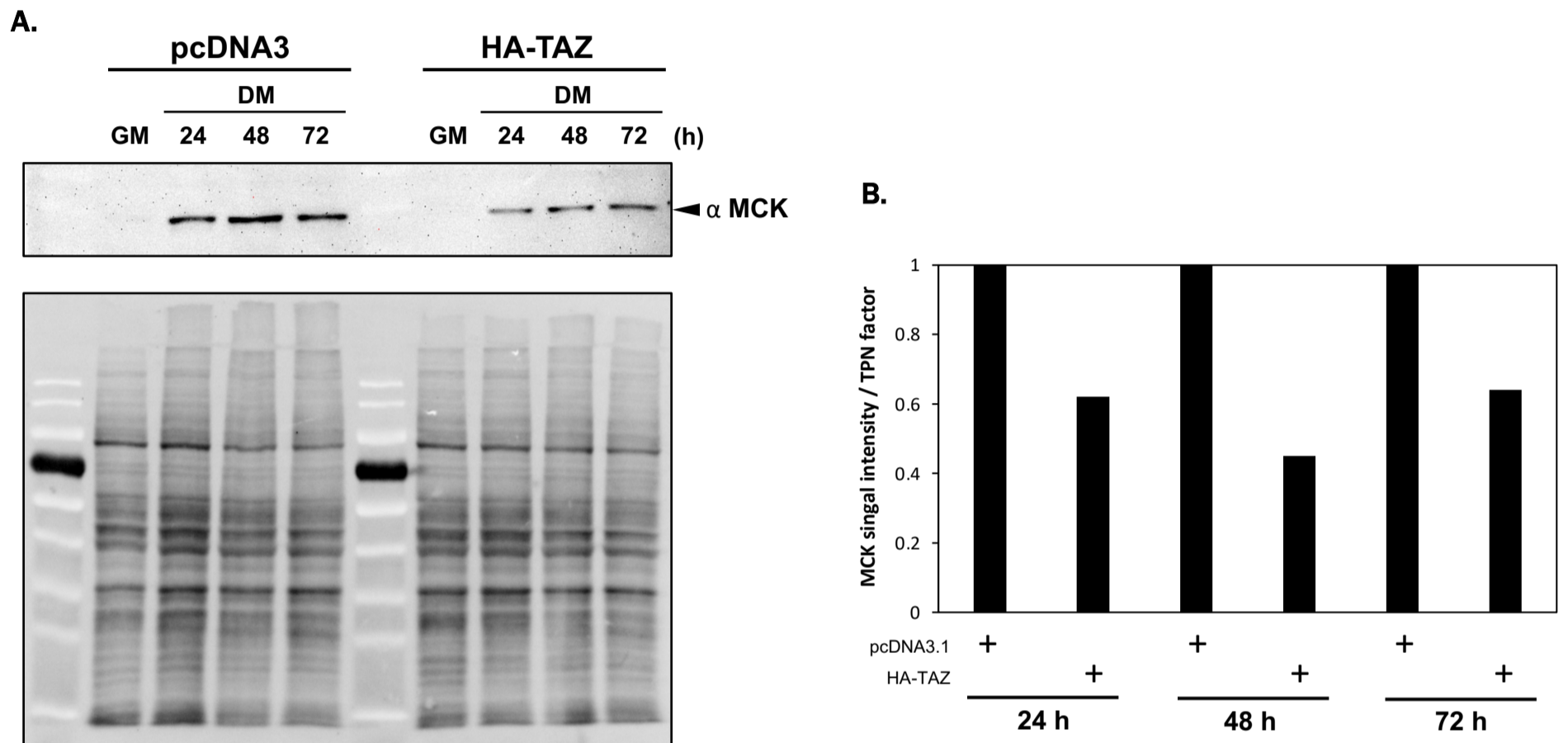


Fig. S2. HA-TAZ and pcDNA3 control were ectopically expressed in C2C12 and grown for 24 hours before switching to differentiation media for 24 to 72 h. Lysates were collected and assessed for expression of MCK by western blot analysis. MCK protein levels were quantified at each timepoint and normalized to the signal band intensity respective to total protein normalization (TPN) factor indicated in the gel image below the western blot. On the right side, a bar graph representing the TPN normalized MCK data at each timepoint is shown, indicating a downregulation of MCK protein levels in differentiating C2C12 cells transfected with TAZ compared to the corresponding control.

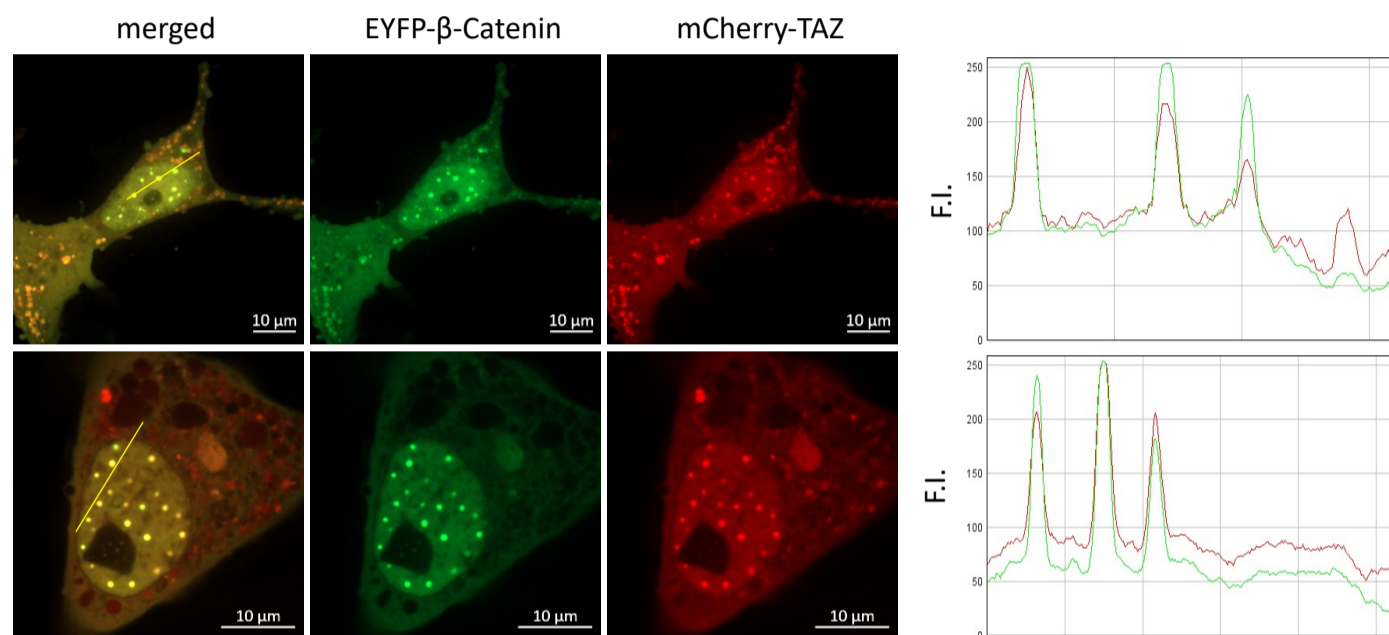


Fig. S3. C2C12 cells were transfected with indicated expression constructs. After 1 day, the cells were subjected to live-cell confocal fluorescence microscopy. The micrographs of samples were analyzed for co-localization by line-scan analysis for fluorescence intensity (F.I.) of green (EYFP-β-catenin) and red (mCherry-TAZ) using ImageJ.

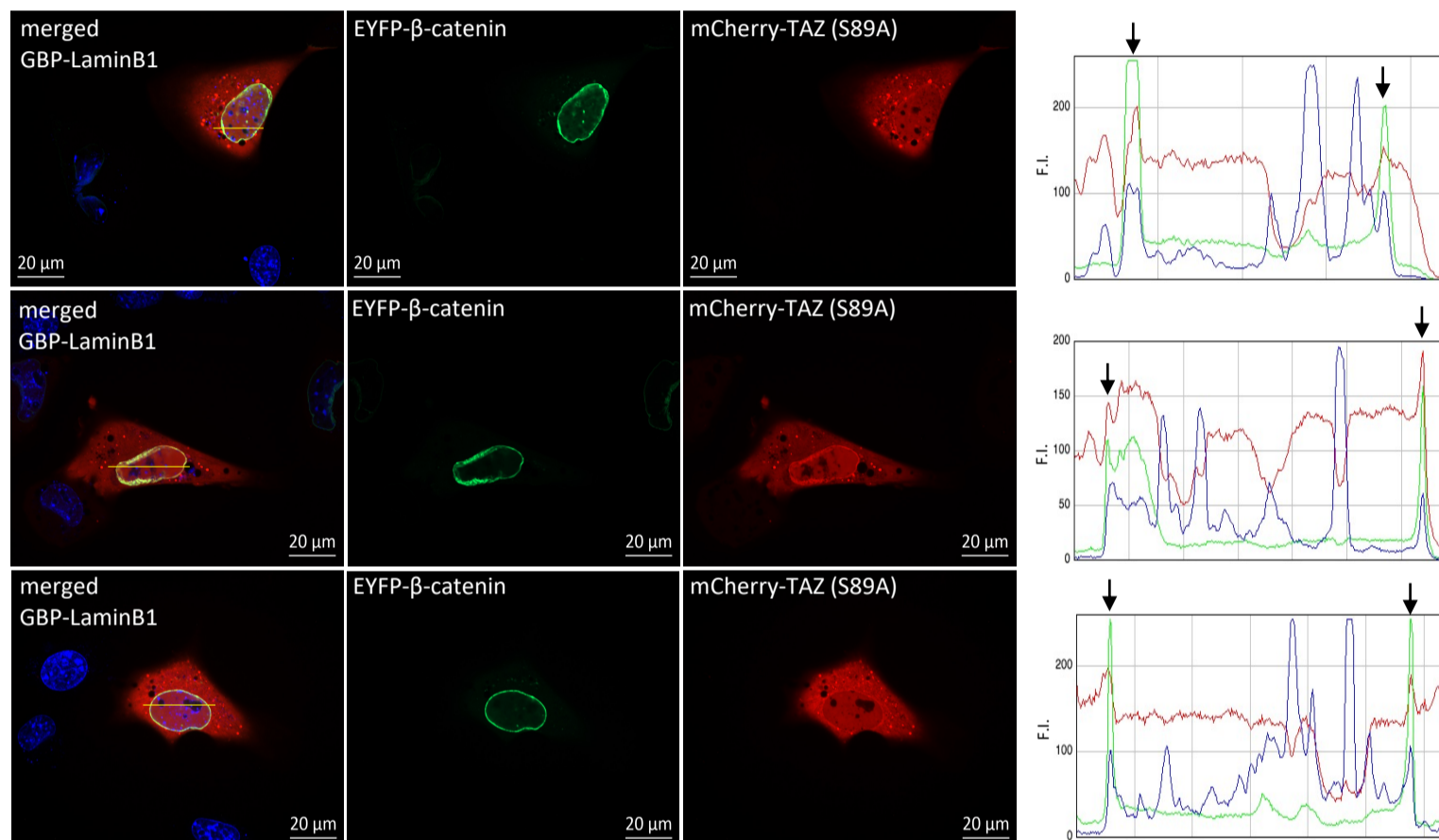
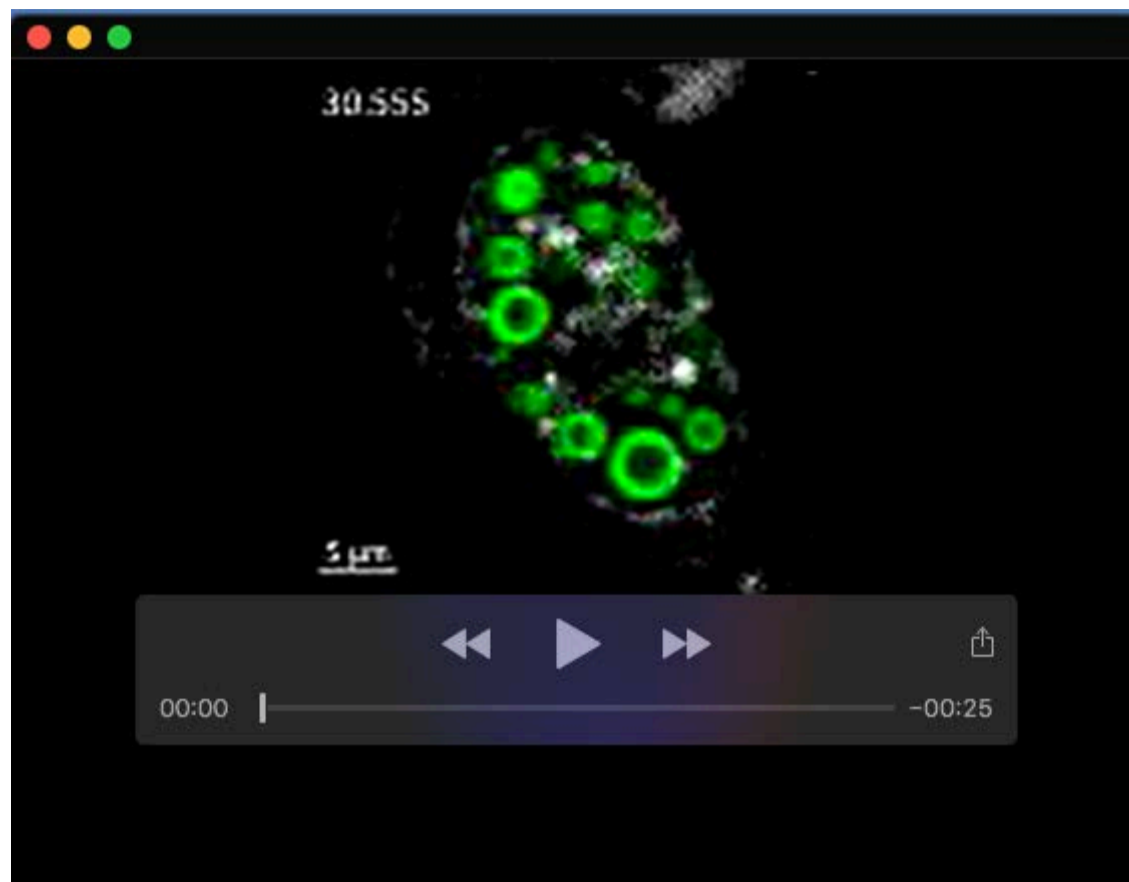
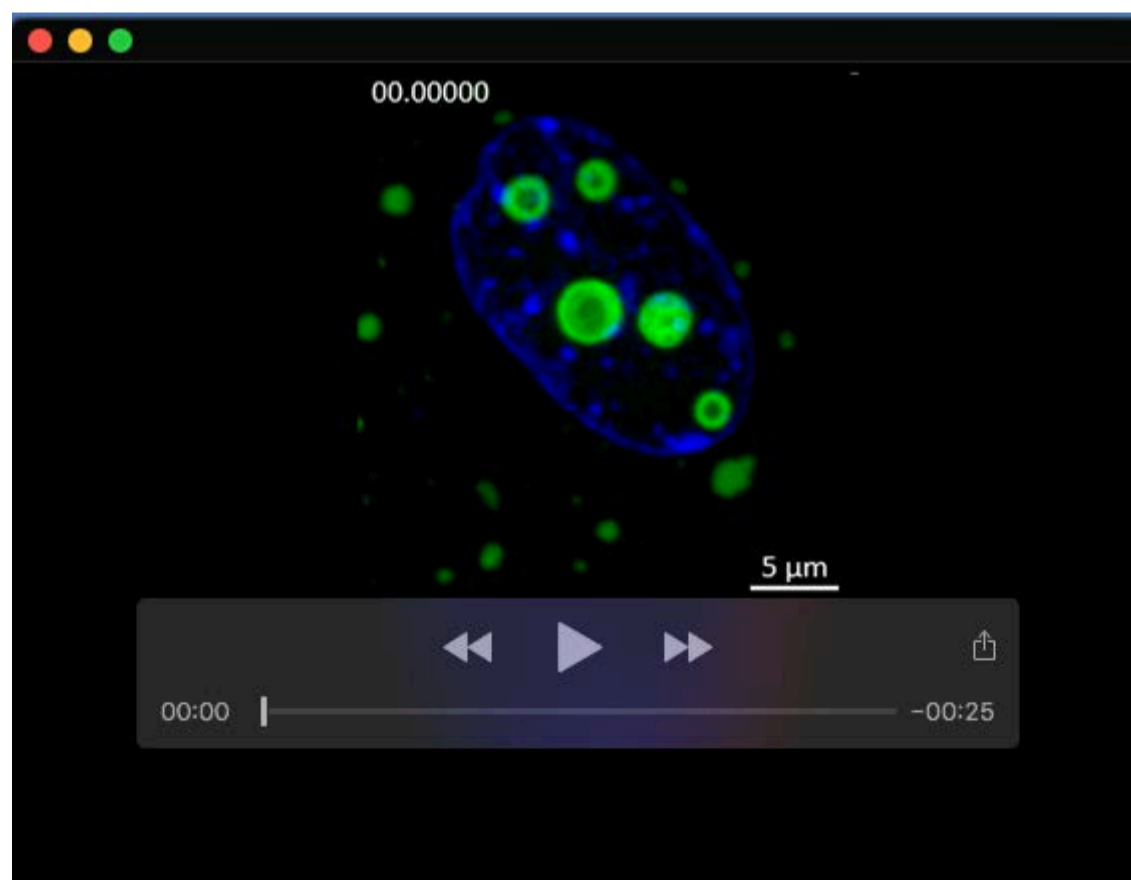


Fig. S4. C2C12 myogenic cells were transfected with the indicated expression constructs. After 1 day, the cells were subjected to live-cell confocal fluorescence microscopy. EYFP-β-catenin was localized to the nuclear envelope by an interaction trap using GBP-LaminB1. A line scan analysis along the indicated yellow line in the merged micrograph depicts recruitment of mCherry-TAZ to the nuclear envelope trapped EYFP-β-catenin, as indicated by arrows.



Movie 1. Time-lapse demonstration of nuclear condensate formation and fusion of EYFP-TAZ by live-cell imaging. Live cell spinning disk confocal imaging of ectopically expressed EYFP-TAZ (green) was carried out indicating the formation of spherical condensate structures inside the nucleus (imaged using Hoechst 33342 in blue). Videos document fusion properties of EYFP-TAZ condensates.



Movie 2. Time-lapse demonstration of nuclear condensate formation and fusion of EYFP-TAZ by live-cell imaging. Live cell spinning disk confocal imaging of ectopically expressed EYFP-TAZ (green) was carried out indicating the formation of spherical condensate structures inside the nucleus (imaged using Hoechst 33342 in white). Videos document fusion properties of EYFP-TAZ condensates.