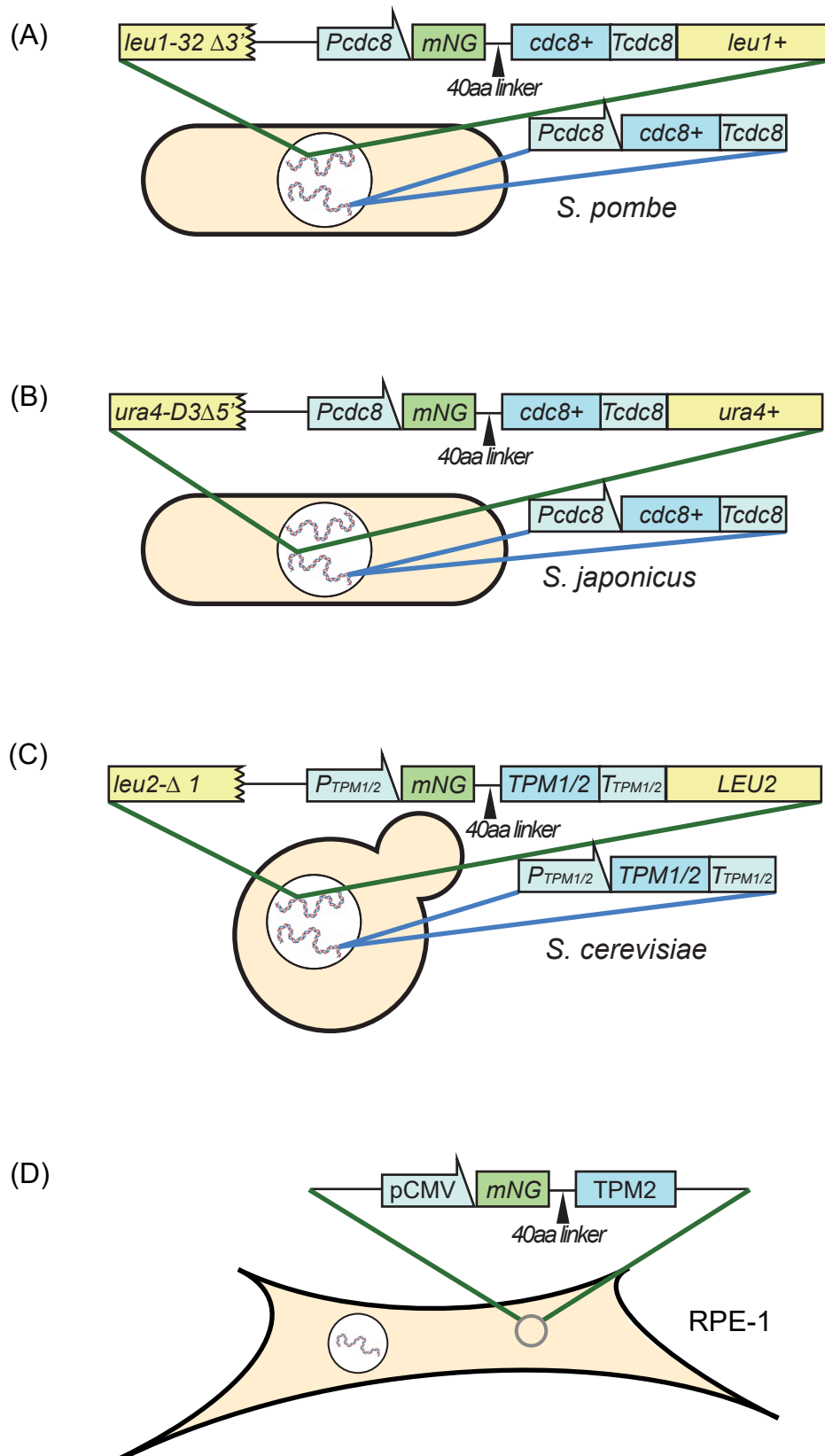
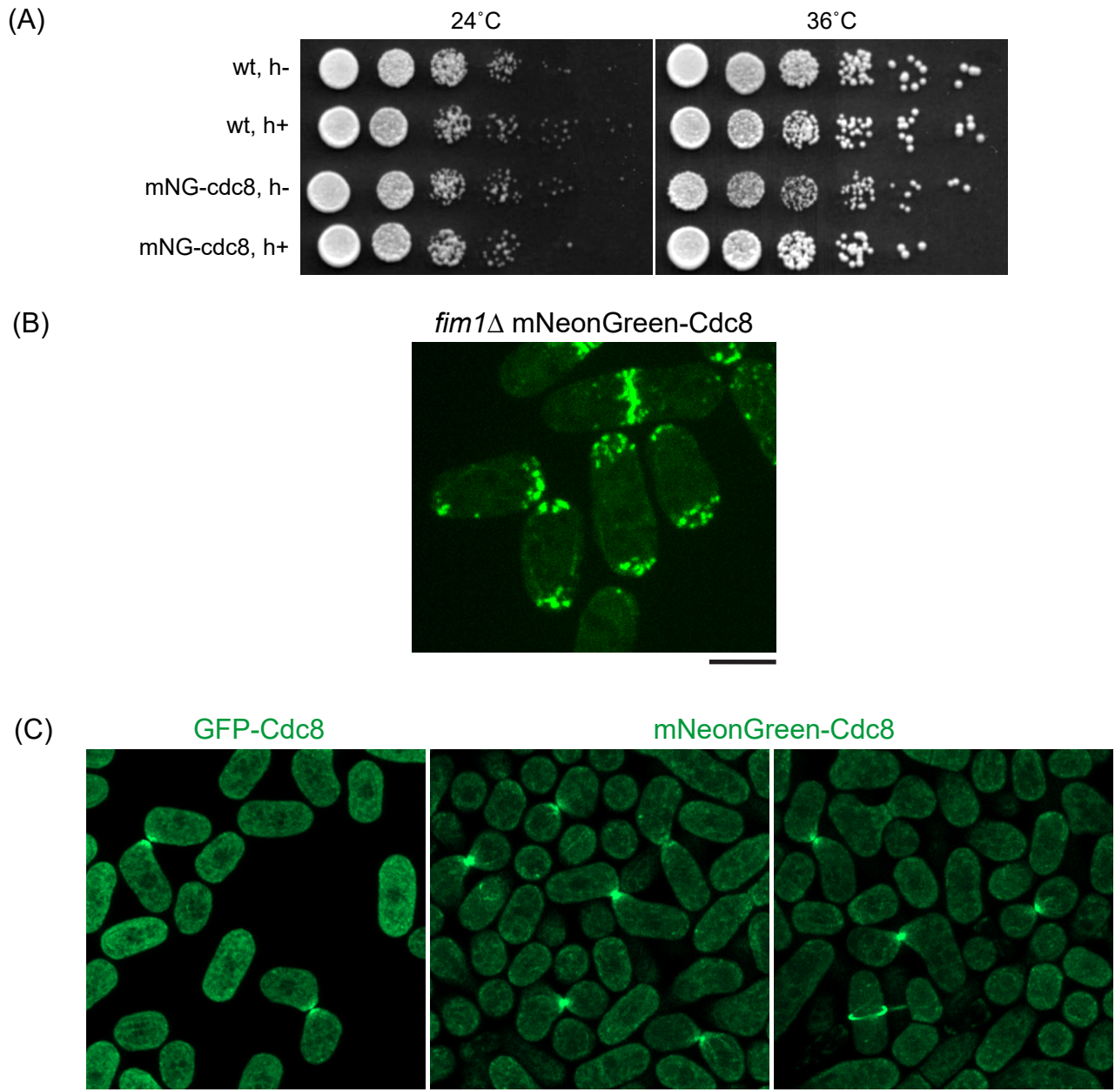


### Supplemental figure 1



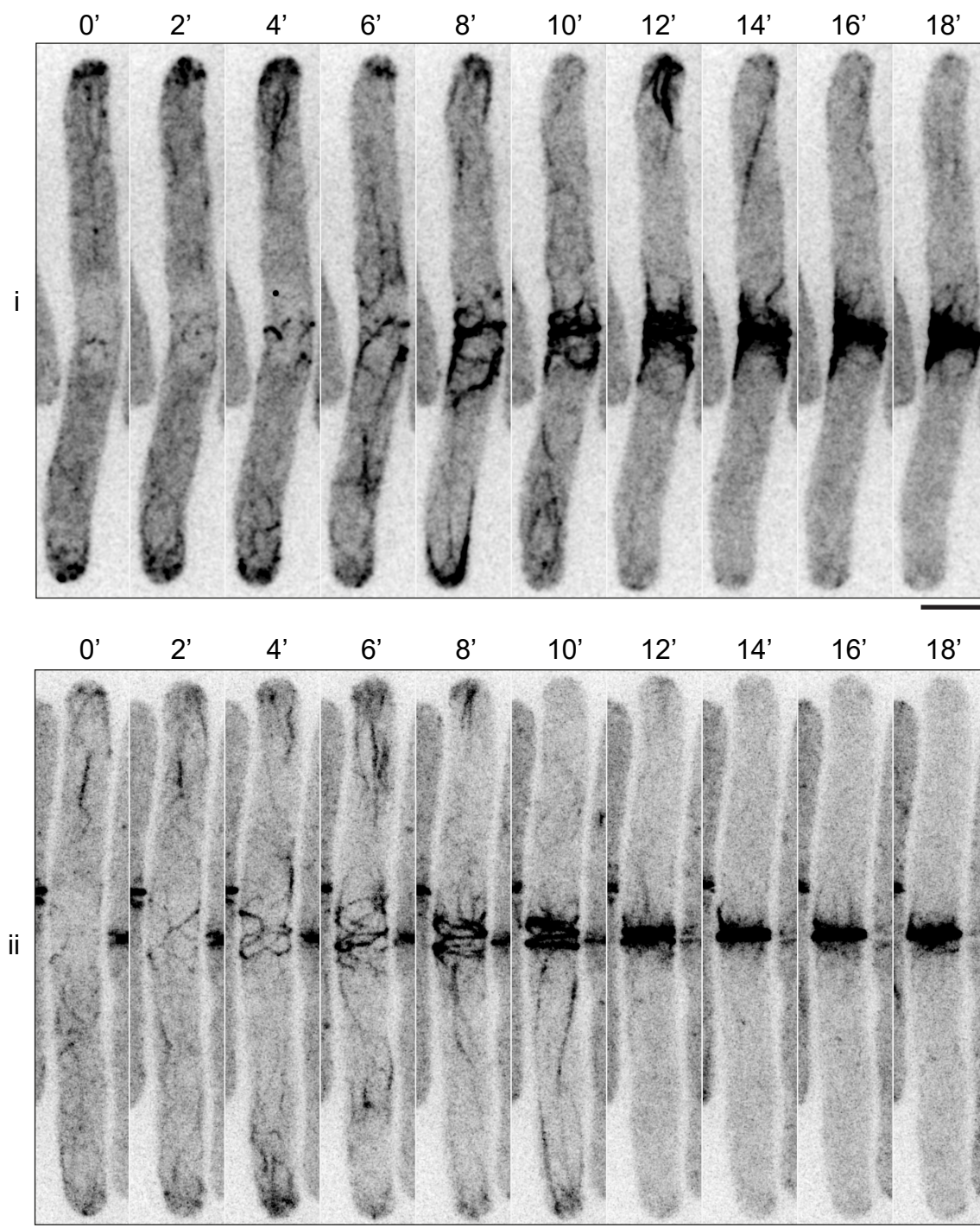
**Fig. S1. Strategy for tagging tropomyosin with mNeongreen:** (A) Schematic illustration showing the insertion of P<sub>*cdc8*</sub> mNG-40 amino acid linker-*cdc8* cassette at *leu1* locus. (B) Schematic illustration showing the insertion of P<sub>*cdc8*</sub> mNG-40 amino acid linker-*cdc8* cassette at the *ura4* locus. The tag was expressed under the native *cdc8* promoter. (C) Schematic illustration showing the insertion of P<sub>*Tpm1/2*</sub> mNG-40 amino acid linker-*Tpm1/2* cassette at the *leu2* locus. (D) Schematic illustration showing the expression of mNG-40 aa linker-TPM2 under the control of a CMV promoter in an RPE-1 cell.

## Supplemental figure 2



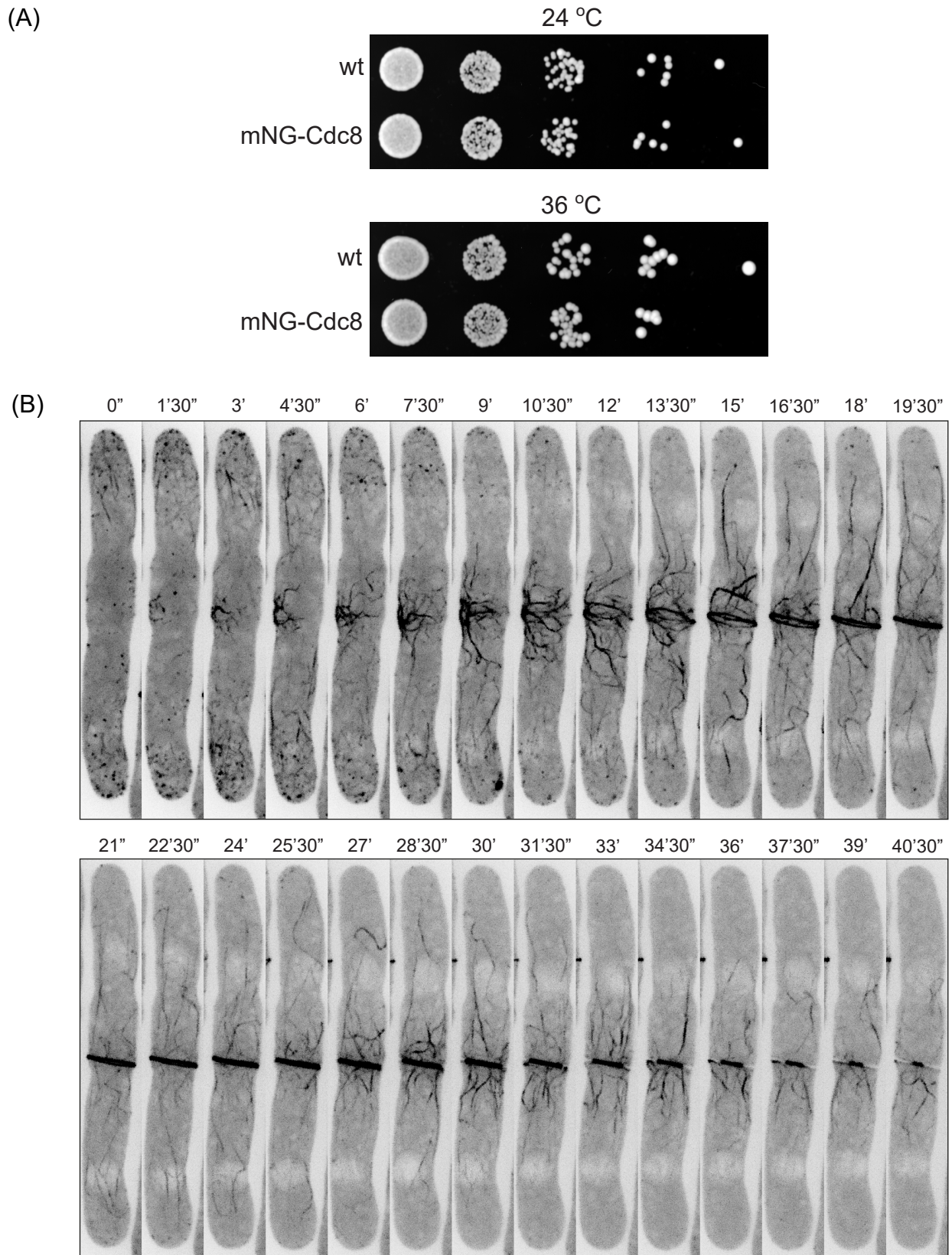
**Fig. S2. Tagging Cdc8 with mNeonGreen in *S. pombe*.** (A) Growth of strains was assessed by manual spot test. Untagged wild-type strains act as a control for mNG-Cdc8 strains. Cells were cultured in YEA media at 24°C to saturation, subjected to six-fold serial dilution, and spotted onto YEA agar. Plates were incubated at 24°C and 36°C for 3 days before being photographed. (B) Fimbrin null mutant cells (*fim1*Δ) cells show obvious mNG-Cdc8 patches. (n=34). (C) Airyscan2 images of mating cells expressing GFP-Cdc8 (left) or mNG-Cdc8 (centre; right) which labels the fusion focus. mNG-Cdc8 allows detection of actin cables not visible with GFP-Cdc8 (n=4). Higher cytosolic GFP signals were detected in the GFP-cdc8 cells. Scale bars are 5µm.

### Supplemental figure 3



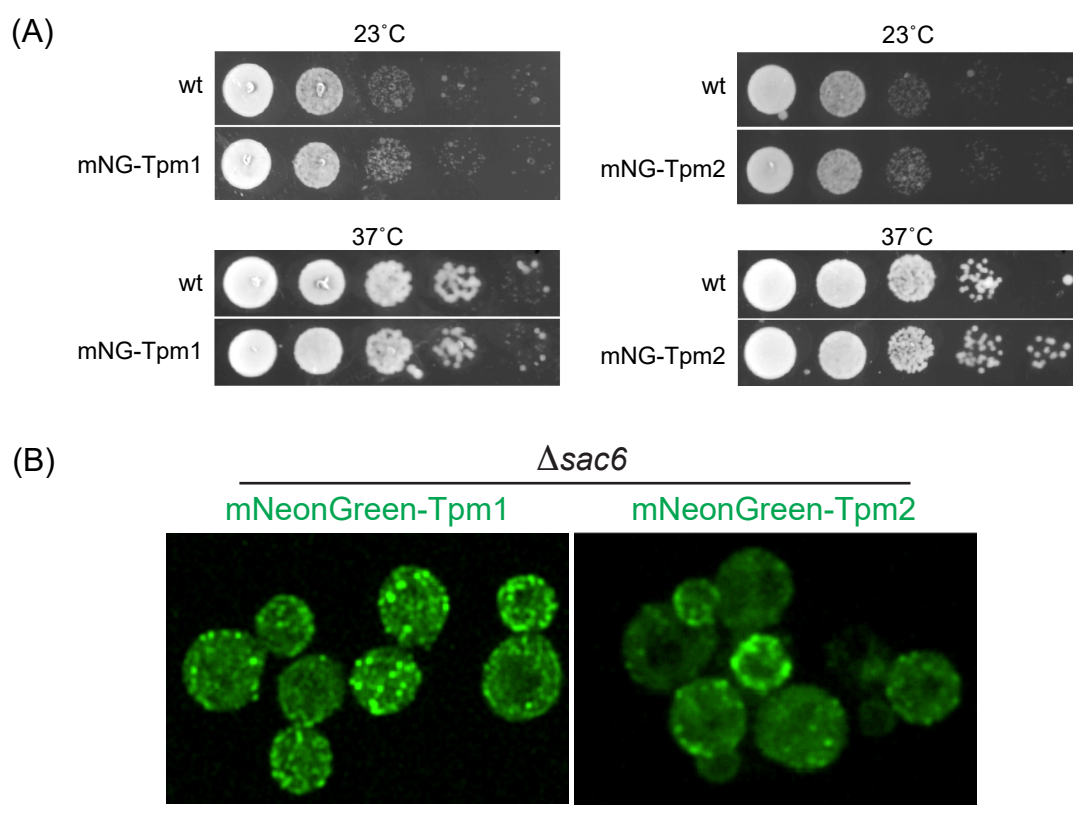
**Fig. S3. Actomyosin ring assembly visualized using mNG-Cdc8 in elongated *cdc25-22* cells.** Panel i and ii are time-lapse images of two *S. pombe cdc25-22* cells expressing mNG-Cdc8 demonstrating medial assembly of Cdc8 cables as well as flow of non-medial cables containing Cdc8-tropomyosin into the CAR. (n=65). Scale bars are 5 μm.

## Supplemental figure 4



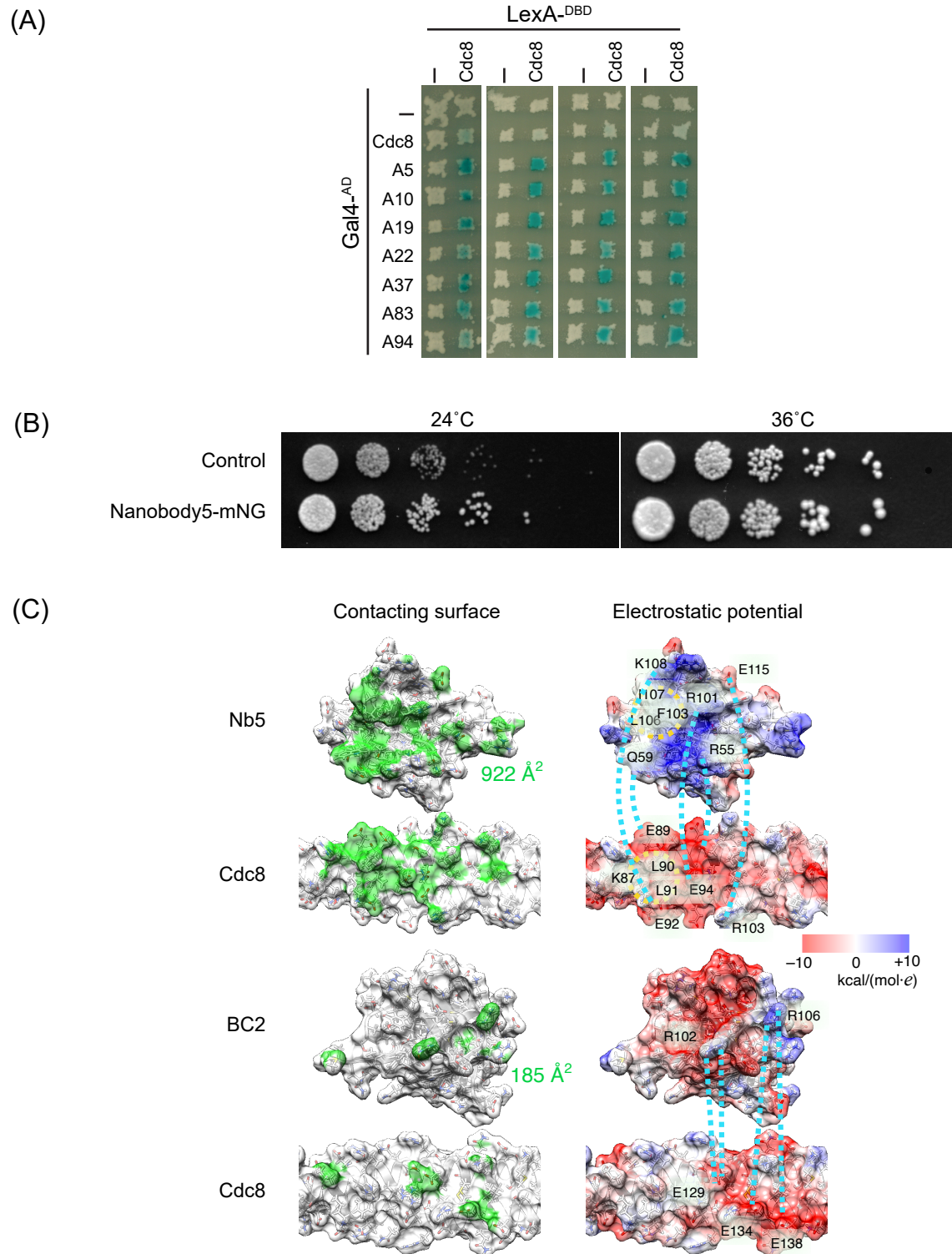
**Fig. S4. Tagging Cdc8 with mNG in *S. japonicus*.** (A) Growth of strains was assessed by manual spot test. Untagged wild-type strains act as a control for mNG-Cdc8 expressing strains. Cells were cultured in YEA media at 24°C to saturation, subjected to five-fold serial dilution, and spotted onto YEA agar. Plates were incubated at 24°C and 36°C for 3 days before being photographed. (B) Time-lapse images of an *S. japonicus cdc25-D9* cell expressing mNG-Cdc8 demonstrating medial assembly of Cdc8 cables as well as flow of non-medial cables containing Cdc8-tropomyosin into the CAR (n=12). Scale bar is 5µm.

### Supplemental figure 5



**Fig. S5. Tagging Tpm1 and Tpm2 with mNG in *S. cerevisiae* cells.** (A) Growth of strains was assessed by manual spot test. Untagged wild-type strains act as a control for mNG-tagged Tpm1 and Tpm2 strains. Cells were cultured in YPD media at 24°C to saturation, subjected to five-fold serial dilution, and spotted onto YPD agar. Plates were incubated at 23°C and 37°C for 3 days before being photographed. (B) Fimbrin null mutant cells (*sac6* $\Delta$  cells) show obvious mNG-Tpm1 (n=209) and mNG-Tpm2 (114) patches. Scale bar is 5 $\mu$ m.

## Supplemental figure 6



**Fig. S6. Cdc8 specific Nanobody-mNeonGreen in *S. pombe*.** (A) Yeast two hybrid interaction shown between Sp Cdc8 and 7 camelid nanobodies. (B) Growth of strains was assessed by manual spot test. Untagged wild-type strains act as a control for Nb5-mNG strains. Cells were cultured in YEA media at 24°C to saturation, subjected to six-fold serial dilution, and spotted onto YEA agar. Plates were incubated at 24°C and 36°C for 3 days before being photographed. (C) Alphafold 2 prediction of Cdc8-tropomyosin–Nb5 contacting surface sites and their electrostatic potential.

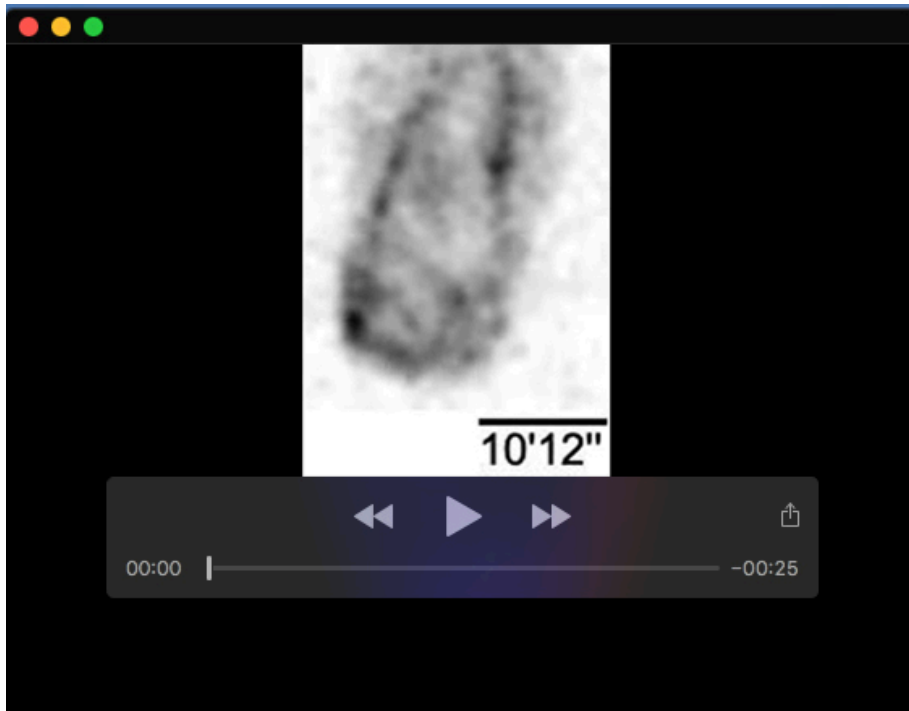


**Table S1. Yeast strains used in this work**

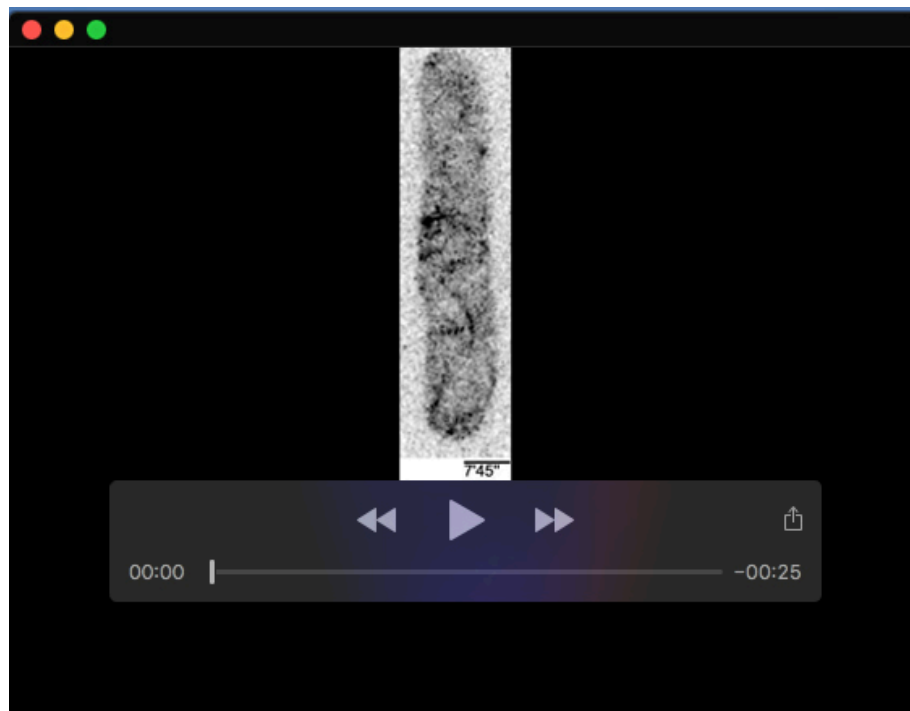
	Strain number	Genotype
<i>S. pombe</i>	MBY101	<i>ade6-210 ura4-D18 leu1-32 h-</i>
	MBY102	<i>ade6-210 ura4-D18 leu1-32 h+</i>
	MBY192	<i>ura4-D18 leu1-32 h-</i>
	MBY12825	<i>leu1-32 pDUAL:p<sup>cdc8</sup>:mNeongreen-40aa:cdc8 ura4-D18 ade6-210, cdc8+ h-</i>
	MBY12828	<i>leu1-32 pDUAL:p<sup>cdc8</sup>:mNeongreen-40aa:cdc8 ura4-D18 ade6-210, cdc8+ h+</i>
	MBY12947	<i>leu1-32 pDUAL-p<sup>cdc8</sup>:mNeongreen-40aa:cdc8 lys1&lt;&lt;pLYS1U-P<sup>act1</sup>:lifeact-mCherry:ura4+ ade6-210, cdc8+</i>
	MBY12994	<i>fim1Δ leu1-32 pDUAL:p<sup>cdc8</sup>:mNeongreen-40aa:cdc8, ura4-D18, ade6-210, cdc8+</i>
	MBY13071	<i>cdc25-22 leu1-32 pDUAL:p<sup>cdc8</sup>:mNeongreen-40aa:cdc8 rlc1-mCherry:ura4+ ade6-210 h+</i>
	MBY13185	<i>leu1-32 p<sup>adh11</sup>:Nanobody5-mNeonGreen</i>
	YSM3935	<i>h90 myo52-tdTomato:natMX leu1-32:p<sup>cdc8</sup>:mNeonGreen-cdc8:term<sup>cdc8</sup>:term<sup>ADH1</sup>:leu1+ lys3+:p<sup>map3</sup>:mCherry:term<sup>ADH1</sup>:bsdMX ade6-M216 ura4-D18</i>
	YSM3936	<i>h90 myo52-tdTomato:natMX leu1-32:p<sup>ADH1</sup>:cdc8Nb5-mNeonGreen:term<sup>ADH1</sup>:leu1+ ade6-M210 ura4-294</i>
	YSM3316	<i>h90 myo52-tdTomato:natMX leu1-32:p<sup>ADH1</sup>:cdc8Nb5-mNeonGreen:term<sup>ADH1</sup>:leu1+ ade6-M210 ura4-294</i>
<i>S. japonicus</i>	SOJ5	<i>matsj-P2028 h-</i>
	SOJ4909	<i>pcdc8-mNeonGreen-40 a.a. linker-cdc8<sup>ORF</sup>-cdc8<sup>3'UTR</sup>::ura4+::ura4sj-D3 h+</i>
	SOJ5001	<i>pcdc8-mNeonGreen-40 a.a. linker-cdc8<sup>ORF</sup>-cdc8<sup>3'UTR</sup>::ura4+::ura4sj-D3 cdc25-D9:kanR:ura4+</i>
	SOJ5221	<i>pcdc8-mNeonGreen-40 a.a. linker-cdc8<sup>ORF</sup>-cdc8<sup>3'UTR</sup>::ura4+::ura4sj-D3 h-</i>
<i>S. cerevisiae</i>	YSP002	ESM356 MATa <i>ura3-52 leu2Δ1 trp1Δ63 his3Δ200</i> (wild type)
	YSP107	As YSP002 except pRS305-P <sub>tpm1</sub> -mNG-40aaL- <i>tpm1-T<sub>tpm1</sub>-leu2</i>
	YSP108	As YSP002 except pRS305-P <sub>tpm2</sub> -mNG-40aaL- <i>tpm2-T<sub>tpm2</sub>-leu2</i>
	YSP191	As YSP002 except pRS305-P <sub>tpm1</sub> -mNG-40aaL- <i>tpm1-T<sub>tpm1</sub>-leu2 Δsac6::His3MX6 abp1-tdtomato::natNT2</i>
	YSP192	As YSP002 except pRS305-P <sub>tpm2</sub> -mNG-40aaL- <i>tpm2-T<sub>tpm2</sub>-leu2 Δsac6::His3MX6 abp1-tdtomato::natNT2</i>

**Table S2. Plasmids used in this study**

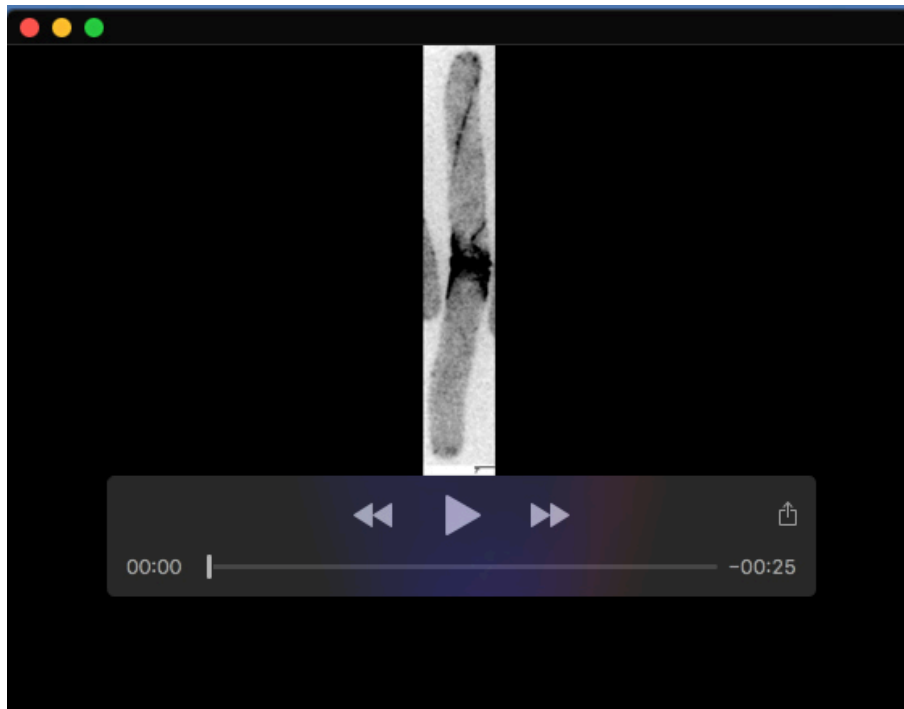
Plasmid name	Description	Reference			
<b>pTH8-77</b>	pDUAL-cdc8(5'UTR)-mNG-40 a.a. linker-cdc8-cdc8(3'UTR)	This Study			
<b>pMB117</b>	pDUAL-Padh11-Nb A5-40 a.a. linker-mNG	This Study			
<b>pSO1233</b>	pBluescriptSK(+)-ura4+( <i>S. japonicus</i> )-cdc8(5'UTR)-mNG-40 a.a. linker-cdc8-cdc8(3'UTR)	This Study			
<b>piSP347</b>	pRS305-Ptpm1-mNG-40aaL-tpm1-Ttpm1	This Study			
<b>piSP349</b>	pRS305-Ptpm2-mNG-40aaL-tpm2-Ttpm2	This Study			
<b>piSP23</b>	pFA6a-His3MX6	Knop et al 1999			
<b>pMB128</b>	pDUAL-Padh11-mNeongreen-40aaL	MB lab			
<b>piSP112</b>	pMM5s	This Study			
<b>piSP113</b>	pMM6s	This Study			
<b>piSP600</b>	pMM5S-Cdc8	This Study			
<b>piSP604</b>	pMM6S-Cdc8	This Study			
<b>piSP198</b>	pMM5S-Cdc8 Nb A5	This Study			
<b>piSP200</b>	pMM5S-Cdc8 Nb A10	This Study			
<b>piSP202</b>	pMM5S-Cdc8 Nb A19	This Study			
<b>piSP203</b>	pMM5S-Cdc8 Nb A22	This Study			
<b>piSP204</b>	pMM5S-Cdc8 Nb A37	This Study			
<b>piSP206</b>	pMM5S-Cdc8 Nb A83	This Study			
<b>piSP207</b>	pMM5S-Cdc8 Nb A94	This Study			
<b>piSP209</b>	pMM6S-Cdc8 Nb A5	This Study			
<b>piSP211</b>	pMM6S-Cdc8 Nb A10	This Study			
<b>piSP213</b>	pMM6S-Cdc8 Nb A19	This Study			
<b>piSP215</b>	pMM6S-Cdc8 Nb A22	This Study			
<b>piSP217</b>	pMM6S-Cdc8 Nb A37	This Study			
<b>piSP218</b>	pMM6S-Cdc8 Nb A83	This Study			
<b>piSP219</b>	pMM6S-Cdc8 Nb A94	This Study			



**Movie 1.** *S. pombe* mNG-Cdc8 patch and cable dynamics.



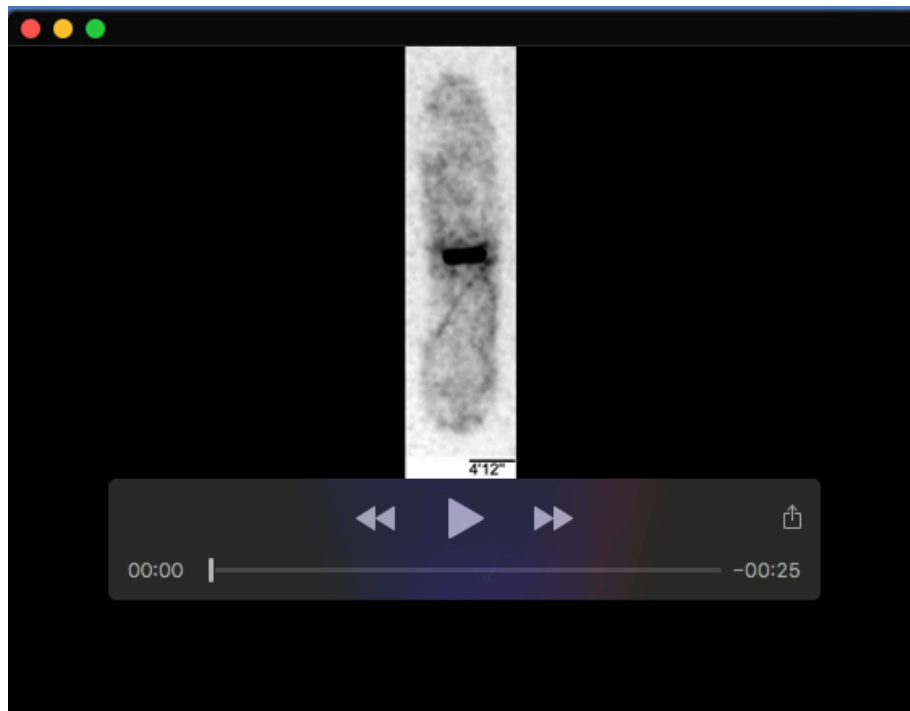
**Movie 2.** mNG-Cdc8 dynamics during CAR assembly in *S. pombe*



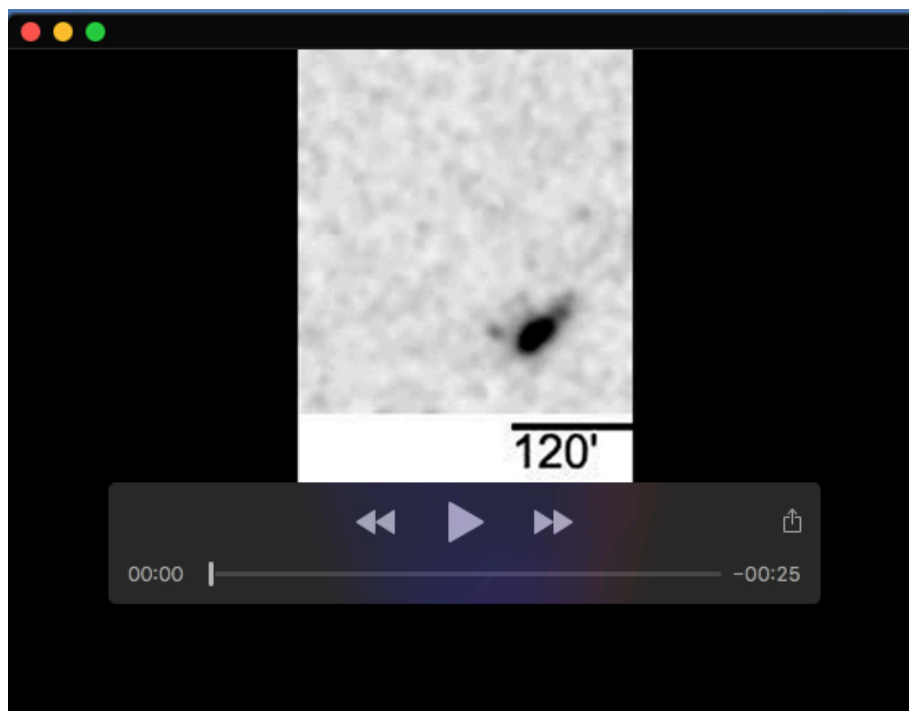
**Movie 3.** mNG-Cdc8 dynamics during CAR assembly in highly elongated *S. pombe* *cdc25-22* cells, demonstrating flow of Cdc8 cables into the CAR during its assembly.



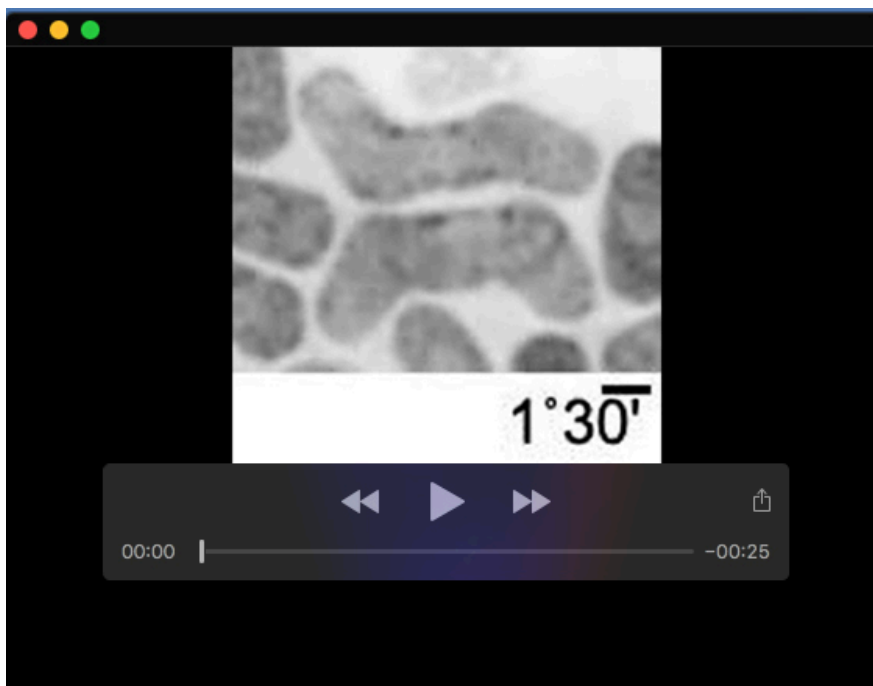
**Movie 4.** mNG-Cdc8 dynamics during CAR assembly in highly elongated *S. pombe* *cdc25-22* cells, demonstrating flow of Cdc8 cables into the CAR during its assembly.



**Movie 5.** mNG-Cdc8 dynamics and cable expulsion during CAR constriction in *S. pombe*.



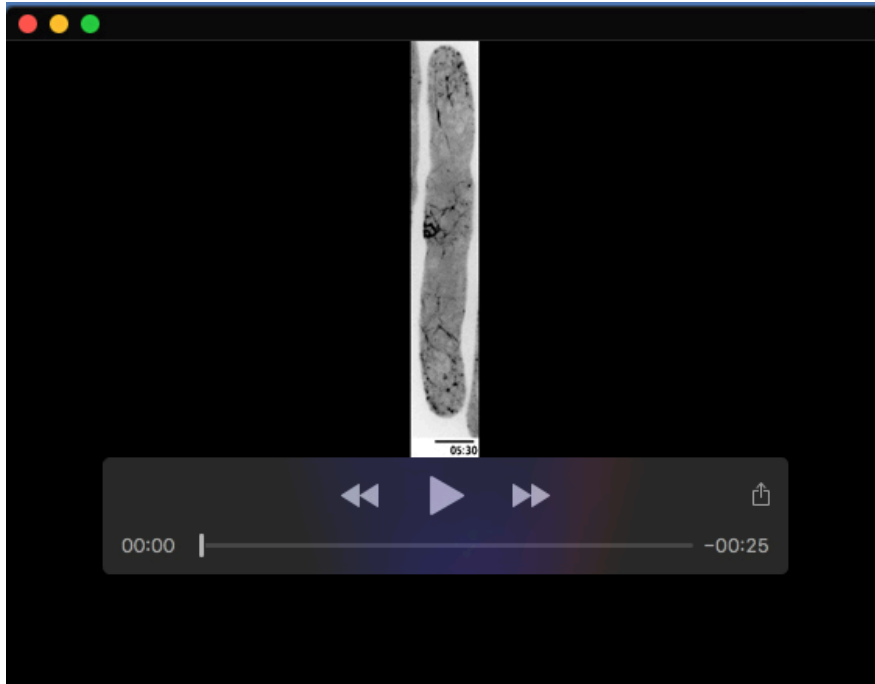
**Movie 6.** Dynamics of mNG-Cdc8 during ATP-dependent constriction of CARs within *S. pombe* cell ghosts.



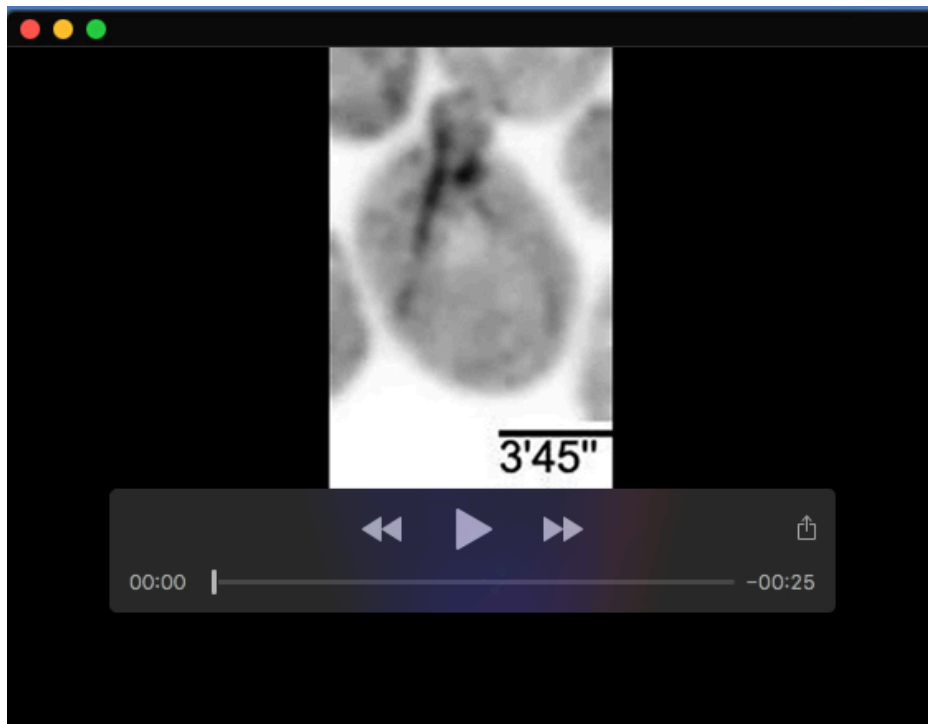
**Movie 7.** Dynamics of *S. pombe* mNG-Cdc8 during mating and sporulation.



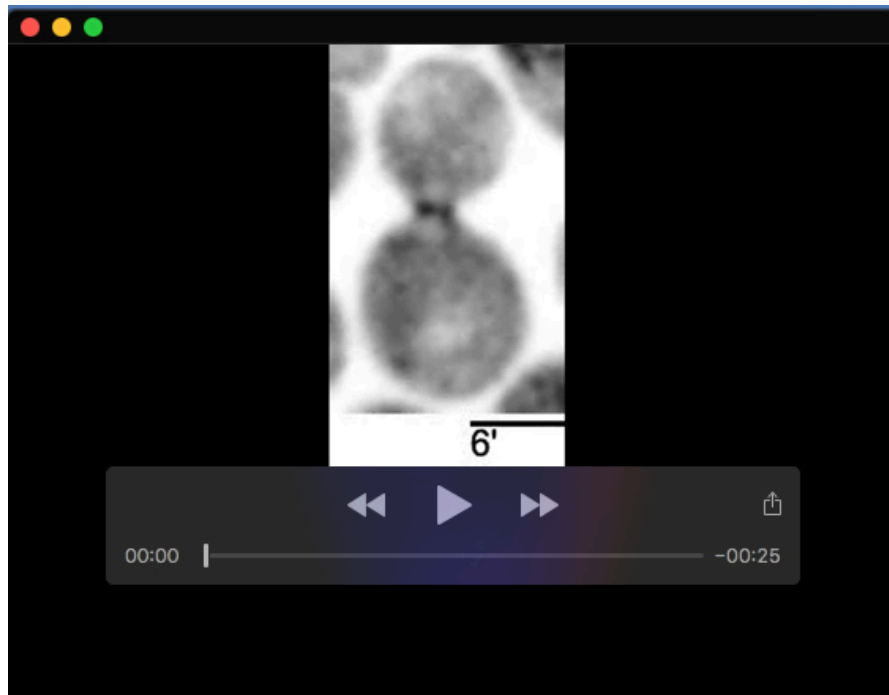
**Movie 8.** Dynamics of *S. japonicus* mNG-Cdc8 during CAR assembly and constriction.



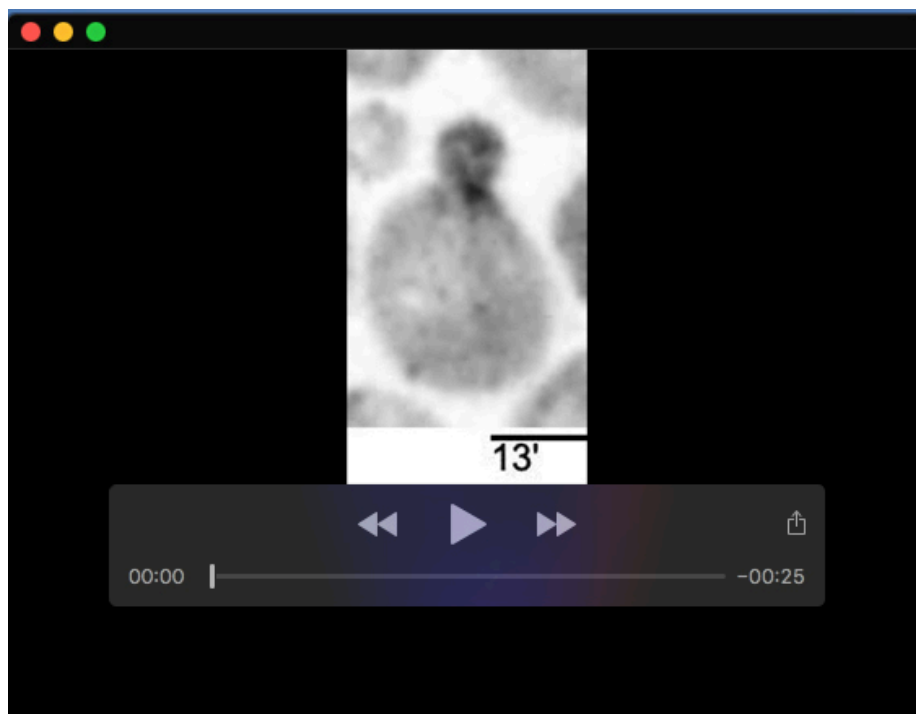
**Movie 9.** Dynamics of *S. japonicus* mNG-Cdc8 during CAR assembly and constriction in elongated *cdc25-D9* cells.



**Movie 10.** Dynamics of *S. cerevisiae* mNG-Tpm1 in cables.

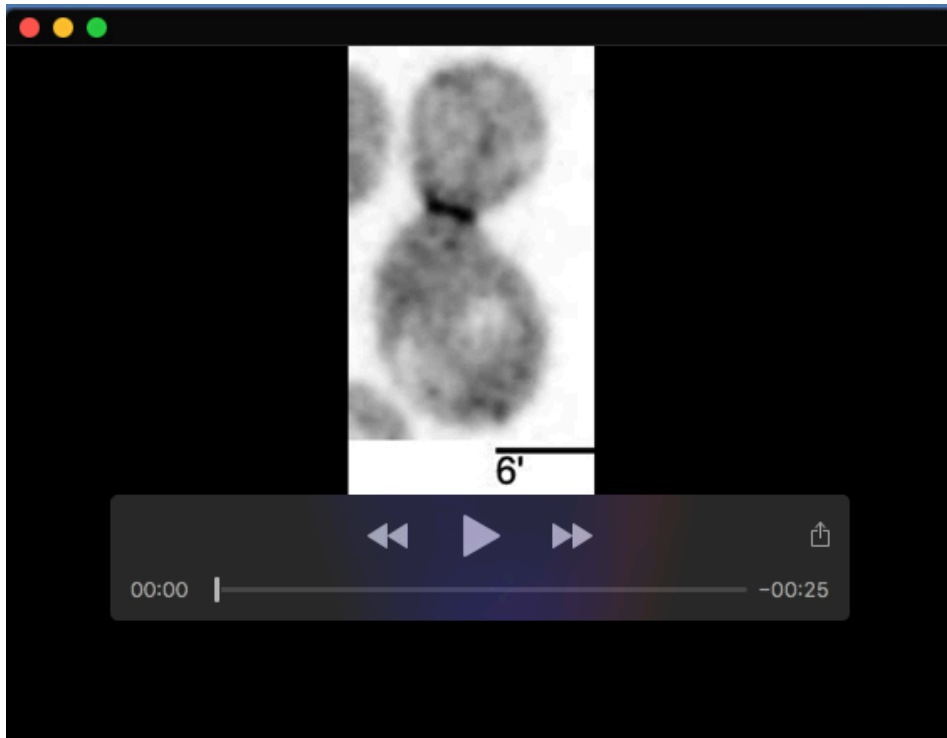


**Movie 11.** Dynamics of *S. cerevisiae* mNG-Tpm1 in the CAR.

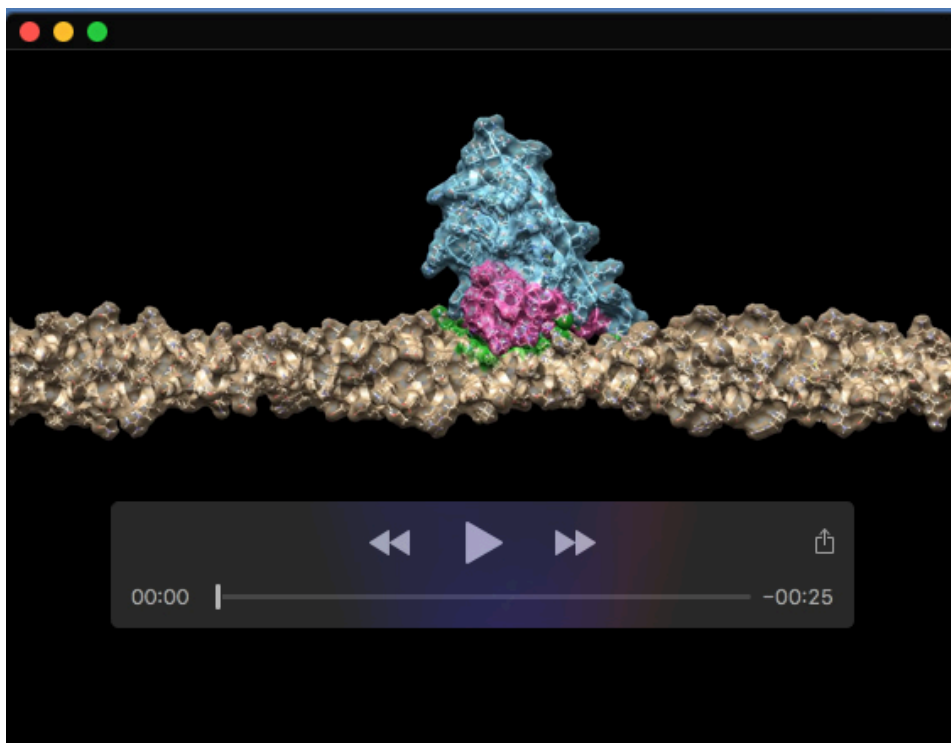


**Movie 12.** Dynamics of *S. cerevisiae* mNG-Tpm2 in cables.

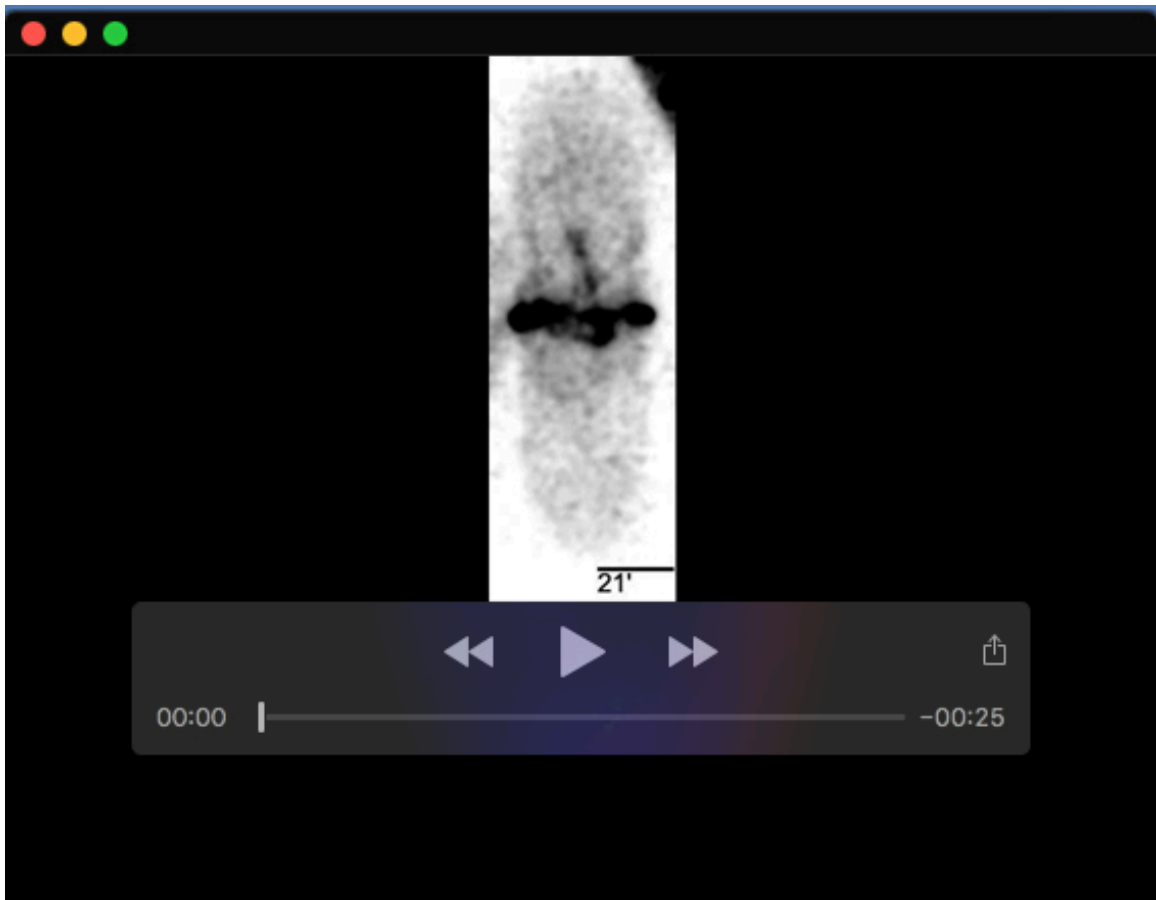




**Movie 13.** Dynamics of *S. cerevisiae* mNG-Tpm2 in the CAR.



**Movie 14.** Simulation of Cdc8-Nanobody 5 interaction with Cdc8 dimer.



**Movie 15.** Dynamics of mNG-Nb5 during CAR assembly in *S. pombe*.