

Fig. S1. Inflammasome activation in THP-1 producer cells. (A) Western blot analysis of producer THP-1 cells treated with 100ng/ml LPS for 24hr. The amount of mature IL-1 β is increased after LPS treatment. (B) IL-1 β ELISA of conditioned media isolated from producer THP-1 cells treated with LPS. Data are shown as mean \pm SEM for $n = 4$.

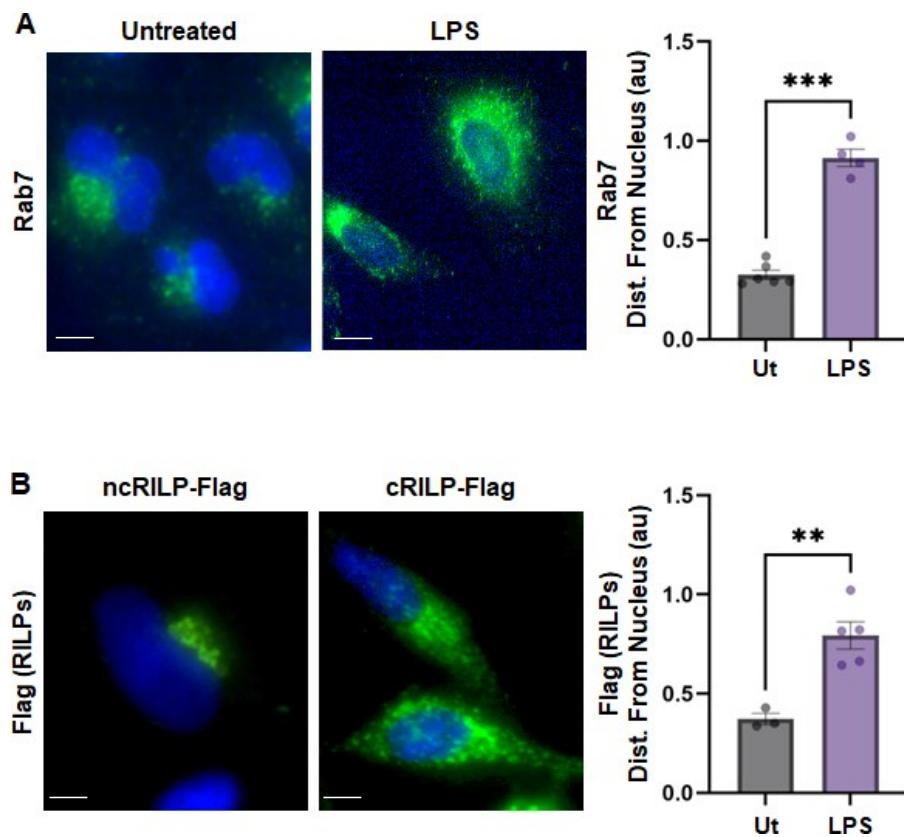


Fig. S2. Localization of Rab7 as a surrogate for RILP cleavage. (A) In HMC3 microglia cells, Rab7 serves as a marker of RILP cleavage in immunofluorescence. At baseline, Rab7 localizes at the perinuclear region, near the mitotic center. This localization mirrors that of non-cleavable RILP (B, ncRILP). After RILP cleavage via inflammasome activation with LPS, Rab7 redistributes throughout the cellular periphery and extends to the plasma membrane. This localization is similar to that seen when cleaved RILP is overexpressed (B, cRILP). The bar graphs represent an ImageJ analysis measuring the distance the Rab7^{+ve} or RILP^{+ve} puncta are from the nucleus. Data are shown as mean ± SEM; *, P ≤ 0.05 for n = a minimum of 15 cells from 3-5 individual experiments. Scale bar: 10μm.

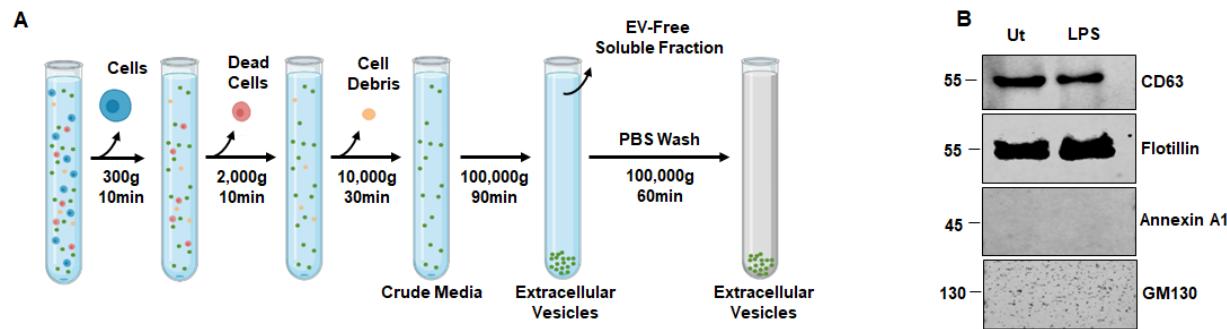


Fig. S3. (A) S3. (A) Cell culture supernatants were collected from control and LPS-treated THP-1 cells and subjected to differential centrifugation to separate the EV-free soluble fraction from the EV-enriched pellet. (B) Western blot analysis showed that the EV pellets contain known exosomal markers (CD63 and Flotillin) but do not contain the microvesicle marker annexin A1 or components of other organelles including the Golgi.

Table S1. Primers used in this study.

Primer Name	Primer Sequence (5'-3')
hIL-1 β Forward	CATGGGATAACGAGGGCTTATGT
hIL-1 β Reverse	CCCAAGGCCACAGGTATT
hIL6_Forward	ACTCACCTCTCAGAACGAATTG
hIL6_Reverse	CCATCTTCCAAGGTTCAGGTTG
hIL33_Forward	GTGACGGTGTGATGGTAAGAT
hIL33_Reverse	AGCTCCACAGAGTGTTCCTTG
hCRP Forward	AGACATGTCGAGGAAGGCTTT
hCRP Reverse	TCGAGGACAGTCCGTAGAA
hHMGB-1-Forward	GGACAAGGCCGTTATGAAA
hHMGB-1-Reverse	GCAGAAGAGGAAGAAGGCCGAA
hCCL2_Forward	ATGAAAGTCTCTGCCGCCCTCT
hCCL2_Reverse	TGAGTGTCAAGTCTCGGAGTT
hTNF α _Forward	TCTTCTCGAACCCCCGAGTGA
hTNF α _Reverse	CCTCTGATGGCACCAACCAG
hIL10_Forward	TCAAGGCGCATGTGAACCTCC
hIL10_Reverse	GATGTCAAACTCACTCATGGCT
hTGF β -1forward	TATGCCAGGAATTGTTGCTG
hTGF β -1 reverse	CAATTCCCTGGCGATACCTCAG
hGAPDH-F	GAAGGTGAAGGTGGAGTC
hGAPDH-R	GAAGATGGTATGGGATTTC
mCRP Forward	GAACTTTCAGCCGAATACATCTTT
mCRP Reverse	CCTTCCTCGACATGTCTGTCT
mIL-1 β -Forward	TCGCTCAGGGTCACAAGAAA
mIL-1 β -Reverse	CATCAGAGGCAAGGAGGAAAAC
mTGF- β -Forward	GTGTGGAGCAACATGTGGAACCTCA
mTGF- β -Reverse	TTGGTTCAGCCACTGCCGTA
mIL-10-Forward	GGTTGCCAAGCCTTATCGGA
mIL-10-Reverse	ACCTGCTCCACTGCCTTGCT
mTNF α -Forward	AGGCTCTGGAGAACAGCACAT
mTNF α -Reverse	TGGCTTCTCTCCTGCACCAAA
mArg1-Forward	CTCCAAGCCAAAGTCCTTAGAG
mArg1-Reverse	AGGAGCTGTCATTAGGGACATC
mCCL2-Forward	GTTGGCTCAGCCAGATGCA
mCCL2-Reverse	AGCCTACTCATTGGGATCATCTTG
mHMGB-1-Forward	GGCGAGCATCCTGGCTTATC
mHMGB-1-Reverse	GGCTGCTTGTCACTGCTG
mIL6_Forward	TAGCCTTCCTACCCCAATTCC
mIL6_Reverse	TTGGTCCTTAGCCACTCCTTC
mIL33_Forward	TCCAACCTCAAGATTCCCCG
mIL33_Reverse	CATGCAGTAGACATGGCAGAA
mGAPDH-6-Forward	CGTCCCGTAGACAAAATGGT
mGAPDH-6-Reverse	TTGAGGTCAATGAAGGGGTC