## **Supplementary Materials and Methods**

Proximate Analysis

Frozen fish remains were homogenized using a Fisher Brand Bead Mill 24 and subsamples of the homogenate were weighed and freeze dried (Labconco Lyophilizer). *Protein:* Protein content was estimated in triplicate (intra-assay CV% <10%) using a BCA assay with a 72% TCA precipitation (Pierce BCA kit, ThermoFisher Scientific, MA, USA), where absorbance was measured at 562 nm. *Lipids:* Lipid content was estimated using a chloroform:methanol extraction as described in Mann and Gallager, 1985 and Johnson et al, 2017. Lipids from 50 mg of freeze-dried homogenized sample were extracted using 100 ul milliQ water and 1.5 ml chloroform:methanol (1:2) (vortexed, incubated at 4°C, centrifuged at 4000 rpm for 5 min). The supernatant was removed and remaining sample was re-extracted in 1.5 ml chloroform:methanol (2:1). The supernatants were pooled, mixed with 950 ul NaCl (0.7%), incubated at 4°C for 30 min, then centrifuged (4000 rpm, 5 min), and the volume of the bottom layer was measured. Dried subsamples of the bottom layer were used to extrapolate lipid content to the entire sample. *Ash Content:* Ash content was determined by drying freeze-dried samples overnight at 100°C to account for any moisture that returned during sample storage. Samples were then weighed (~30 mg) before being combusted in a muffle furnace at 450°C for 12 h and then re-weighed.

**Table S1**. Dietary and whole-body Proximate composition (% wet weight)

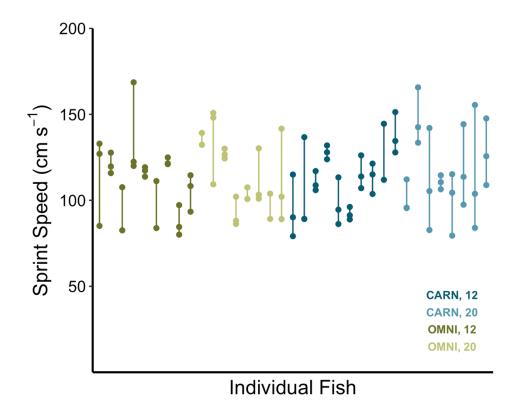
Dietary Proximate composition (% wet weight)									
	Experiment 1		Experiment 2						
	Ulva	Artemia	Ulva	Artemia					
% Moisture	$82.04 \pm 1.63$	$87.48 \pm 0.91$	$75.33 \pm 3.81$ $86.83 \pm 0.38$						
% Protein	$1.47 \pm 0.27$	$4.75 \pm 0.51$	$1.95 \pm 0.88$	$5.59 \pm 0.62$					
% Lipid	$0.42 \pm 0.05$	$1.23 \pm 0.14$	$0.55 \pm 0.10$	$1.84 \pm 0.08$					
% Ash	$10.71 \pm 1.87$	1.44 ± NA	9.93 ± NA	$1.78 \pm 0.04$					
Whole body Proximate composition (% wet weight)									
	12°C		20°C						
	Carnivorous	Omnivorous	Carnivorous	Omnivorous					
% Moisture	$70.15 \pm 1.15$	$72.25 \pm 1.40$	$72.98 \pm 0.74$	$71.79 \pm 0.45$					
% Protein	$13.40 \pm 1.09$	13.91 ± 1.21	12.61 ± 1.45	$10.76 \pm 0.85$					
% Lipid	$3.88 \pm 0.41$	$3.08 \pm 0.25$	$3.70 \pm 0.63$ $3.93 \pm 0.10$						
% Ash	$5.45 \pm 0.84$	$4.31 \pm 0.50$	$4.60 \pm 0.60$	$5.26 \pm 0.60$					

Represented are means and standard error values for dietary proximate composition in *Ulva* sp., *Artemia* sp., and proximate body composition from whole opaleye from experiments 1 and 2. Proximate body composition were statistically analyzed using 2-way ANOVA and no significant differences were found between treatment groups. When sample size <3 standard error was not calculated and is listed as NA.

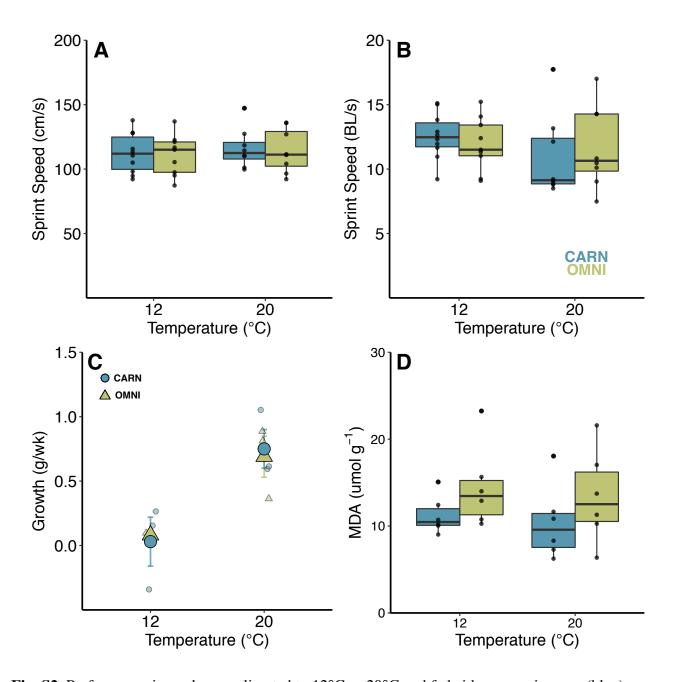
Table S2. AIC Outputs for Polynomial Curves.

AIC outputs for warm ABT test f <sub>hmax</sub> polynomial curves							
Model	Formula		AIC	ΔAIC			
Model 1	poly(acute_temp, 3) * diet * temp + (1   fish_id)	18	5282.61153	0			
Model 2	poly(acute_temp, 3) * temp + diet + (1   fish_id)	11	5295.12706	12.515531			
Model 3	poly(acute_temp, 3) * temp + (1   fish_id)	10	5297.84122	15.2296905			
Model 4	poly(acute_temp, 2) * diet * temp + (1   fish_id)	14	5331.73874	49.1272062			
Model 5	poly(acute_temp, 3) * diet + temp + (1   fish_id)		5425.26489	142.653365			
Model 6	poly(acute_temp, 4) + temp + diet + (1   fish_id)		5427.25882	144.64729			
Model 7	poly(acute_temp, 4) + temp * diet + (1   fish_id)		5428.15216	145.540635			
Model 8	poly(acute_temp, 3) + temp + diet + (1   fish_id)	8	5429.2756	146.664069			
Model 9	poly(acute_temp, 3) + temp * diet + (1   fish_id)	9	5430.17554	147.564011			
Model 10	poly(acute_temp, 3) + temp + (1   fish_id)	7	5432.27141	149.659875			
Model 11	poly(acute_temp, 3) + diet + (1   fish_id)	7	5433.3873	150.775766			
Model 12	poly(acute_temp, 3) + (1   fish_id)	6	5435.91176	153.300235			
Model 13	poly(acute_temp, 2) + temp + diet + (1   fish_id)	7	5491.9684	209.356872			
Model 14	poly(acute_temp, 2) + temp * diet + (1   fish_id)	8	5492.77923	210.167701			
Model 15	acute_temp + temp + diet + (1   fish_id)	6	5742.45159	459.840058			
Model 16	acute_temp + temp * diet + (1   fish_id)	7	5743.27212	460.660589			
Model 17	acute_temp + temp + (1   fish_id)	5	5745.43438	462.822848			
Model 18	acute_temp + diet + (1   fish_id)	5	5749.62894	467.017406			
Model 19	acute_temp + (1   fish_id)	4	5751.89944	469.287913			
AIC outputs for cold test f <sub>hmax</sub> polynomial curves							
Model	Formula	df	AIC	ΔAIC			
Model 1	poly(acute_temp, 4) + diet + (1   fish_id)	8	1620.06402	0			
Model 2	poly(acute_temp, 4) * diet + (1   fish_id)	12	1623.74238	3.67835332			
Model 3	poly(acute_temp, 3) + diet + (1   fish_id)	7	1636.74603	16.6820023			
Model 4	poly(acute_temp, 3) * diet + (1   fish_id)	10	1639.36915	19.3051258			
Model 5	poly(acute_temp, 3) + (1   fish_id)	6	1640.02832	19.9642973			
Model 6	poly(acute_temp, 2) * diet + (1   fish_id)	8	1640.27173	20.2077023			
Model 7	poly(acute_temp, 2) + diet + (1   fish_id)	6	1641.6658	21.6017758			
Model 8	acute_temp + diet + (1   fish_id)	5	1992.02421	371.960185			
Model 9	acute_temp + (1   fish_id)	4	1994.25531	374.191287			

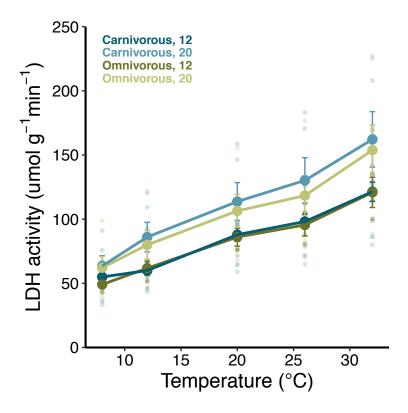
Represented are model formulas as input into R and AIC output results. df = degrees of freedom, AIC = Akaike Information Criterion  $\triangle$ AIC = AIC(model)—AIC(min AIC value), acute\_temp = acute temperature, fish\_id = individual fish.



**Fig. S1.** Figure illustrating repeatability of sprint performance across individuals. Each dot indicates a max sprint performance (cm s<sup>-1</sup>) calculated from an individual sprint trial. Colors indicate treatments with dark blue (carnivorous diet at 12°C), dark green (omnivorous diet at 12°C), light blue (carnivorous diet at 20°C), light green (omnivorous diet at 20°C).



**Fig. S2.** Performance in opaleye acclimated to 12°C or 20°C and fed either a carnivorous (blue) or omnivorous (green) diet. Presented are **A**) sprints measured as speed in cm s<sup>-1</sup> and **B**) sprints measured as speed in BL s<sup>-1</sup>, **C**) Growth rate (average fish mass (g) gained per week per tank) **D**) Lipid Peroxidation (LPO) in liver tissue measured as malondialdehyde concentration (MDA) in μmol gram<sup>-1</sup> of liver tissue. In panel A, B, D box plots represent interquartile ranges (boxes and whiskers), median values (solid lines) and outliers (> 1.5 beyond interquartile range) are plotted as data points outside the whiskers. In panel C, large circles and triangles indicate mean (± SEM) values for the carnivorous (*Artemia* sp.) and omnivorous diet treatments (*Artemia* sp. and *Ulva* sp.), respectively.



**Fig. S3.** Lactate dehydrogenase (LDH) activity in  $\mu$  mol per gram wet white muscle tissue weight in opaleye acclimated to 12°C (dark colors) or 20°C (light colors) and fed either a carnivorous (*Artemia* sp., represented as blues) or omnivorous diet (*Artemia* sp. and *Ulva* sp., represented as greens). Circles represent mean values and error bars indicate SEM. For each sample, LDH activity was measured at 5 different temperatures (8, 12, 20, 26, 32°C). Lactate dehydrogenase activity was higher at 20°C compared to 12°C but did not differ across diets. Lactate dehydrogenase activity also increased with acute temperature exposure. Acute temp: df = 4,  $\chi^2$  = 1061.711, p<0.001; acclimation temp: df = 1,  $\chi^2$  = 5.132, p = 0.023; diet: df = 1,  $\chi^2$  = 0.172, p = 0.679; acute temp × acclimation temp: df = 4,  $\chi^2$  = 22.526, p < 0.001.

## References

**Johnson, J.S., Clements K.D., and Raubenheimer, D.,** (2017). The Nutritional Basis of Seasonal Selective Feeding by a Marine Herbivorous Fish. *Mar. Biol.* **164,** 201.

Mann, R., and Gallager, S.M., (1985). Physiological and biochemical energetics of larvae of Teredo navalis L. and Bankia gouldi (Bartsch) (Bivalvia: Teredinidae). *J. Exp. Mar. Biol. Ecol.* **85**, 211-228.